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**Jiang et al.**

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(54) **IMMUNOENHANCER-LINKED  
OLIGOMERIC HIV ENVELOPE PEPTIDES**

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**C07K 14/005** (2006.01)

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**2319/40** (2013.01); **C12N 2740/16122**  
(2013.01); **C12N 2740/16134** (2013.01)

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See application file for complete search history.

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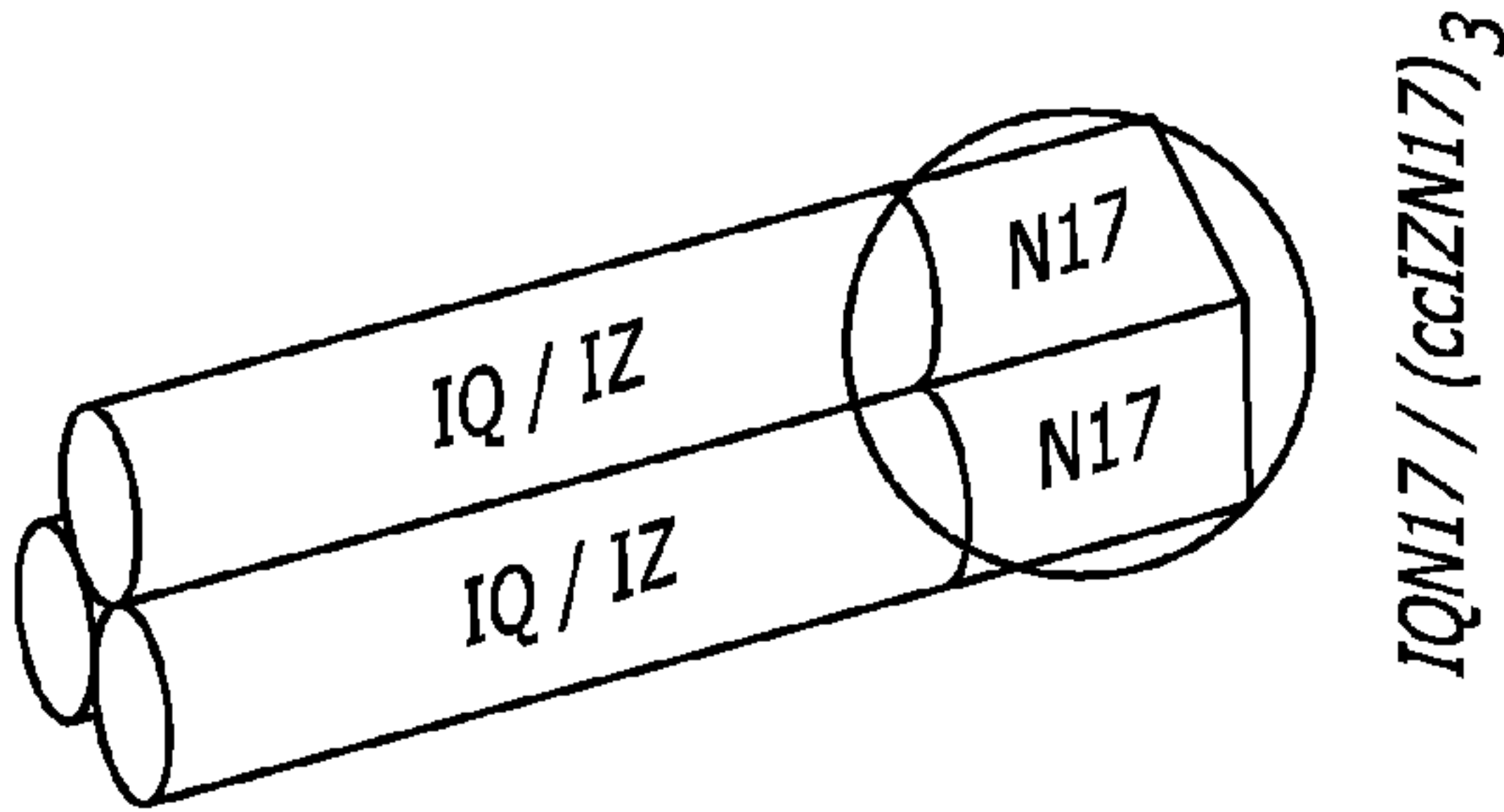
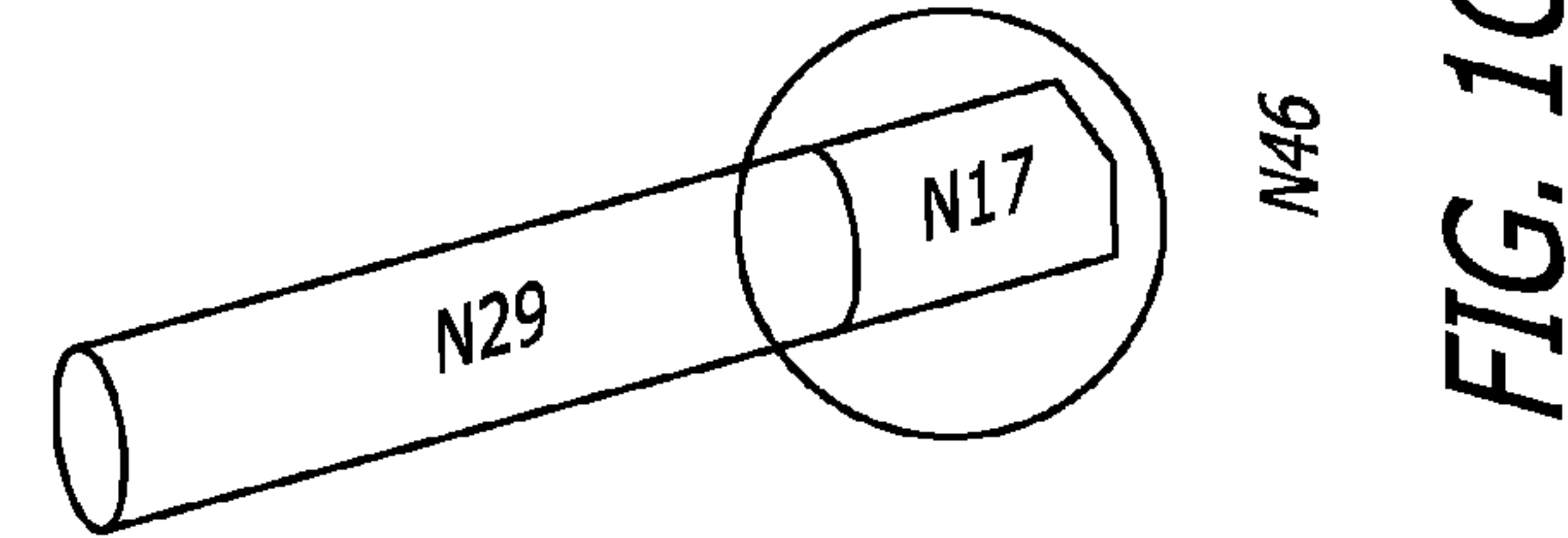
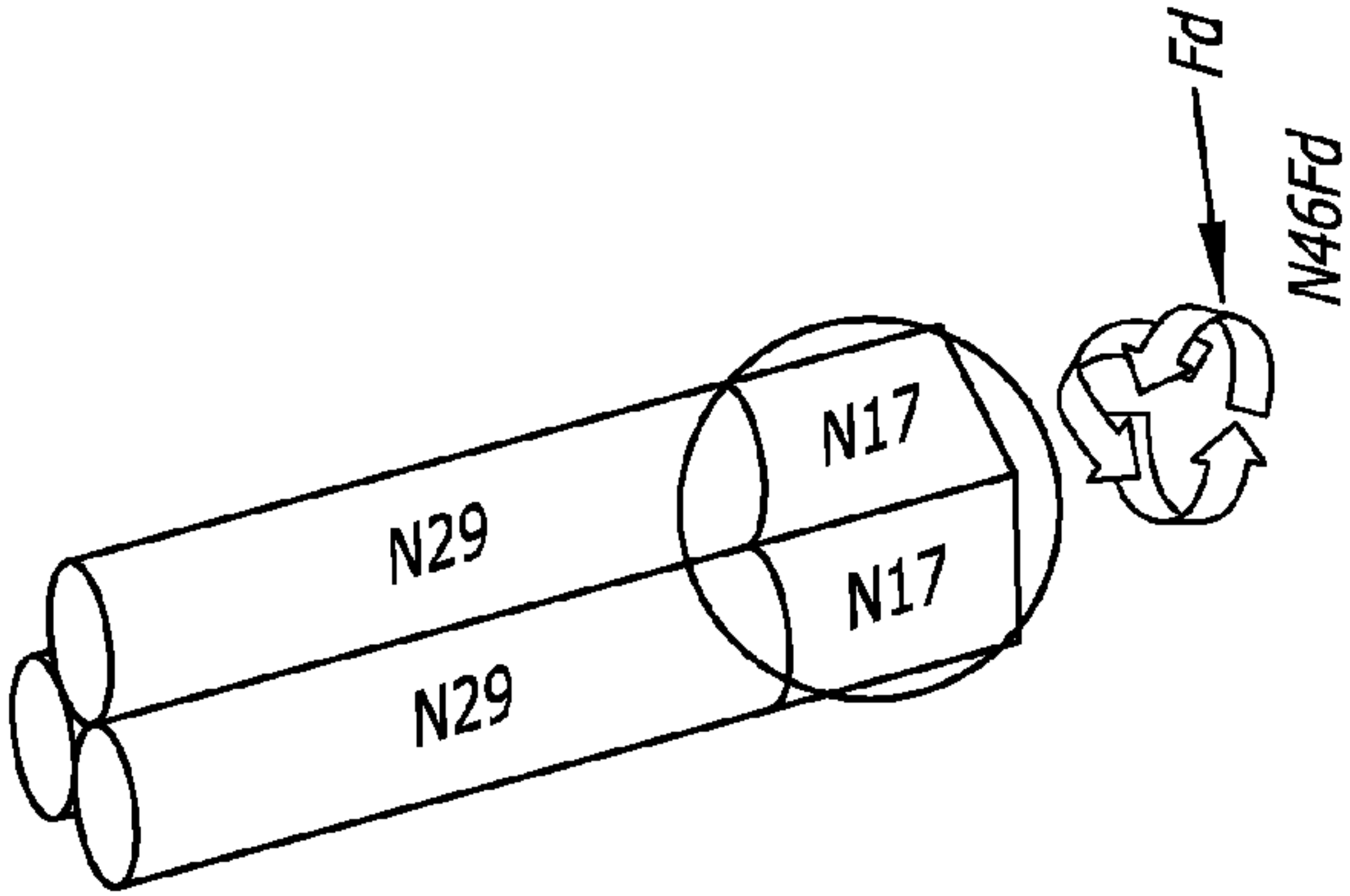
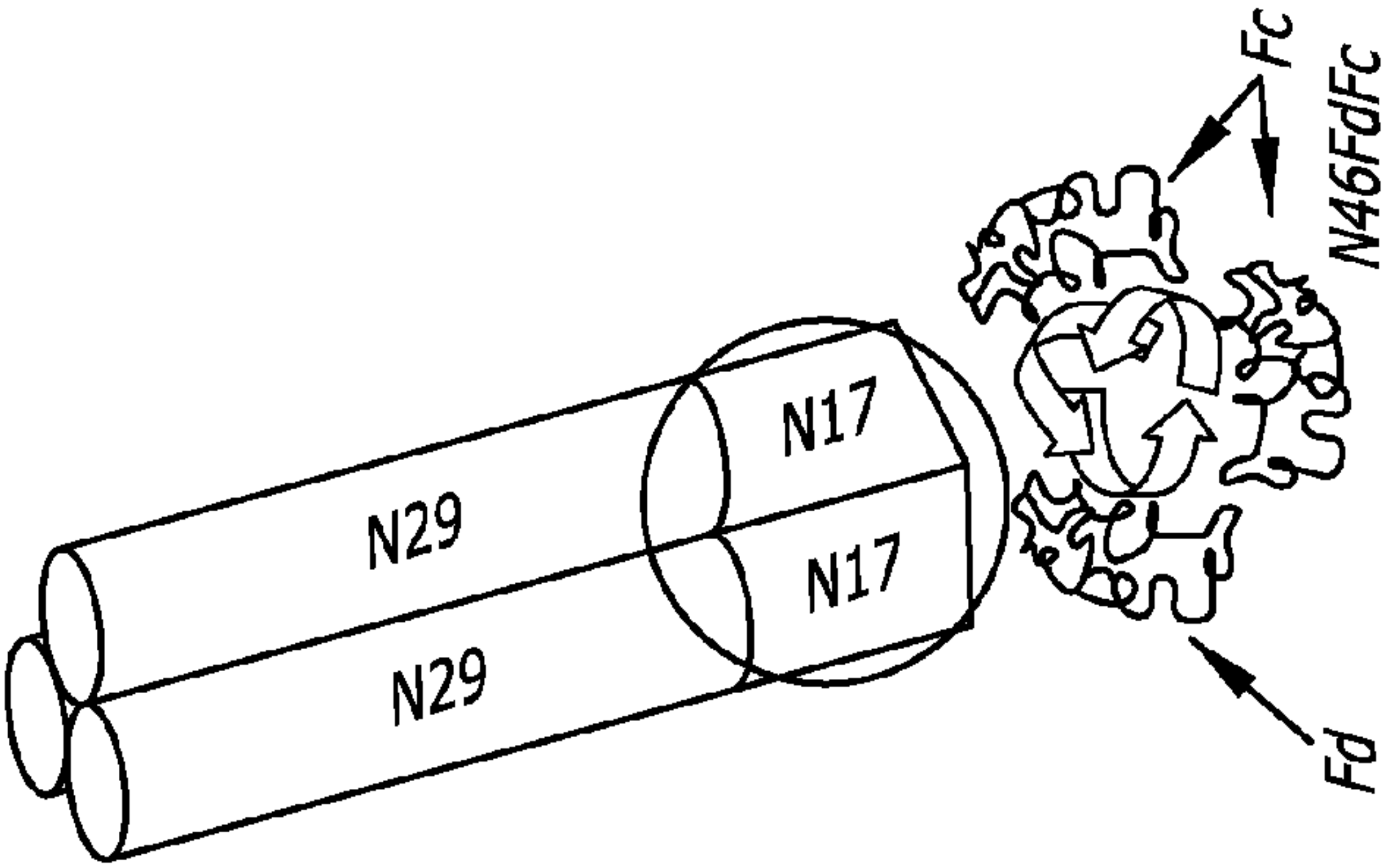
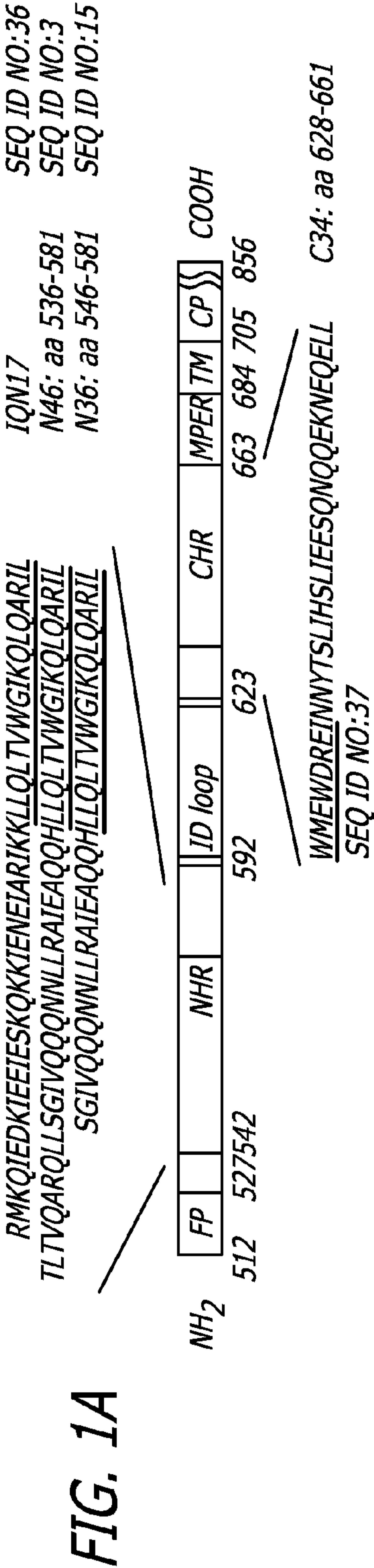
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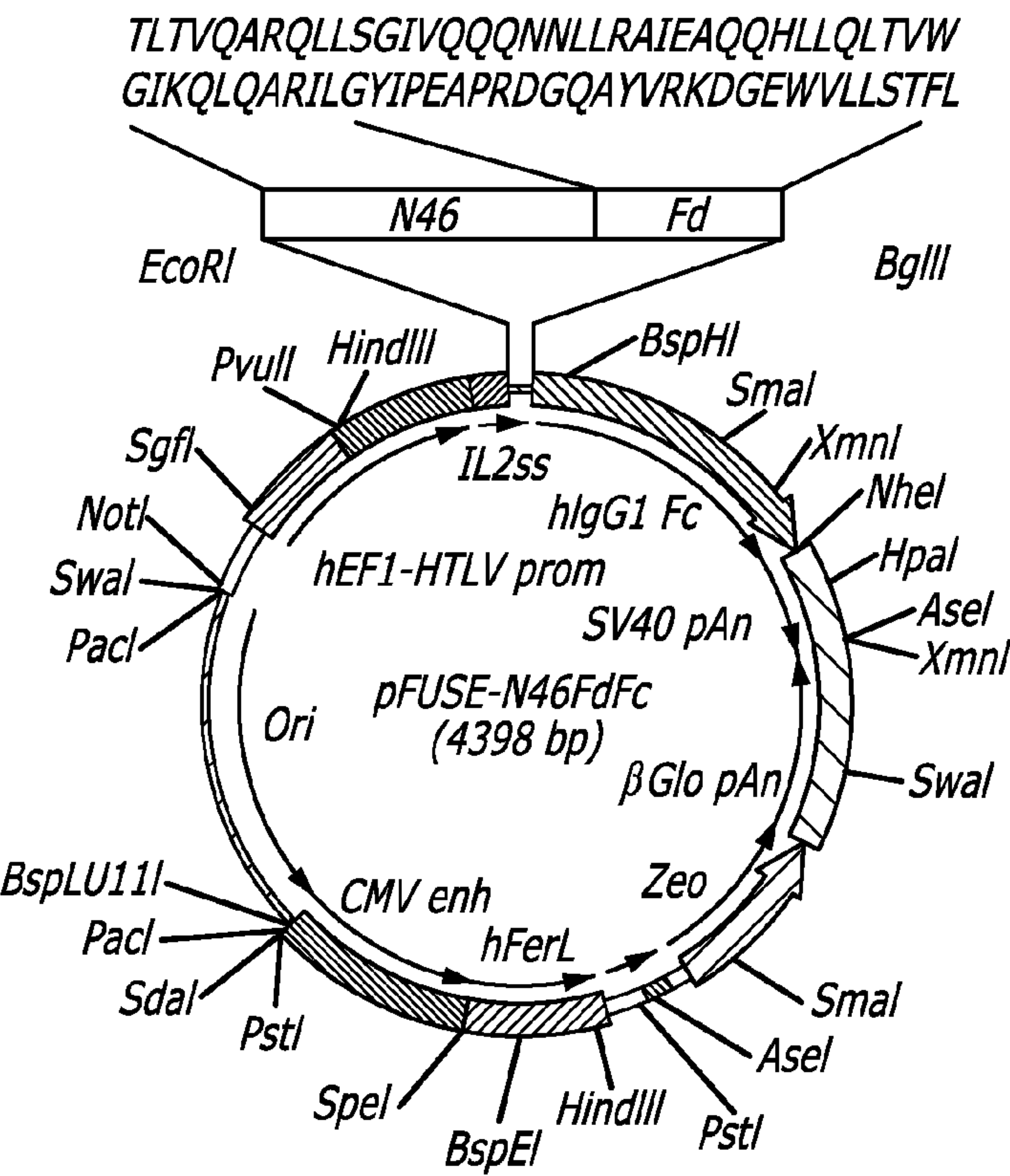
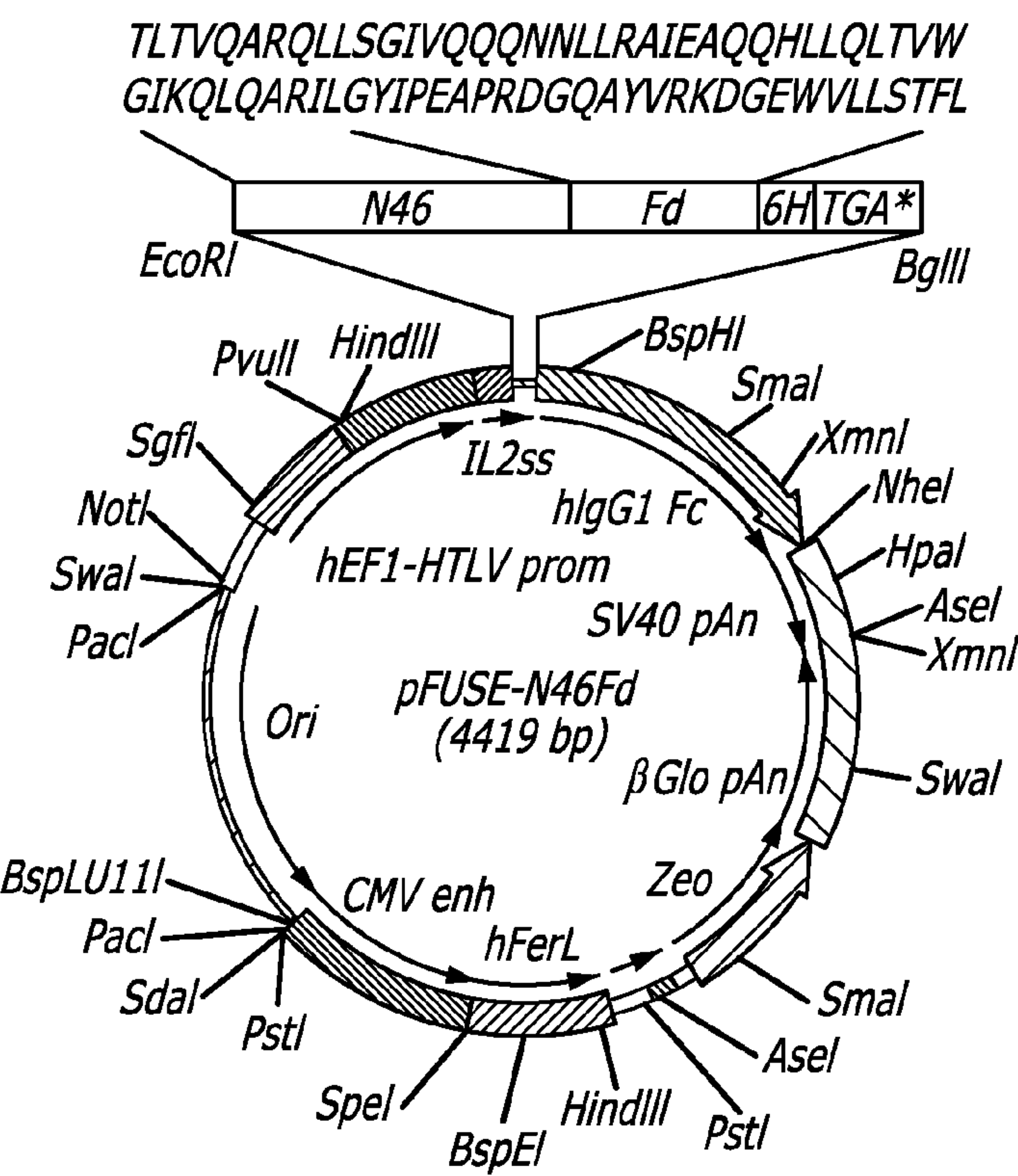
(57) **ABSTRACT**

Provided herein are immunogenic compositions comprising  
fusion proteins, the fusion proteins comprising lentivirus  
gp41 or a fragment thereof, a trimerization or oligomerization  
motif and an immunoenhancer that elicit potent and broad  
HIV neutralizing antibody responses in the immunized hosts.  
Also disclosed are methods of making and using the immu-  
nogenic compositions.

**15 Claims, 6 Drawing Sheets**







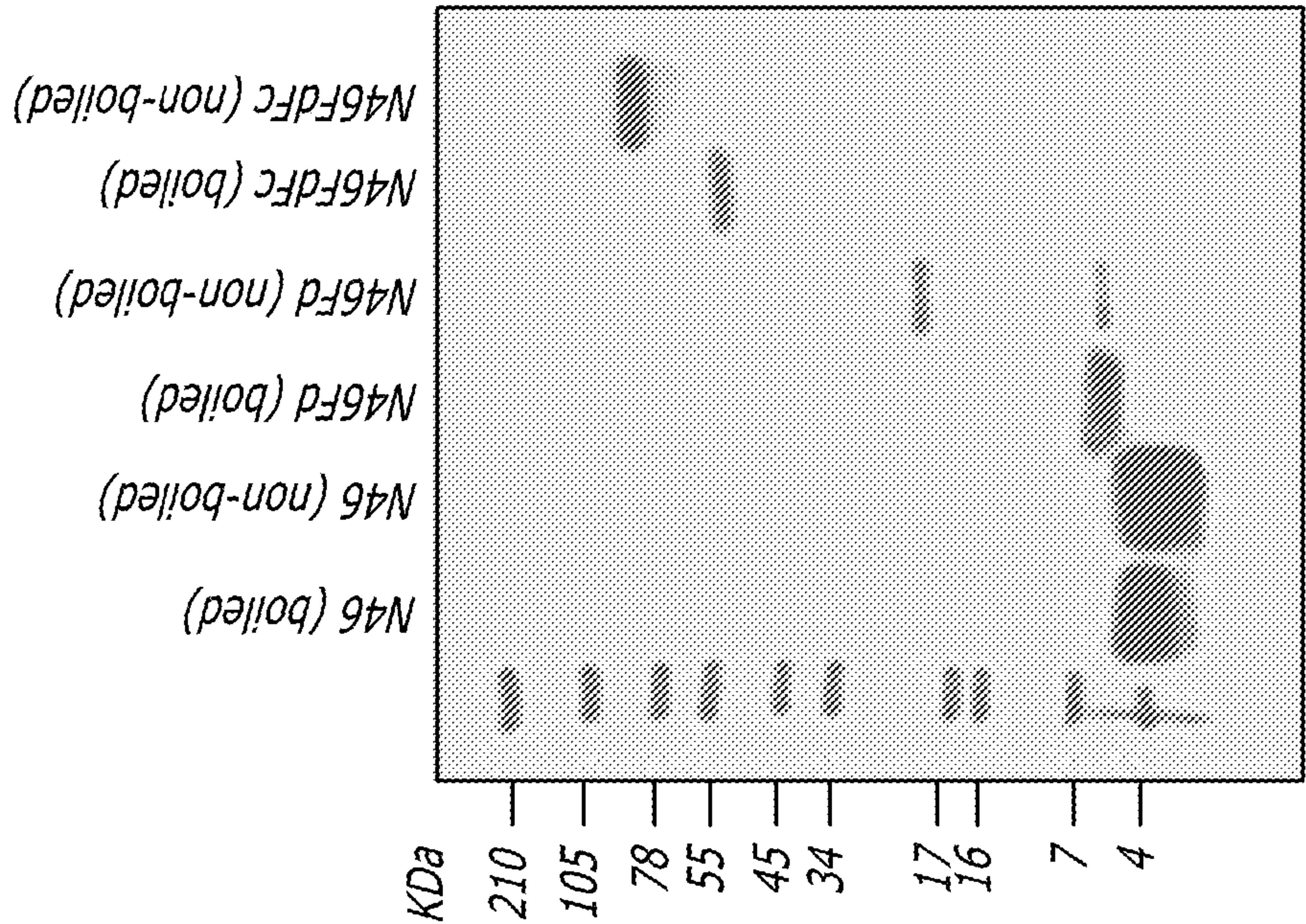


FIG. 3B

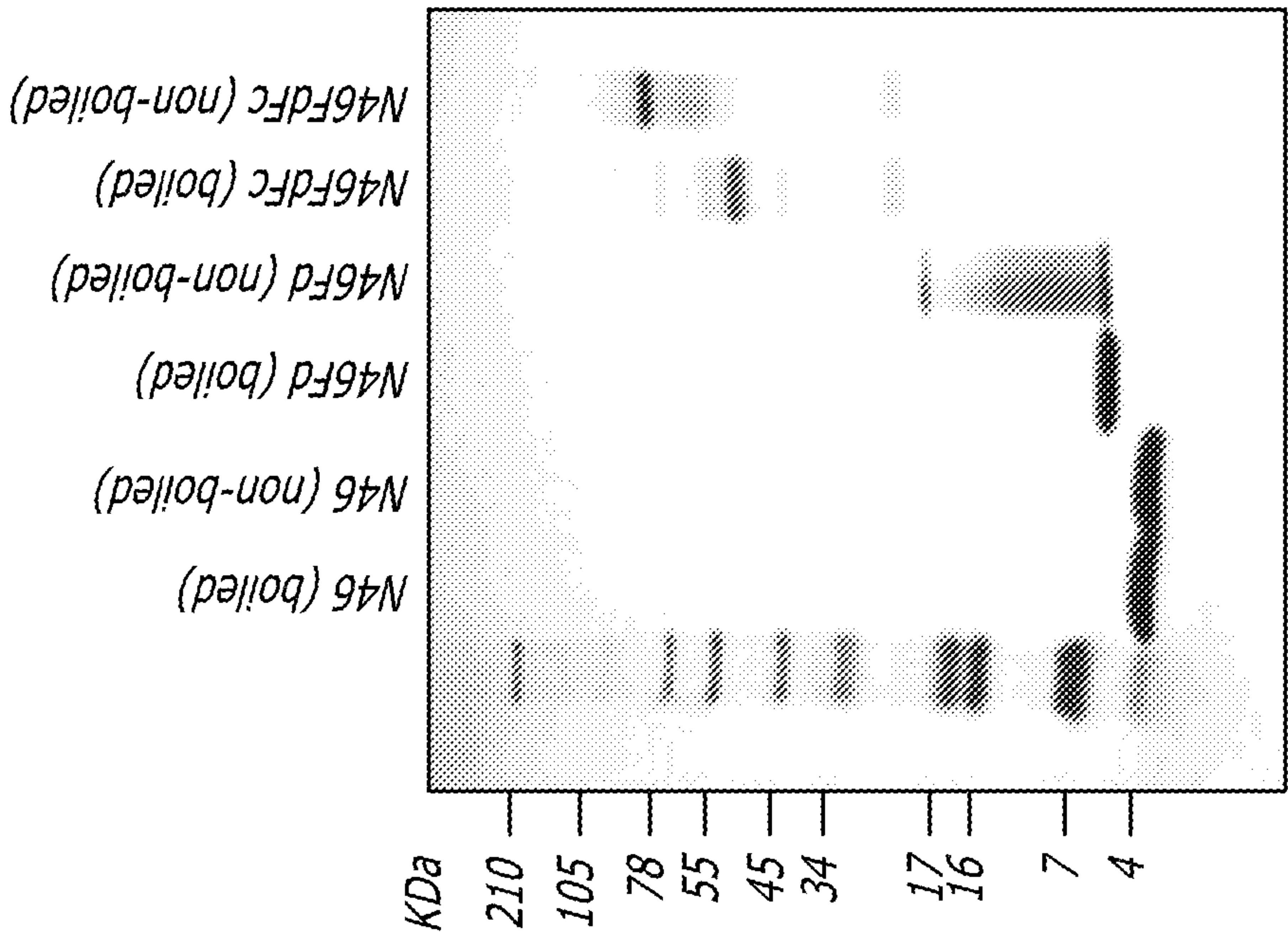


FIG. 3A



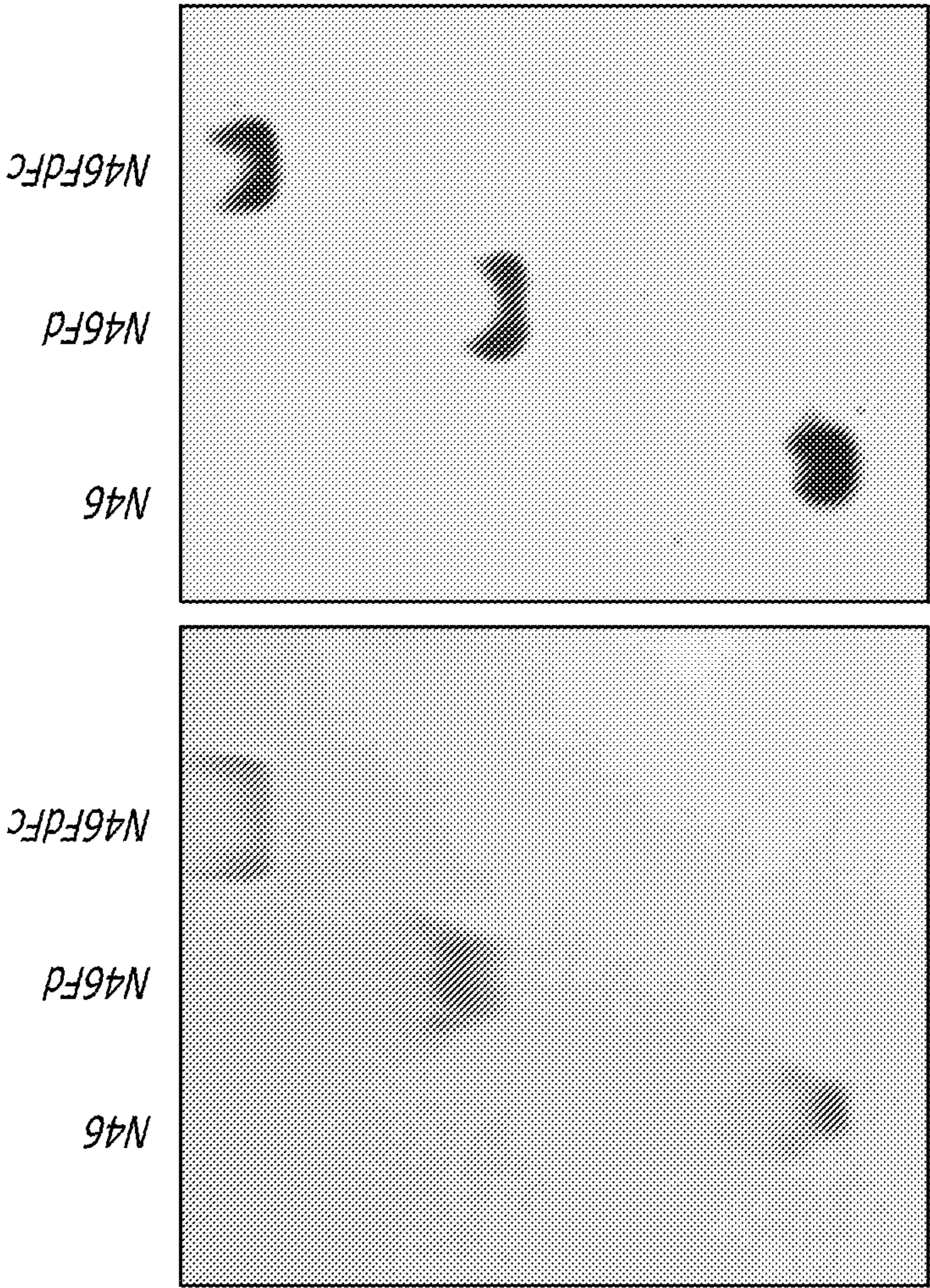
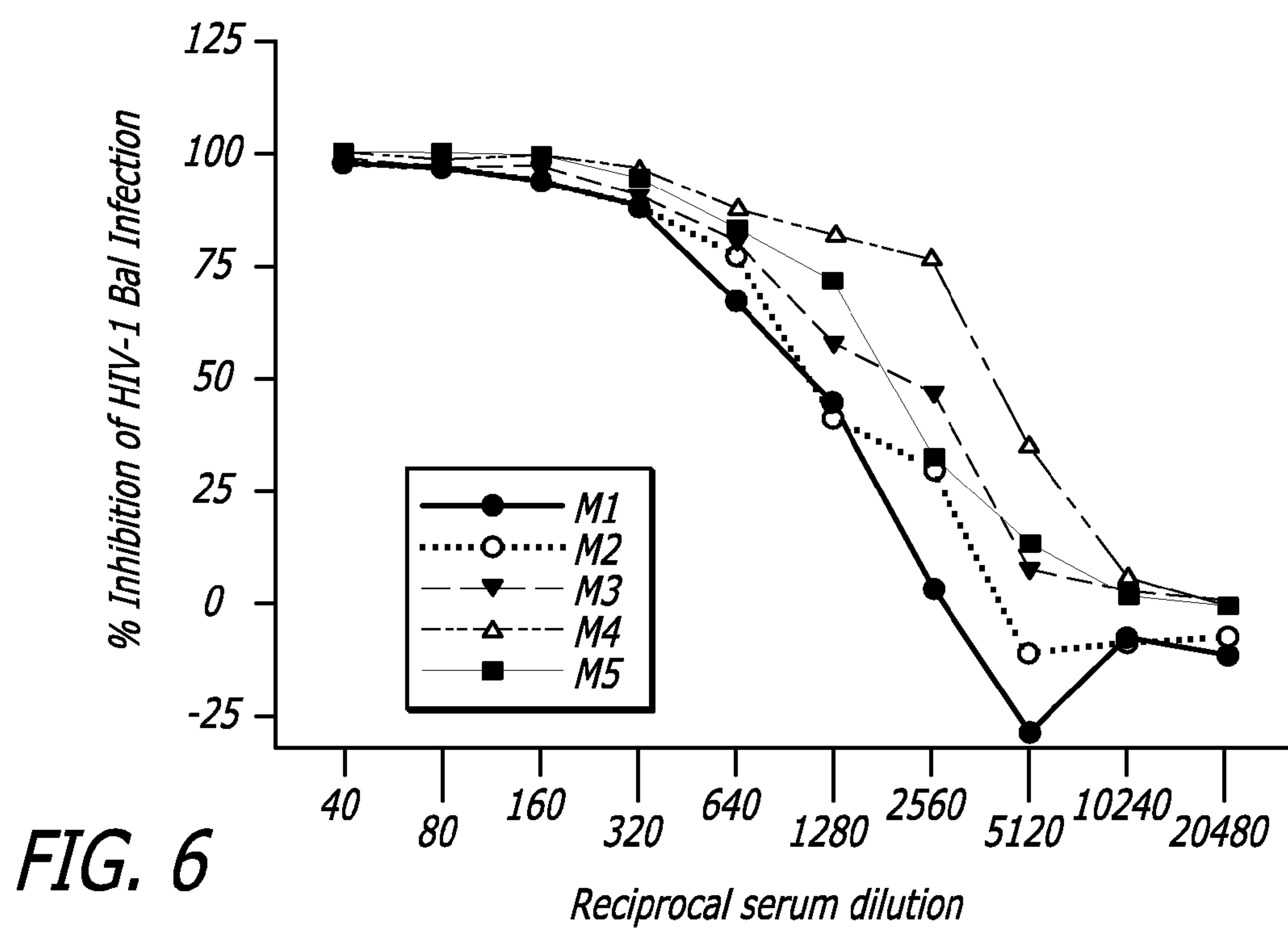
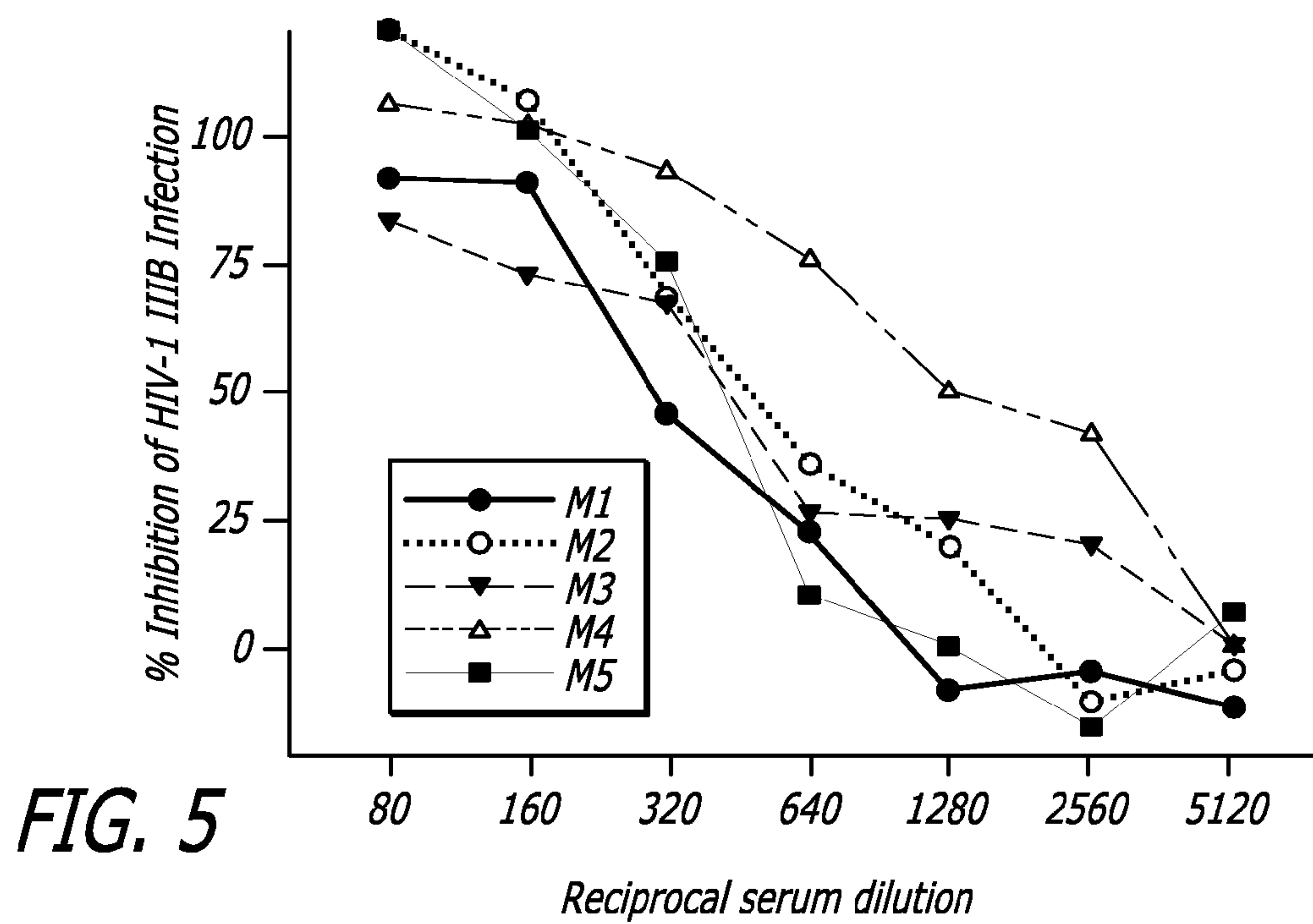


FIG. 4A

FIG. 4B



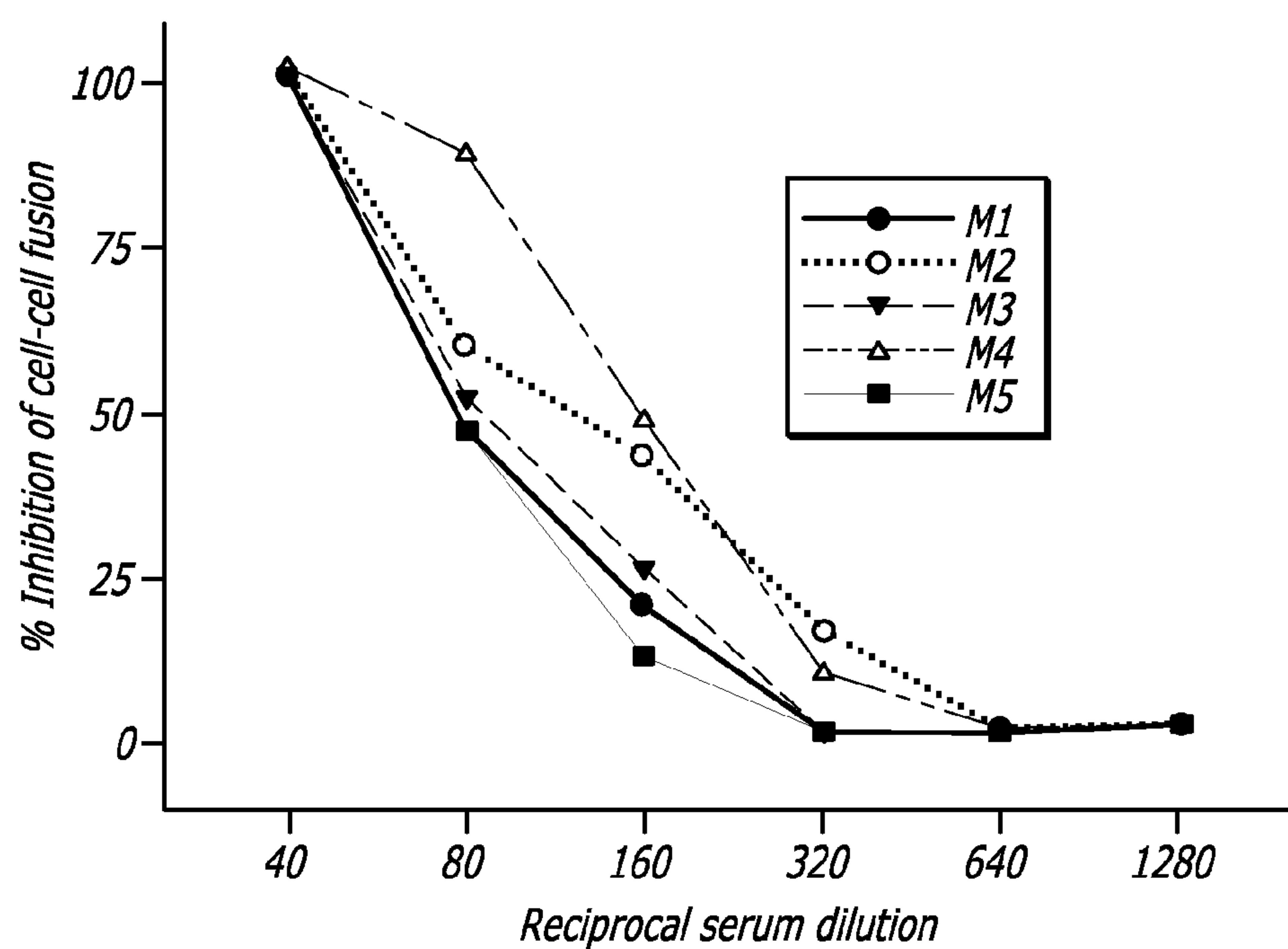


FIG. 7

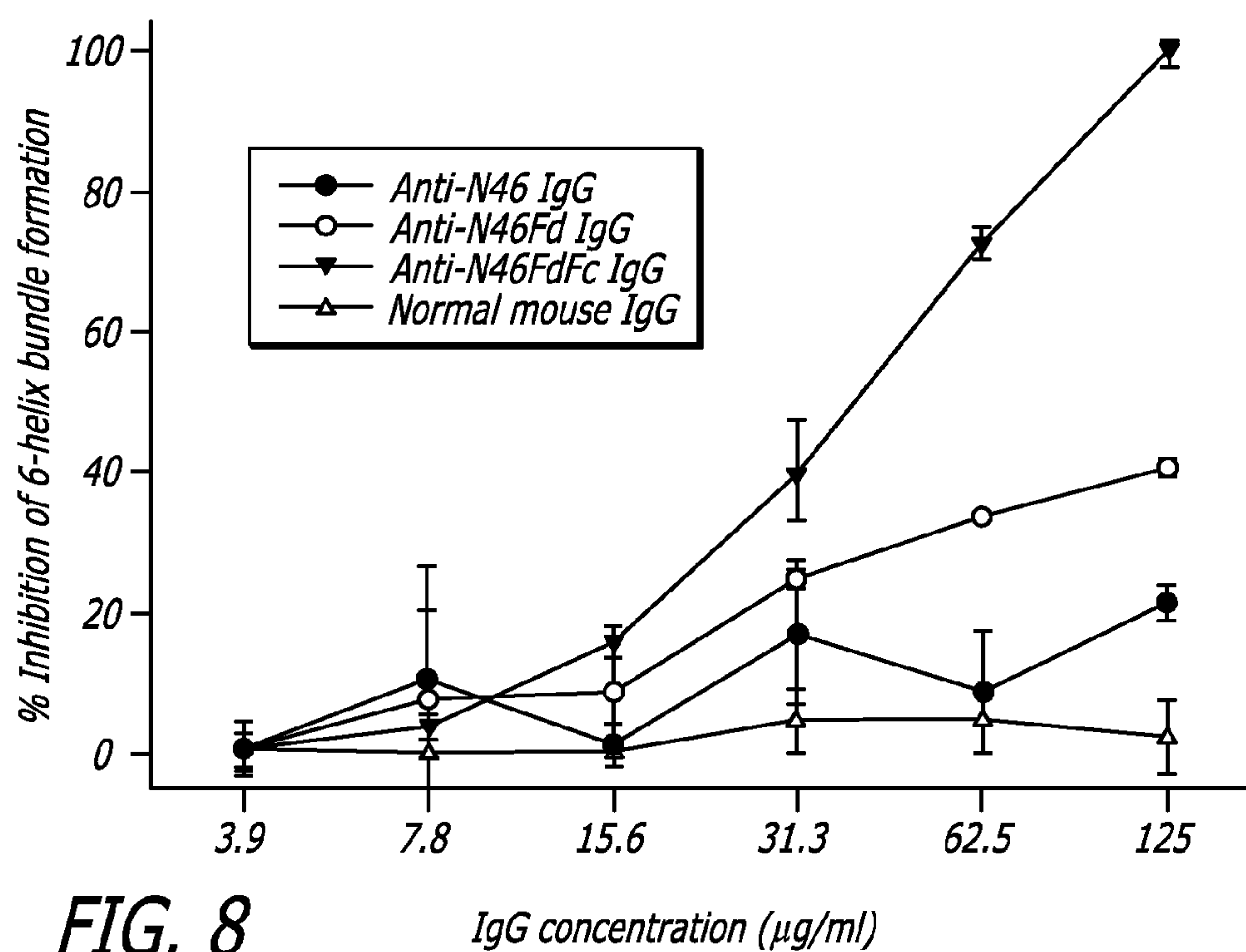


FIG. 8



# IMMUNOENHANCER-LINKED OLIGOMERIC HIV ENVELOPE PEPTIDES

## CROSS REFERENCE TO RELATED APPLICATION

The present application claims the benefit under 35 USC §119(e) to U.S. Provisional Patent Application 61/106,101 filed Oct. 16, 2008, the entire contents of which are incorporated by reference herein.

## FIELD OF THE INVENTION

The present disclosure relates to the field of immunogenic compositions for the prevention of HIV infection.

## BACKGROUND OF THE INVENTION

More than 60 million people worldwide have been infected by the human immunodeficiency virus (HIV) and nearly half have died of the resultant Acquired Immunodeficiency Syndrome (AIDS) since 1981. About 2.7 million new infections were reported in 2007. Therefore, development of an effective and safe HIV vaccine is urgently needed to contain the spread of HIV and AIDS. However, most previous efforts to achieve this goal have failed. Clinical trials of the first T-cell vaccine (Merck's MRKAd5 HIV-1 gag/pol/nef trivalent vaccine) were terminated recently because the data showed that the vaccine was unable to prevent HIV infection and could not lower virus levels in vaccinated volunteers who became infected. The first B-cell vaccine (VaxGen's AIDSVax, a bivalent gp120-based subunit vaccine) tested in clinical trials also failed to protect volunteers from HIV infection, possibly due to its inability to elicit broad neutralizing antibody responses because of great variability and high glycosylation of gp120. Since the HIV-1 envelope glycoprotein (Env) transmembrane subunit gp41 has relatively conserved sequence and less glycosylation sites than gp120, it may be a better target than gp120 for vaccine development. Indeed, two human monoclonal antibodies (mAbs) targeting gp41, 2F5 and 4E10, exhibit much broader neutralizing activity than those targeting gp120 (mAbs 2G12 and b12).

HIV-1 gp41 (SEQ ID NO. 2) plays an essential role in virus fusion with the target cell. HIV-1 gp41 consists of three essential functional regions: fusion peptide (FP), N-terminal heptad repeat (called NHR or HR1, which refer to the same sequence and are considered equivalent as used herein) and C-terminal heptad repeat (called CHR or HR2, which refer to the same sequence and are considered equivalent as used herein) (FIG. 1A). Both NHR and CHR contain a number of leucine zipper-like motifs which have tendency to form coiled coil structures. Peptides derived from the NHR and CHR regions are effective in inhibiting HIV-1 fusion with the target cells and one of the CHR-peptides, enfuvirtide was licensed by the United States Food and Drug Administration (US FDA) in 2003 as the first member of a new class of anti-HIV drugs-HIV fusion inhibitors.

HIV fusion with the host cell is initiated by binding of Env surface subunit 120 to the primary receptor CD4 and a co-receptor, CXCR4 or CCR5, resulting in a series of conformational changes in gp41, including insertion of FP into the target cell membrane and association of CHR-helices with the NHR-trimer, a prehairpin intermediate, to form a stable six-helix bundle (6-HB) core, which bring the viral envelope and target cell membrane into close proximity for fusion. X-ray crystallographic studies have shown that 6-HB consists of three molecules of a NHR-peptide that form the inner

trimeric coiled-coil and three copies of a CHR-peptide that pack obliquely in an anti-parallel configuration into the highly conserved hydrophobic grooves on the surface of the internal NHR-trimer. Each groove has a deep hydrophobic pocket (FIG. 1B, circled at bottom), which plays an important role in viral fusion and maintaining the stability of the 6-HB. Accordingly, it is proposed that a CHR-peptide (e.g., enfuvirtide or C34) inhibits HIV-1 fusion by binding to the viral gp41 NHR-trimer at the fusion-intermediate state to block the formation of fusion-active core of gp41. Therefore, the gp41 NHR-trimer is a crucial target for HIV therapeutics, and may also serve as an important target for HIV vaccines.

However, the gp41 NHR-peptides cannot form stable and soluble trimers in vitro spontaneously because these peptides have tendency to aggregate in physiological solutions. To study the structure, function and immunogenicity of the gp41 prehairpin intermediate, several soluble and stable NHR-trimer mimetics have been created, including IQN17 (FIG. 1C), in which a 17-mer NHR-peptide (aa 565-581) involving in formation of the gp41 hydrophobic pocket is linked with GCN4-pI<sub>Q</sub>I (IQ) motif, a soluble trimeric coiled coil, and similar mimetics with higher stability, including IZN17, IZN36 and (ccIZN17)<sub>3</sub>, in which IQ is replaced with more stable trimerization motif, IZ. Other examples of NHR-trimer mimetics include NCCG-gp41, N35<sub>NCCG</sub>-N13, and 5-Helix in which one CHR peptide of the 6-HB is missing so as to expose the groove of the NHR-trimer. Although all these NHR-trimer mimetics properly present the hydrophobic groove and pocket and are effective in interacting with viral gp41 CHR to inhibit HIV-1 fusion, none of them could induce detectable neutralizing antibody responses in immunized animals. It is believed that the accessibility of the prehairpin intermediate of gp41 to antibody molecules (e.g., IgG) is limited because antisera directed against NHR-peptides exhibited no neutralizing activity at 37° C., but were effective under suboptimal temperature (31.5° C.) to prolong fusion intermediates. But interestingly, IgG1 m44, a human mAb directed against gp41 is much more potent than Fab m44 in neutralizing infection by primary HIV-1 isolates. Another human mAb, D5, that specifically binds to the pocket of NHR-trimer is highly potent to neutralize HIV-1 infection. Although rabbit antisera induced against N35<sub>NCCG</sub>-N13 showed no neutralizing activity, purified IgG from the antisera (about 5-10% of total IgGs) with high-binding affinity to the NHR-trimer could significantly inhibit HIV-1 Env-mediated cell fusion. These data suggest that the gp41 NHR-trimer in the prehairpin fusion intermediate state is accessible to antibodies, which is not restricted by either antibody size or the presence of a kinetic barrier, but may be limited by the affinity of antibodies to bind with the NHR-trimer. Therefore, it is essential to design an immunogen with proper conformation and increased immunogenicity that can induce antibodies with high-binding affinity to the gp41 prehairpin intermediate.

## SUMMARY OF THE INVENTION

The N-terminal heptad repeat (NHR or HR1) alpha-helical trimer of HIV-1 envelope protein (Env) transmembrane subunit gp41 plays a crucial role in virus fusion with the target cell and represents an important target for therapeutics (e.g., enfuvirtide) and vaccines. Disclosed herein is a subunit oligomeric immunogenic composition comprising at least a portion of the HIV-1 gp41, a trimerization motif and an immunoenhancer.

In one embodiment presented herein, an immunogenic composition for induction of an immune response against a



lentivirus is provided, said immunogenic composition comprising a fusion protein comprising a gp41 sequence; a trimerization or oligomerization motif; and an immunoenhancer.

In another embodiment, the gp41 sequence is from a lentivirus selected from the group consisting of HIV-1, HIV-2 or SIV. In another embodiment, the gp41 sequence is selected from a portion of the gp41 molecule selected from the group consisting of the N-terminal heptad repeat (NHR or HR1) region, the C-terminal heptad repeat (CHR or HR2) region, the fusion peptide (FP) region and the membrane proximal external region (MPER) of gp41.

In another embodiment, the trimerization or oligomerization motif is selected from the group consisting of foldon, IQ, and IZ.

In another embodiment, the immunoenhancer is selected from the group consisting of the Fc domain of immunoglobulin G, complement component C3d, and Onchocerca volvulus activation associated protein-1 (Ov-ASP-1). In another embodiment, the Fc domain of immunoglobulin G or complement component C3d is from a mammal selected from the group consisting of mouse, rabbit, pig, non-human primate and human.

In yet another embodiment, the fusion protein comprises the gp41 sequence, the trimerization or oligomerization sequence and the immunoenhancer in that order. In yet another embodiment, the fusion protein further comprises a His tag or a GST sequence.

In still another embodiment, the immunogenic composition further comprises an adjuvant.

In one embodiment disclosed herein, a method is provided for inducing an immune response to HIV comprising the steps of administering the immunogenic composition of claim 1 to a mammal in need thereof; and inducing an immune response in said mammal to said HIV.

In another embodiment, the immunogenic composition is administered by a route selected from the group consisting of subcutaneous, intramuscular, intraperitoneal, and mucous immunization. In yet another embodiment, the immune response results in the production of neutralizing antibodies in said mammal.

In one embodiment presented herein, an immunogenic composition is provided for induction of an immune response against a lentivirus comprising a fusion protein, the fusion protein comprising a N46 sequence of human immunodeficiency virus gp41; a foldon trimerization motif; and a human immunoglobulin G Fc sequence.

#### BRIEF DESCRIPTION OF THE DRAWINGS

FIG. 1 depicts the structures of the HIV-1 gp41 and NHR-trimers as well as the NHR- and CHR-peptides.

FIG. 2 depicts the map of the expression vectors pFUSE-N46Fd (FIG. 2A) and pFUSE-N46FdFc (FIG. 2B) encoding N46Fd and N46FdFc, respectively.

FIG. 3 depicts the SDS-PAGE analysis of N46, N46Fd, and N46FdFc.

FIG. 4 depicts the analysis of N46, N46Fd, and N46FdFc by using Acid-N-PAGE (FIG. 4A) and Western blot (FIG. 4B) using anti-N46 antibodies.

FIG. 5 depicts neutralization of HIV-1 IIIB (X4) Bal (R5) infection mediated by antisera against N46FdFc.

FIG. 6 depicts neutralization of HIV-1 Bal (R5) infection mediated by antisera against N46FdFc.

FIG. 7 depicts the inhibition of HIV-1-mediated cell-cell fusion mediated by antisera against N46FdFc.

FIG. 8 depicts the inhibition of the gp41 six-helix bundle formation by IgG purified from antisera against N46FdFc.

#### DEFINITION OF TERMS

To facilitate an understanding of the following Detailed Description, Examples and appended claims it may be useful to refer to the following definitions. These definitions are non-limiting in nature and are supplied merely as a convenience to the reader.

Gene: A "gene" as used herein refers to at least a portion of a genetic construct having a promoter and/or other regulatory sequences required for, or that modify the expression of, the genetic construct.

Host: As used herein "host" refers to the recipient of the present immunogenic compositions. Exemplary hosts are mammals including, but not limited to, primates, rodents, cows, horses, dogs, cats, sheep, goats, pigs and elephants. In one embodiment of the present invention the host is a human. For the purposes of this disclosure host is synonymous with "vaccinee."

Immunogen: As used herein the term "immunogen" shall mean any substrate that elicits an immune response in a host. Immunogens of the present disclosure include, but are not limited to human immunodeficiency proteins.

Immunogenic Composition: An "immunogenic composition" as used herein comprises an expressed protein or a recombinant vector, with or without an adjuvant, that expresses and/or secretes an immunogen in vivo and wherein the immunogen elicits an immune response in the host. The immunogenic compositions disclosed herein may or may not be immunoprotective or therapeutic. When the immunogenic compositions may prevent, ameliorate, palliate or eliminate disease from the host then the immunogenic composition may optionally be referred to as a vaccine. However, the term immunogenic composition is not intended to be limited to vaccines.

The term "HIV fusion" refers to a critical step of virus life cycle necessary for a virion or HIV-infected fusing with a target vesicle or cell.

"ND50" refer to the antibody concentration or antiserum titer that results in a 50% neutralization, respectively, of virus infection.

"IC50 or EC50" refer to the antibody concentration or antiserum titer that results in a 50% reduction or inhibition, respectively, in virus infection or virus-mediated cell-cell fusion.

#### DETAILED DESCRIPTION OF THE INVENTION

The N-terminal heptad repeat (NHR or HR1) alpha-helical trimer of HIV-1 envelope protein (Env) transmembrane subunit gp41 plays a crucial role in virus fusion with the target cell and represents an important target for therapeutics (e.g., enfuvirtide) and vaccines. Disclosed herein is a subunit immunogenic composition comprising a fusion protein comprising at least a portion of the HIV-1 gp41, a trimerization motif and an immunoenhancer. The immunogenic compositions disclosed herein may be administered in any formulations and with any adjuvant.

In one embodiment the composition comprises a fusion protein comprising an NHR-peptide N46 (SEQ ID NO. 3) derived from the HIV-1 HXB2 (subtype B) gp41 (SEQ ID NO. 2), a trimerization motif foldon (Fd) (SEQ ID NO. 18), and the human immunoglobulin G Fc domain (hFc) (SEQ ID NO. 21) as an immunoenhancer, designated N46FdFc (SEQ ID NO. 25). The N46 peptide and the recombinant protein



without Fc, N46Fd (SEQ ID NO. 24) were used as controls. Unlike the N46 peptide and N46Fd, N46FdhFc elicited potent neutralizing antibody responses in the immunized mice against infection by laboratory-adapted and primary HIV-1 strains.

In another embodiment, a fusion protein is provided comprising a gp41 NHR sequence derived from one of the following sequences: HIV-1 94UG103, subtype A (SEQ ID NO. 4); 92US657, subtype B (SEQ ID NO. 5); HIV-1 93IN101, subtype C (SEQ ID NO. 6); HIV-1 92UG001, subtype D (SEQ ID NO. 7); HIV-1 92THA009, subtype NE (SEQ ID NO. 8); HIV-1 93BR020, subtype F (SEQ ID NO. 9); HIV-1 RU570, subtype G (SEQ ID NO. 10); HIV-1 BCF02, group O (SEQ ID NO. 11); or from HIV-2 CBL20 (SEQ ID NO. 12), a trimerization motif and an immunoenhancer.

The fusion protein described herein comprises at least one gp41 NHR sequence derived from the entire NHR(HR1) region or from part of the NHR(HR1) region, e.g., N17 (SEQ ID NO. 13), N36 (SEQ ID NO. 14), N28 (SEQ ID NO. 15), N36 (SEQ ID NO. 16), N51 (SEQ ID NO. 17), or N63 (SEQ ID NO. 18) derived from HIV-1 HXB2 (subtype B).

The fusion protein described herein comprises at least one gp41 NHR(HR1) sequence from a lentivirus. Exemplary lentiviruses include, but are not limited to, any strain of HIV-1, HIV-2 or simian immunodeficiency virus (SIV).

In one embodiment, the fusion protein described herein comprises an entire gp41 sequence from HIV-1 HXB2 (SEQ ID NO. 2). In another embodiment, the fusion protein comprises an entire gp41 sequence from HIV-1, HIV-2 or SIV.

Moreover, fusion proteins disclosed herein comprise a trimerization or oligomerization motif that includes, but is not limited to, foldon (SEQ ID NO. 18), IQ (the IQ calmodulin-binding motif, SEQ ID NO. 19), and IZ (isoleucine zipper motif, SEQ ID NO. 20). Foldon is a trimerization or oligomerization motif from the T4 bacteriophage fibrin. Additionally, the gp41 component and the immunoenhancer can be physically linked by 2,2-bipyridine-5-carboxylic acid or intramolecular disulfide bonds.

Additionally, the fusion proteins include an immunoenhancer that includes, but is not limited to, Fc fragment of human IgG (SEQ ID NO. 20), mouse IgG Fc (SEQ ID NO. 21), rabbit IgG Fc (SEQ ID NO. 22), C3d of human complement, or an immunomodulator, such as a cytokine.

In one embodiment, the immunoenhancer is immunoglobulin Fc fragment. The immunoglobulin molecule consists of two light chains and two heavy chains held together by disulfide bonds such that the chains form a Y shape. The base of the Y (carboxy terminus of the heavy chain) plays a role in modulating immune cell activity. This region is called the Fc (fragment, crystallizable) region, and is composed of two heavy chains that contribute two or three constant domains depending on the class of the antibody. By binding to specific proteins, the Fc region ensures that each antibody generates an appropriate immune response for a given antigen. The Fc region also binds to various cell receptors, such as Fc receptors, and other immune molecules, such as complement proteins. By doing this, it mediates different physiological effects including opsonization, cell lysis, and degranulation of mast cells, basophils and eosinophils.

TABLE 1

Oliomeric immunogenic composition fusion protein segment sequences	
Sequence Identifier	Amino Acid Sequence
SEQ ID NO. 1 gp160 of HIV-1 HXB2, subtype B	MRVKEKYQHLWRWGWRWGTMLLGMLMICSATEKLWVTVYYGVVW KEATTTLFCASDAKAYDTEVHNVWATHACVPTDPNPQEVVLNVNTE FNMWKNDMVEQMHEDIISLWDQSLKPCVKLTPLCVSLKCTDLKNDTN TNSSSGRMIMEKGEIKNCSFNIISTIRGKVQKEYAFFYKLDIIPIDNDTTS YKLTSCNTSVITQACPKVSFEPIPIHYCAPAGFAILKCNNKTFNGTGPCT NVSTVQCTHGIRPVVSTQLLLNGLSLAESEEVVIRSVMFTDNAKTIIIVQLNT SVEINCTRPNNNTRKRIRIQRGPGRAFVTIGKIGNMRQAHNCNISRAKWN NTLKQIASKLREQFGNNKTIIFKQSSGGDPEIVTHSFNCGGEFFYCNST QLFNSTWFNSTWSTEGSNNTGSDTITLPCRIKQIINMWQKVGKAMYA PPISGQIRCSSNITGLLLTRDGGNSNNESEIFRPGGGDMRDNRSELY KYKVVKIEPLGVAPTAKRRVVQREKRAVGIGALFLGFLGAAGSTMGA ASMTLTVQARQLLSGIVQQQNNLLRAIEAQQHLLQLTVWGIKQLQARIL AVERYLKDQQLGIWGC SGKLICTTAVPWNASWSNKSLEQIWNHTTW MEWDREINNYTSLIHSLIEESQNQQEKNQEELLELDKWASLWNWFNIT NWLWYIKLFIWVGGVLVGLRIVFAVLSIVNRVRQGYSPLSFQTHLPTPRG PDRPEGIEEEGGERDRDRSIRLVNGSLALIWDRLSLCLFSYHRLRDL LIVTRIVELLGRRGWEALKYWWNLLQYWSQELKNSAVSLLNATAIAVA EGTDRVIEVVGACRAIRHIPRRIRQGLERILL
SEQ ID NO. 2 gp41 of HIV-1 HXB2, subtype B	AVGIGALFLGFLGAAGSTMGAASMTLTVQARQLLSGIVQQQNNLLRAIE AQQHLLQLTVWGIKQLQARILAVERYLKDQQLGIWGC SGKLICTTAVP WNASWSNKSLEQIWNHTTWMEWDREINNYTSLIHSLIEESQNQQEKN EQELLELDKWASLWNWFNITNWLWYIKLFIWVGGVLVGLRIVFAVLSIVN RVRQGYSPLSFQTHLPTPRGPDRPEGIEEEGGERDRDRSIRLVNGSLA LIWDDLRLSLCLFSYHRLRDL LIVTRIVELLGRRGWEALKYWWNLLQY WSQELKNSAVSLLNATAIAVAEGTDRVIEVVGACRAIRHIPRRIRQGL ERILL
SEQ ID NO. 3 N46 from gp41 of HIV-1 HXB2, subtype B	TLTVQARQLLSGIVQQQNNLLRAIEAQQHLLQLTVWGIKQLQARI
SEQ ID NO. 4 N46 from gp41 of HIV-1 94UG103, subtype A	TLTVQARQLLSGIVQQQSNLLRAIEAQQHLLKLTWVGIKQLQARVL



TABLE 1-continued

Oliomeric immunogenic composition fusion protein segment sequences	
Sequence Identifier	Amino Acid Sequence
SEQ ID NO. 5 N46 from gp41 of HIV-1 92US657, subtype B	TLTVQARQLLSGIVQQQSNLLRAIEAQQHLLQLTVWGIKQLQARVL
SEQ ID NO. 6 N46 from gp41 of HIV-1 93IN101, subtype C	TLTAQARQLLSGIVQQQSNLLRAIEAQQHLLQLTVWGIKQLQTRVL
SEQ ID NO. 7 N46 from gp41 of HIV-1 92UG001, subtype D	TLTVQARQLLSGIVQHQNLLMAIEAQQHLLQLTVWGIKQLQARIL
SEQ ID NO. 8 N46 from gp41 of HIV-1 92THA009, subtype A/E	TLTVQARQLLGIVQQQSNLLRAIEAQQHLLQLTVWGIKQLQARVL
SEQ ID NO. 9 N46 from gp41 of HIV-1 93BR020, subtype F	TLTVQARQLLSGIVQQQSNLLRAIEAQQHLLQLTVWGIKQLQARVL
SEQ ID NO. 10 N46 from gp41 of HIV-1 RU570, subtype G	TLTVQVKKLLFGIVQQQSNLLRAIEAQQHLLQLTVWGIKQLQARVL
SEQ ID NO. 11 N46 from gp41 of HIV-1 BCF02, group O	ALTVRTHTLIKGIVQQQDNLLRAIQQQQLRLSVWGIRQLRARLL
SEQ ID NO. 12 N46 from gp41 of HIV-2 CBL20	TLSAQSRTLLAGIVQQQQQLLDVVKRQQEMLRLTVWGTKNLQARVT
SEQ ID NO. 13 N17 from gp41 of HIV-1 HXB2	LLQLTVWGIKQLQARIL
SEQ ID NO. 14 N28 from gp41 of HIV-1 HXB2	IEAQQHLLQLTVWGIKQLQARILAVERY
SEQ ID NO. 15 N36 from gp41 of HIV-1 HXB2	SGIVQQQNNLLRAIEAQQHLLQLTVWGIKQLQARIL
SEQ ID NO. 16 N51 from gp41 of HIV-1 HXB2	QARQLLSGIVQQQNNLLRAIEAQQHLLQLTVWGIKQLQARILAVERYLK QQ
SEQ ID NO. 17 N63 from gp41 of HIV-1 HXB2	STMGAASMTLTVQARQLLSGIVQQQNNLLRAIEAQQHLLQLTVWGIKQ LQARILAVERYLKDQ
SEQ ID NO. 18 Foldon	GYIPEAPRDGQAYVRKDGEWLLSTFL
SEQ ID NO. 19 IQ	RMKQIEDKIEEIESKQKKIENEIARIKK
SEQ ID NO. 20 IZ	IKKEIEAIKKEQEAIKKKIEAIEKEI
SEQ ID NO. 21 human IgG Fc	RSDKTHTCPPCPAPELLGGPSVFLFPPKPKDTLMISRTPEVTCVVVDV SHEDPEVKFNWYVDGVEVHNAKTKPREEQYNSTYRVVSVLTVLHQD WLNQKEYKCKVSNKALPAPIEKTISKAKGQPREPQVYTLPPSREEMTK NQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTPPVLDSDGSFFLYS KLTVDKSRWQQGNVFCFSVMHEGLHNHYTQKSLSLSPGK
SEQ ID NO. 22 mouse IgG Fc	RSPRGPTIKCPPCKCPAPNLLGGPSVFIFPPKIKDVLMSLSPIVTCVV VDVSEDDPDVQISWFWNNVEVHTAQTQTHREDYNSTLRVVSALPIQH QDWMSGKEFKCKVNNKDLPAPIERTISKPKGSVRAPQVYVLPPEEE

TABLE 1-continued

Oliomeric immunogenic composition fusion protein segment sequences	
Sequence Identifier	Amino Acid Sequence
	MTKKQVTLTTCMVTDFMPEDIYVEWTNNGKTELNYKNTEPVLDS DGSY FMYSKLRVEKKNWVERNSYSCSVHEGLHNHHTTKSFSRTPGK
SEQ ID NO. 23 rabbit IgG Fc	RSSKPTCPPPELLGGPSVFI FPPKPKDTLMI SRTPEVTCVVVDVSQDDP EVQFTWYINNEQVRTARPPPLREQQFNSTIRVVSTLPIAHQDWLRGKEF KCKVHNKALPAPIEKTISKARGQPLEPKVYTMGPPREELSSRSVSLTC MINGFYPSDISVEWEKNGKAEDNYKTTPAVLDS DGSYFLYSKLSVPTS EWQRGDVFTCSVMHEALHNHYTQKSISRSPGK
SEQ ID NO. 24 N46Fd	TLTVQARQLLSGIVQQQNNLLRAIEAQQHLLQLTVWGIKQLQARIL <b>GYIP</b> <b>EAPRDGQAYVRKDGEWVLLSTFL</b>
SEQ ID NO. 25 N46hFc	TLTVQARQLLSGIVQQQNNLLRAIEAQQHLLQLTVWGIKQLQARILRSD KHTCPCPAPELLGGPSVFLFPPKPKDTLMI SRTPEVTCVVVDVSHE DPEVKFNWYVDGVEVHNAKTKPREEQYNSTYRVVSVLTVLHQDWLN GKEYKCKVSNKALPAPIEKTISKAKGQPREPQVYTLPPSREEMTKNQV SLTCLVKGFYPSDIAVEWESNGQPENNYKTTPPVLDSDGSFFLYSKLT VDKSRWQQGNV FSCSVMHEGLHNHYTQKSLSLSPGK
SEQ ID NO. 26 N46FdhFc	TLTVQARQLLSGIVQQQNNLLRAIEAQQHLLQLTVWGIKQLQARIL <b>GYIP</b> <b>EAPRDGQAYVRKDGEWVLLSTFL</b> RS DKHTCPCPAPELLGGPSVFL FPPKPKDTLMI SRTPEVTCVVVDVSHEDPEVKFNWYVDGVEVHNAKT KPREEQYNSTYRVVSVLTVLHQDWLNGKEYKCKVSNKALPAPIEKTIS KAKGQPREPQVYTLPPSREEMTKNQVSLTCLVKGFYPSDIAVEWESN GQPENNYKTTPPVLDSDGSFFLYSKLTVDKSRWQQGNV FSCSVMHE GLHNHYTQKSLSLSPGK
SEQ ID NO. 27 N46FdmFc	TLTVQARQLLSGIVQQQNNLLRAIEAQQHLLQLTVWGIKQLQARIL <b>GYIP</b> <b>EAPRDGQAYVRKDGEWVLLSTFL</b> RS DKHTCPCPAPELLGGPSVFL FPPKPKDTLMI SRTPEVTCVVVDVSHEDPEVKFNWYVDGVEVHNAKT KPREEQYNSTYRVVSVLTVLHQDWLNGKEYKCKVSNKALPAPIEKTIS KAKGQPREPQVYTLPPSREEMTKNQVSLTCLVKGFYPSDIAVEWESN GQPENNYKTTPPVLDSDGSFFLYSKLTVDKSRWQQGNV FSCSVMHE GLHNHYTQKSLSLSPGK
SEQ ID NO. 28 N46FdrFc	TLTVQARQLLSGIVQQQNNLLRAIEAQQHLLQLTVWGIKQLQARIL <b>GYIP</b> <b>EAPRDGQAYVRKDGEWVLLSTFL</b> RS DKHTCPCPAPELLGGPSVFL FPPKPKDTLMI SRTPEVTCVVVDVSHEDPEVKFNWYVDGVEVHNAKT KPREEQYNSTYRVVSVLTVLHQDWLNGKEYKCKVSNKALPAPIEKTIS KAKGQPREPQVYTLPPSREEMTKNQVSLTCLVKGFYPSDIAVEWESN GQPENNYKTTPPVLDSDGSFFLYSKLTVDKSRWQQGNV FSCSVMHE GLHNHYTQKSLSLSPGK
SEQ ID NO. 29 IQN46	<b>RMQIEDKIEEIESKQKKIENEIARIKKT</b> TLTVQARQLLSGIVQQQNNLLR AIEAQQHLLQLTVWGIKQLQARIL
SEQ ID NO. 30 IQN46hFc	<b>RMQIEDKIEEIESKQKKIENEIARIKKT</b> TLTVQARQLLSGIVQQQNNLLR AIEAQQHLLQLTVWGIKQLQARILRSDKHTCPCPAPELLGGPSVFLF PPKPKDTLMI SRTPEVTCVVVDVSHEDPEVKFNWYVDGVEVHNAKTK PREEQYNSTYRVVSVLTVLHQDWLNGKEYKCKVSNKALPAPIEKTISK AKGQPREPQVYTLPPSREEMTKNQVSLTCLVKGFYPSDIAVEWESNG QPENNYKTTPPVLDSDGSFFLYSKLTVDKSRWQQGNV FSCSVMHEGL HNHYTQKSLSLSPGK
SEQ ID NO. 31 IZN46	<b>IKKEIEAIKKEQEAIKKKIEAIEKEI</b> TLTVQARQLLSGIVQQQNNLLRAIEA QQHLLQLTVWGIKQLQARIL
SEQ ID NO. 32 IZN46hFc	<b>IKKEIEAIKKEQEAIKKKIEAIEKEI</b> TLTVQARQLLSGIVQQQNNLLRAIEA QQHLLQLTVWGIKQLQARILRSDKHTCPCPAPELLGGPSVFLFPPK PKDTLMI SRTPEVTCVVVDVSHEDPEVKFNWYVDGVEVHNAKTKPRE EQYNSTYRVVSVLTVLHQDWLNGKEYKCKVSNKALPAPIEKTISKAKG QPREPQVYTLPPSREEMTKNQVSLTCLVKGFYPSDIAVEWESNGQPE NNYKTTPPVLDSDGSFFLYSKLTVDKSRWQQGNV FSCSVMHEGLHNH YTQKSLSLSPGK

As used herein, nucleotide sequences which are substan-  
tially the same share at least about 90% identity, and amino  
acid sequences which are substantially the same typically  
share more than 95% amino acid identity. It is recognized,  
however, that proteins (and DNA or mRNA encoding such  
proteins) containing less than the above-described level of  
homology arising as splice variants or that are modified by  
conservative amino acid substitutions (or substitution of

degenerate codons) are contemplated to be within the scope  
of the present disclosure. As readily recognized by those of  
skill in the art, various ways have been devised to align  
sequences for comparison, e.g., Blosum 62 scoring matrix, as  
described by Henikoff and Henikoff in Proc. Natl. Acad. Sci.  
USA 89:10915 (1992). Algorithms conveniently employed  
for this purpose are widely available (see, for example,  
Needleman and Wunsch in J. Mol. Bio. 48:443 (1970).



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Therefore, disclosed herein are amino acid sequences 85%, 90%, 95%, 98%, 99% or 100% identical to SEQ ID NOs:1-32.

Also disclosed herein are conservative amino acid substitutions of amino acids sequences SEQ ID NOs:1-32. Conservative amino acids substitutions are defined as changed, which although they alter the primary sequence of the protein or peptide, do not normally alter its function. Conservative amino acid substitutions typically include substitutions within the following groups: glycine and alanine; valine, isoleucine, and leucine; aspartic acid and glutamic acid; asparagine and glutamine; serine and threonine; lysine and arginine; and phenylalanine and tyrosine.

In one embodiment, the claimed fusion proteins can be constructed using overlapping primers. In another embodiment, the DNA sequence (GGCTATATTCCG GAAGCGC-CGCGTGATGGCCAGGCGTATGTGCG-TAAAGATGGCGAATGGGTGCTG

CTGTCTACCTTTCTG; SEQ ID NO:36) encoding Fd (aa sequence: GYIPEAPRDG QAYVRKDGWVLLSTFL) is synthesized first. Separate PCR products of the gp41 sequence and Fd are generated and the gp41 segment-Fd fusion fragment is amplified by one-round PCR using a gp41 forward primer and Fd reverse primer with gp41 (or a fragment thereof) and Fd DNA (PCR products) as templates. The amplified gp41 segment-Fd PCR product is then inserted into the hFc vector, to get an gp41-Fd-hFc recombinant plasmid encoding gp41-Fd-hFc fusion proteins. For the purposes of this discussion, the term "gp41" or "gp41 component" refers to the full-length gp41 protein, or a fragment thereof.

The gp41 component (gp41), trimerization or oligomerization motif component (TM) and the immunoenhancer component (IE) of the fusion proteins can be constructed in a variety of orientations including, but not limited to, gp41-TM-IE, gp41-IE-TM, IE-gp41-TM, TM-gp41-IE, etc. In one embodiment, the components are arranged gp41-TM-IE.

In one embodiment, pFUSE-hIgG1-Fc (human Fc, hFc), pFUSE-mIgG2a-Fc2 (murine Fc, mFc), or pFUSE-rIgG2-Fc2 (rabbit Fc, rFc) vectors are used for construction of the disclosed fusion proteins. In another embodiment, the fusion proteins can be expressed from other mammalian cell expression vectors, including, but not limited to, pcDNA3.1, pcDNA6-His, PEE13.1, PEE1.41, pCMV-NEO-BAM, pSV2, and pCMV1,2,3,4,5,6. In another embodiment, the fusion proteins can be expressed from insect cell expression vectors including, but not limited to, pAcGP67, pFastBac Dual, and pMT/V5-His-TOPO. In yet another embodiment, the fusion proteins can be expressed from *E. coli* expression vectors including, but not limited to, pET, pET-SUMO, and pGEX vectors with GST.

The following expression systems are suitable for use in expressing the disclosed fusion proteins: mammalian cell expression systems such as, but not limited to, pcDNA expression system, and GS Gene expression system; insect cell expression systems such as, but not limited to, Bac-to-Bac expression system, baculovirus expression system and DES expression systems; and *E. coli* expression systems including, but not limited to, pET, pSUMO and GST expression systems.

Advantages of proteins expressed in mammalian cell expression systems include the follows. The mammalian cell expression system is a relatively mature eukaryotic system for expression of recombinant proteins. There is a far higher chance to get correctly folded soluble proteins with proper glycosylation, making the expressed protein maintain native conformation and keep sufficient bioactivity if necessary. This system can either transient or stable express recombi-

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nant antigens, and promote signal synthesis. Recombinant proteins expressed in this way may keep good antigenicity and immunogenicity. However, both insect and bacterial expression systems provide inexpensive and efficient expression of active proteins, particularly when glycosylation is not required for bioactivity.

The purification systems are dependent on whether a tag is linked or fused with the fusion proteins. When the fusion proteins are fused with IgG Fc vectors, Protein A or Protein G affinity chromatography is used for the purification. If the fusion proteins are fused with GST proteins, the GST columns will be used for the purification. If the fusion proteins link with 6xHis tag at the N- or C-terminal, the expressed proteins are purified using His tag columns. If no tag is linked with recombinant proteins, the expressed proteins are purified using Fast protein liquid chromatography (FPLC), High performance liquid chromatography (HPLC) or other chromatographies.

In certain embodiments, the immunogenic compositions further comprise an adjuvant. Adjuvants suitable for use in animals include, but are not limited to, Freund's complete or incomplete adjuvants, Sigma Adjuvant System (SAS), and Ribi adjuvants. Adjuvants suitable for use in humans include, but are not limited to, MF59 (an oil-in-water emulsion adjuvant), aluminum hydroxide, -phosphate or -oxide, HAVLO-GEN® (an acrylic acid polymer-based adjuvant, Intervet Inc., Millsboro, Del.), polyacrylic acids, oil-in-water or water-in-oil emulsion based on, for example a mineral oil, such as BAYOL™ or MARCOL™ (Esso Imperial Oil Limited, Canada), or a vegetable oil such as vitamin E acetate, saponins, and *Onchocerca volvulus* activation-associated protein-1 (ASP-1) (see US 20060039921, which is incorporated by reference herein for all it discloses regarding ASP-1 adjuvants). However, components with adjuvant activity are widely known and, generally, any adjuvant may be utilized that does not adversely interfere with the efficacy or safety of the vaccine and/or immunogenic composition.

Vaccine and immunogenic compositions according to the various embodiments disclosed herein can be prepared and/or marketed in the form of a liquid, frozen suspension or in a lyophilized form. Typically, vaccines and/or immunogenic compositions prepared according to the present disclosure contain a pharmaceutically acceptable carrier or diluent customarily used for such compositions. Carriers include, but are not limited to, stabilizers, preservatives and buffers. Suitable stabilizers are, for example SPGA, Tween compositions (such as are available from A.G. Scientific, Inc., San Diego, Calif.), carbohydrates (such as sorbitol, mannitol, starch, sucrose, dextran, glutamate or glucose), proteins (such as dried milk serum, albumin or casein) or degradation products thereof. Non-limiting examples of suitable buffers include alkali metal phosphates. Suitable preservatives are thimerosal, merthiolate and gentamicin. Diluents include water, aqueous buffer (such as buffered saline), alcohols and polyols (such as glycerol).

Also disclosed herein are methods for inducing an immune response to HIV using the disclosed fusion proteins. Generally, the vaccine or immunogenic composition may be administered subcutaneously, intradermally, submucosally, or intramuscularly in an effective amount to prevent infection from HIV and/or treat an infection with HIV. An effective amount is defined as an amount of immunizing fusion protein that will induce immunity in the vaccinated animals, against challenge by a virulent virus. Immunity is defined herein as the induction of a significant higher level of protection in a population of the animal after vaccination compared to an unvaccinated group.



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Further, in various formulations of the vaccines and/or immunogenic compositions, suitable excipients, stabilizers and the like may be added.

## EXAMPLES

## Example 1

## Design of Immunoenhancer-Linked HIV-1 gp41 NHR-Trimer-Based Vaccine

As shown in FIG. 1A, the HIV-1 HXB2 gp41 consists of an ectodomain, a transmembrane (TM) and cytoplasmic (CP) domains. The ectodomain contains: fusion peptide (FP); N-terminal heptad repeat (NHR), which has a pocket-forming sequence (underlined); Immunodominant (ID) loop; C-terminal heptad repeat (CHR), which contains a pocket-binding sequence (underlined); and a membrane proximal external region (MPER). The residue numbers of each region correspond to their positions in gp160 of HIV-1 HXB2. FIGS. 1B-E show the gp41 NHR-trimeric structures: B) IQN17 and (ccIZN17)<sub>3</sub>, which consists of a 17-mer pocket-forming sequence and a trimerization motif, GCN4-pIQI (IQ) or IZ; C) N46, a NHR-peptide (residues 536-581) containing the N17 pocket-forming sequence; D) N46Fd, a recombinant protein consisting of N46 and foldon (Fd), a trimerization motif; and E) N46FdhFc, a recombinant protein containing N46Fd and human IgG Fc. The conserved hydrophobic pocket is circled.

## Example 2

## Construction of Plasmids Encoding N46Fd and N46FdhFc

The N46 sequence was derived from the NHR region of HIV-1HXB2 gp41. The pHXB2-env plasmid was employed as a template to synthesize a DNA fragment encoding the N46 amino acid sequence. All primers and the DNA fragment encoding the Fd sequence were synthesized by Integrated DNA Technologies (Coralville, Iowa). The N46 DNA fragment was then linked with Fd DNA fragment using an overlapping primer design program. The N46Fd DNA fragment was applied as the template for PCR amplification using the forward primer 5'-CCGGAATTCGACGCTGACGGTACAGG-3' (SEQ ID NO:33) and the reverse primer 5'-GGAA-GATCTTCAGTGGTGGTGGTG GTGGTGCAGAAAGG-TAGA-3' (SEQ ID NO:34). The amplified N46Fd DNA fragment was digested by EcoRI and BglII and inserted into the vector pFUSE-hIgG1-Fc2, designated pFUSE-N46Fd and encoded the N46Fd fusion protein. Subsequently, the pFUSE-N46FdFc plasmid DNA encoding the N46FdhFc protein was constructed using the same method of preparation for pFUSE-N46Fd with the following exceptions: (i) the pFUSE-N46Fd instead of the pHXB2-N46Fd plasmid was used as the template, and (ii) the following reverse primer was used: 5'-GGAAGATCTCAGAAAGGTAGACAG-3' (SEQ ID NO:35; containing no stop codon). The sequences of all constructs were confirmed by DNA sequencing. FIG. 2 shows the maps of the plasmids encoding N46Fd (A) and N46FdhFc (B). "6H" and "TGA\*" represent 6-His tag and stop codon, respectively.

## Example 3

Expression of N46Fd and N46FdhFc in Mammalian Cells and *E. Coli*

The plasmids of pFUSE-N46Fd and pFUSE-N46FdhFc were transformed into DH5 $\alpha$  cells. A single colony from

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freshly streaked selective plate containing 25  $\mu$ g/ml Zeocin was picked and inoculated in a starter culture of 5 ml LB medium containing 25  $\mu$ g/ml Zeocin with shaking at 250 rpm for 8 hrs at 37° C. Then 2 ml of the starter culture supernatants was applied to 400 ml LB containing 25  $\mu$ g/ml Zeocin. After incubation overnight at 37° C., the broth was centrifuged at 6,000 rpm (4,500 $\times$ g) for 15 min. The plasmid was purified using a Plasmid Purification Kit (QIAGEN, Valencia, Calif.) and then transfected to 293T cells using Lipofectamine 2000 reagent (Invitrogen, Carlsbad, Calif.) according to the manufacturer's protocol. Fresh DMEM was supplied next day. N46Fd and N46FdhFc were purified as described above.

For expression of N46FdhFc in *E. coli*, the plasmids pGEX-N46Fd and pGEX-N46FdhFc were transformed into Rosetta 2 (DE3) LysS complete cells, respectively, which were incubated in LB medium containing 100  $\mu$ g/ml ampicillin at 37° C. overnight. One colony was selected and transferred to 5 ml of LB medium. After incubation at 37° C. for 16 hrs, 2 ml culture supernatants were collected and transferred to 400 ml LB containing 100  $\mu$ g/ml ampicillin and 0.5 mM IPTG to induce protein expression. After culture for 4 hrs, the pellets were collected after centrifugation at 6,000 rpm (4,500 $\times$ g) for 20 min, and then sonicated three times (10 min each time). After centrifugation at 14,000 rpm (20,000 $\times$ g) for 30 min and filtration with 0.45  $\mu$ m membrane, the supernatants were passed through a GST-binding column and the GST-tag was removed by treatment with PreScission Protease (GE Healthcare, N.J.). N46Fd was purified by using a Ni-NTA His-Bind Superflow column since the foldon fragment in N46Fd can bind with his-binding beads. N46FdhFc was further purified by loading the eluted fractions to Protein A-Sepharose 4 Fast Flow column (Amersham Biosciences, Piscataway, N.J.). After washing, N46FdhFc was eluted with glycine-HCl buffer (pH 2.7) and neutralized with 1 M Tris-HCl buffer (pH 9), which was replaced by PBS using 10 kd filter (Millipore Corporation, Billerica, Mass.) through centrifugation at 2,900 rpm (2,003 $\times$ g) for 3 times and 20 min for each time. The purified protein was stored at -4° C. until use.

## Example 4

## Characterization of N46, N46Fd and N46FdhFc

The recombinant fusion proteins N46Fd and N46FdhFc as well as the synthetic peptide N46 were analyzed by SDS-PAGE, acid native PAGE, and Western blot. Purified N46, N46Fd and N46FdhFc (boiled and unboiled) were analyzed by SDS-PAGE using 10-20% Tris-Tricine Gradient Gels (Invitrogen). The peptide or proteins were also analyzed by acid native PAGE as described by Sackett et al. (J. Biol. Chem. 281(31):21755-62, 2006). Briefly, 10% polyacrylamide continuous native gels at pH 3.4 were prepared using 30 mM  $\beta$ -alanine and 42 mM formic acid and pre-run in the same buffer containing 100  $\mu$ M TCEP as a reducing agent. Gel electrophoresis was performed with a constant voltage of 125 V at room temperature for 2 h. The gel was then stained with Coomassie blue and imaged with a Fluor Chem 8800 imaging system (Alpha Innotech Corp., San Leandro, Calif.). For western-blotting, the peptide or proteins within the gels after separation by SDS-PAGE or acid native PAGE were transferred onto a 0.45  $\mu$ m-pore nitrocellulose membrane (Amersham Pharmacia Biotech, UK) at 100 V for 2 h and then 60 V for 1 h. The blotted membranes were rinsed with PBST (PBS with 0.1% Tween 20) three times for 5 min and then blocked in fresh PBST containing 5% non-fat dried milk at 4° C. overnight. After several washes, the membranes were incubated in rabbit anti-N46 IgG (1  $\mu$ g/ml). HRP-conjugated goat



anti-rabbit IgG (for anti-N46) and the ECL substrate solution (Amersham) were added sequentially. The blots were then visualized with autoradiography films.

As shown in FIG. 3A, without boiling, N46Fd was found in trimeric and monomeric forms, while N46FdhFc was found mainly in trimeric form under reducing condition because the Fd-stabilized trimer is resistant to reducing reagent. With boiling, N46Fd and N46FdhFc were mainly in monomeric and dimeric forms, respectively. N46 was in monomeric form under reducing condition with or without boiling. All the major bands reacted with polyclonal anti-N46 antibodies by Western blots (FIG. 3B). N46, N46Fd and N46FdhFc displayed no bands in native PAGE because they contain a net positive charge, while each of them showed a single band in acid native PAGE (stained with Coomassie blue) (FIG. 4A). All three immunogens reacted with rabbit anti-N46 antibodies by Western blots (FIG. 4B). These results suggest that N46Fd and N46FdhFc are in trimeric or other oligomeric forms.

Example 5

N46FdhFc Induced Potent Neutralizing Antibodies Against HIV-1

Balb/c mice (6-8 weeks old, 5 mice/group) were immunized subcutaneously with N46 peptide, recombinant N46Fd or N46FdhFc, in the presence of complete Freund's adjuvant (CFA, Sigma) and boosted three times with the same antigen plus incomplete Freund's adjuvant (IFA) as outline in Table 2:

TABLE 2

Mouse immunization procedure	
Day	Procedure
0	Prebleed, primary immunization, s.c. injection of 20 µg of an antigen with CFA
30	Boost, s.c. injection of 10 µg of an antigen with IFA
50	Boost, i.p. injection of 10 µg of an antigen in PBS
57	Boost, i.p. injection of 10 µg of an antigen in PBS
64	Terminal bleed

All sera were heat-inactivated at 56° C. for 30 min and kept at 4° C. until use.

Balb/c mice were immunized with adjuvant and N46, N46Fd or N46FdhFc and individual mouse serum was collected seven days after the last boost and tested for neutralizing activity against HIV-1 IIIB infection by the XTT assay for measuring HIV-1-mediated CPE. The XTT assay was conducted as previously described in Jiang et al. (J. Exp. Med. 174, 1557-1563, 1991). Briefly, MT-2 cells were infected with HIV-1 IIIB strain at 100 TCID<sub>50</sub> (50% tissue culture infective dose) in RPMI 1640 medium containing 10% FBS in the presence or absence of an antibody or antiserum at a series of 2-fold dilution at 37° C. overnight. The culture supernatants were then removed and fresh media were added. On day 6 post-infection, 50 µl of XTT solution (1 mg/ml) containing 0.02 µM of phenazine methosulphate (PMS) was added. After 4 hrs, absorbance at 450 nm (A<sub>450</sub>) was measured with an ELISA reader and the ND50 values were calculated as described above. As shown in FIG. 5 and Table 3, sera from mice immunized with N46 and N46Fd exhibited no detectable inhibitory activity, while sera from all the five mice vaccinated with N46FdhFc exhibited potent neutralizing activity.

These antisera were further tested for their neutralizing activity against infection by HIV-1 Bal (subtype B, R5) and an enfuvirtide-resistant HIV-1 strain NL4-3V38A/N42T in

CEMx174 5.25M7 cells using luciferase assay by using a luciferase assay. Briefly, 50 µl of an antibody or antiserum from immunized mice at a series of dilution was incubated with an equal volume of an HIV-1 isolate at 0.01 multiplicity of infection (MOI) at 37° C. for 30 min, followed by addition of the mixture to 100 µl CEMx174 5.25M7 or TZM-bl cells (1×10<sup>5</sup>/ml) that were pre-cultured in a 96-well plate at 37° C. overnight. After further culture at 37° C. for 3 days, the cells were harvested and lysed with 50 µl lysing reagent. The luciferase activity was analyzed using a luciferase kit (Promega Corp.) and a luminometer (Model: Ultra 386, Tecan) according to the manufacture's instruction. The ND50 value of an antiserum was calculated using the software CalcuSyn. Similarly, anti-N46FdhFc antisera exhibited potent neutralizing activity against these two HIV-1 strains (FIG. 6 and Table 3).

The neutralizing activity of antibodies or antisera against primary HIV-1 isolates was determined as previously described in Jiang et al. (Antimicrob. Agents Chemother. 48, 4349-4359, 2004). Briefly, the peripheral blood mononuclear cells (PBMCs) were isolated from the blood of healthy donors using a standard density gradient (Histopaque-1077, Sigma) centrifugation. After incubation at 37° C. for 2 h, the nonadherent cells were collected and resuspended at 5×10<sup>5</sup>/ml in RPMI 1640 medium containing 10% FBS, 5 µg of phytohemagglutinin (PHA)/ml, and 100 U of IL-2/ml, followed by incubation at 37° C. for 3 days. The PHA-stimulated cells were infected with the corresponding primary HIV-1 isolates at a multiplicity of infection (MOI) of 0.01 in the absence or presence of an antibody at a series of dilutions. The supernatants were collected 7 days post infection and mixed with equal volumes of 5% Triton X-100 for the detection of the p24 protein using ELISA as described in Jiang et al. with modification. Briefly, the wells of polystyrene plates (Immulon 1B, Dynex Technology, Chantilly, Va.) were coated with mouse anti-p24 mAb 183-12H-5C (NIH ARRRP) at 5 µg/ml in 0.05 M carbonate buffer (pH 9.6) at 4° C. overnight, followed by washes with PBS-T buffer (PBS containing 0.1% Tween-20) and blocking with PBS containing 2% dry fat-free milk (Bio-Rad Inc., Hercules, Calif.). Triton X-100 virus lysates were then added and incubated at 37° C. for 1 h. After extensive washes, human anti-p24 mAb (37G12, purchased from Polymun, Vienna, Austria), biotin labeled anti-human IgG1 (Santa Cruz Biotech., Santa Cruz, Calif.), SA-HRP and TMB were added sequentially. Reactions were terminated by addition of 1N H<sub>2</sub>SO<sub>4</sub>. Absorbance at 450 nm was recorded in an ELISA reader (Ultra 384, Tecan). Recombinant protein p24 (US Biological, Swampscott, Mass.) was used for establishing a standard dose response curve. These antisera against N46FdhFc also significantly neutralized infection of PBMCs by primary HIV-1 isolates 94UG103 (A, X4R5) and 92US657 (B, R5). However, the antisera directed against N46 and N46Fd at 1:40 dilution had no significant neutralizing activity against any of the viruses tested (Table 3). These results suggest that N46FdhFc is able to elicit antibodies with broad neutralizing activities.

TABLE 3

Neutralizing antibody titers of antisera directed against N46, N46Fd, and N46FdhFc			
HIV-1 (subtype, tropism)	ND50 of antisera against		
	N46	N46Fd	N46FdhFc
IIIB (B, X4) <sup>1</sup>	<40	<40	663
Bal (B, R5) <sup>2</sup>	<40	<40	2314



TABLE 3-continued

Neutralizing antibody titers of antisera directed against N46, N46Fd, and N46FdhFc			
HIV-1	ND50 of antisera against		
(subtype, tropism)	N46	N46Fd	N46FdhFc
NL43 V38A/N42T (B, R5) <sup>2</sup>	<40	<40	732
94UG103 (A, X4R5) <sup>3</sup>	<40	<40	265
92US657(B, R5) <sup>3</sup>	<40	<40	142

<sup>1</sup>Neutralizing activity of the mouse antisera against infection by HIV-1 IIIB was tested in MT-2 cells by XTT assay;  
<sup>2</sup>Neutralizing activity of the mouse antisera against infection by HIV-1 Bal and NL43 was tested in CEMx174 5.25M7 cells using luciferase assay;  
<sup>3</sup>Neutralizing activity of the mouse antisera against infection by primary HIV-1 isolates 94UG103 and p2US657 was tested in PBMCs by p24 assay.

Example 6

Antibodies Induced by N46FdhFc Inhibited HIV-1-Mediated Cell-Cell Fusion

The HIV gp41 fusion intermediate (NHR-trimer) plays a critical role in the HIV-1 fusion process. Therefore, the antibodies induced against the gp41 NHR-trimer may have inhibitory activities on HIV-induced cell-cell fusion. The inhibitory activities of the antibodies or antisera on HIV-induced cell-cell fusion were determined using a dye transfer assay as described by Jiang et al. (Antimicrob. Agents Chemother. 48, 4349-4359, 2004). Briefly, H9/HIV-1<sub>IIIB</sub> cells were pre-labeled with a fluorescent dye, Calcein AM (Molecular Probes, Inc.), and incubated with a testing antibody or antiserum at a series of dilution at 37° C. for 30 min in a 96-well cell culture plate. Then the CD4+MT-2 cells were added to the H9/HIV-1<sub>IIIB</sub> cells at a ratio of 10:1, followed by incubation at 37° C. for 2 hrs. The fused and unfused calcein-labeled HIV-1-infected cells were counted under an inverted fluorescence microscope with an eyepiece micrometer disc. The percent inhibition of cell fusion by an antibody or antiserum and the EC50 values were calculated using the software CalcuSyn. Antisera from all the five mice immunized with N46FdhFc exhibited potent HIV-1 fusion inhibitory activity with average 1050 of 1:154 (FIG. 7), while the antisera from mice immunized with N46 and N46Fd exhibited no detectable inhibitory activity at 1:40.

Example 7

Antibodies Induced by N46FdhFc Bound Preferably to the gp41 Pocket Region

To determine the target site of the neutralizing antibodies elicited by the N46FdhFc, antibody titers in the anti-N46, anti-N46Fd and anti-N46FdhFc antisera was compared against a series of antigens, including N46, Fd, IQN17, and (ccIZN17)<sub>3</sub>. For assessing the titers of antibodies in mouse sera against the gp41 NHR-trimer, the wells of a 96-well polystyrene plate (Costar) were coated with N46, Fd, or NHR-trimer, e.g., IQN17, or (ccIZN17)<sub>3</sub>, at 10 µg/ml in 0.1 M Tris-HCl buffer (pH 8.8) at 4° C. overnight, and blocked with 2% non-fat milk in PBS for 3 hrs at 37° C. Mouse sera in a series of 2-fold dilution were added. The plate was washed with the washing buffer (PBS containing 0.01% Tween 20) for 6 times to remove any unbound antibodies. Horseradish peroxidase (HRP) linked goat-anti-mouse IgG was added, followed by incubation at 37° C. for 30 min. The reaction was

visualized by addition of the substrate 3,3',5,5'-tetramethylbenzidine (TMB) and A450 was measured by using an ELISA reader (Tecan US).

As shown in Table 4, anti-N46 sera exhibited much higher binding titers against the N46 peptide and lower antibody titers against IQN17 and (ccIZN17)<sub>3</sub> than those directed against N46Fd and N46FdhFc. This suggests that anti-N46 antibodies mainly bind to linear epitopes in the NHR domain, while the antibodies to N46Fd and N46FdhFc preferably react with conformational epitopes in the NHR-trimers. Notably, antisera directed against N46FdhFc bound to IQN17 and (ccIZN17)<sub>3</sub> much more strongly than the anti-N46Fd antisera. This indicates that N46FdhFc elicits high levels of antibodies that may bind specifically to the pocket-forming sequence (N17), which overlaps the C-terminal fragment of N46 in the immunogen (FIG. 1), and that the neutralizing epitope(s) may be located in the pocket region. High titers of anti-Fd and anti-human IgG Fc antibodies were detected in the mouse antisera against N46FdhFc. However, the antisera from the mice immunized only with Fd or human IgG Fc had no virus neutralizing activity (data not shown), suggesting that the anti-Fd and anti-Fc antibodies in the anti-N46FdhFc sera are not responsible for the HIV-1 neutralization activity. Interestingly, the titer of antibodies to Fd in the anti-N46Fd sera (1:320,000) was much higher than that in the anti-N46FdhFc sera (1:12,126) (Table 4). This suggests that, although Fd in the fusion protein is highly immunogenic, the addition of the human IgG Fc to the C-terminus of N46Fd (FIG. 1) might suppress the immunogenicity of Fd. This may be one of the reasons why N46FdhFc, but not N46Fd, elicits neutralizing antibody responses. Apparently, fusion of the Fc domain of human IgG to the C-terminus of N46Fd is critical for the design of N46FdhFc as an immunogen. Firstly, the Fc domain in N46FdhFc may suppress the immunogenicity of Fd, perhaps by covering Fd (FIG. 1E) to prevent contact of Fd with antigen-presenting cells (APCs), and thus resulting in reduction of the immune interference of Fd with the immunogenicity of NHR-trimer. Secondly, the Fc domain in an immunogen may have an immunoenhancing effect because Fc-tagged protein can bind the Fc receptor (FcR) on dendritic cells or other APCs and thus promote internalization of the immunogen and major histocompatibility complex (MHC) class II-restricted antigen presentation. Thirdly, conjugation of a protein with Fc may prolong the half-life of the immunogen in immunized animals and thus enhance its immunogenicity.

TABLE 4

The titers of antisera binding to N46, Fd, and NHR-trimers.			
Antigen	Reciprocal of geometric mean titers of mouse antisera		
	Anti-N46	Anti-N46FD	Anti-N46FdhFc
N46	161,270	10,079	9,190
Foldon	<100	320,000	12,126
IQN17	22,449	35,919	211,121
(ccIZN17) <sub>3</sub>	17,818	17,959	320,000

Example 8

Antibodies Induced by N46FdhFc Blocked the gp41 Six-Helix Bundle Formation

To investigate the mechanism by which anti-N46FdhFc antibodies neutralize HIV-1 infection and block HIV-1-me-



diated cell-cell fusion, the inhibitory activities of antibodies in the mouse anti-N46, anti-N46Fd and anti-N46FdHFc antisera were compared with normal mouse IgG as control. The inhibitory activity of antibodies on the 6-HB core formation between N46 (SEQ ID NO. 3) and biotinylated C34 (C34-biotin) was determined by an ELISA with a conformation-specific monoclonal antibody (mAb) NC-1 (Jiang et al. J. Virol. 72, 10213-10217, 1998) and as previously described (He et al. Proc. Natl. Acad. Sci. USA 105, 16332-16337, 2008). Briefly, an antibody at a series of dilutions was pre-incubated with equal amount of N46 at 37° C. for 30 min, followed by addition of C34-biotin (0.5 μM). The mixture was added to the wells of a microplate coated with mAb NC-1 IgG (2 μg/ml in 0.1M Tris, pH 8.8) and blocked with 2% non-fat milk in PBS. The plate was then incubated for 30 min and washed with the washing buffer for 6 times to remove any unbound peptide. SA-HRP and TMB were added sequentially and A450 was measured. The percent inhibition of 6-HB formation and the IC<sub>50</sub> values were calculated.

As shown in FIG. 8, anti-N46FdHFc IgG effectively blocked the 6-HB core formation between N46 and C34 as determined by ELISA. However, normal mouse IgG and antibodies specific for N46 and N46Fd had no significant effect on the inhibition of gp41 6-HB core formation. These results suggest that N46FdHFc elicits antibodies that bind preferably to the viral gp41 pocket region and thus block the fusion-active 6-HB core formation between the viral gp41 NHR and CHR, and consequently result in the inhibition of HIV-1 fusion with the target cell and viral entry.

Unless otherwise indicated, all numbers expressing quantities of ingredients, properties such as molecular weight, reaction conditions, and so forth used in the specification and claims are to be understood as being modified in all instances by the term “about.” Accordingly, unless indicated to the contrary, the numerical parameters set forth in the specification and attached claims are approximations that may vary depending upon the desired properties sought to be obtained by the present invention. At the very least, and not as an attempt to limit the application of the doctrine of equivalents to the scope of the claims, each numerical parameter should at least be construed in light of the number of reported significant digits and by applying ordinary rounding techniques. Notwithstanding that the numerical ranges and parameters setting forth the broad scope of the invention are approximations, the numerical values set forth in the specific examples are reported as precisely as possible. Any numerical value, however, inherently contains certain errors necessarily resulting from the standard deviation found in their respective testing measurements.

The terms “a,” “an,” “the” and similar referents used in the context of describing the invention (especially in the context of the following claims) are to be construed to cover both the singular and the plural, unless otherwise indicated herein or clearly contradicted by context. Recitation of ranges of values herein is merely intended to serve as a shorthand method of referring individually to each separate value falling within the range. Unless otherwise indicated herein, each individual value is incorporated into the specification as if it were indi-

vidually recited herein. All methods described herein can be performed in any suitable order unless otherwise indicated herein or otherwise clearly contradicted by context. The use of any and all examples, or exemplary language (e.g., “such as”) provided herein is intended merely to better illuminate the invention and does not pose a limitation on the scope of the invention otherwise claimed. No language in the specification should be construed as indicating any non-claimed element essential to the practice of the invention.

Groupings of alternative elements or embodiments of the invention disclosed herein are not to be construed as limitations. Each group member may be referred to and claimed individually or in any combination with other members of the group or other elements found herein. It is anticipated that one or more members of a group may be included in, or deleted from, a group for reasons of convenience and/or patentability. When any such inclusion or deletion occurs, the specification is deemed to contain the group as modified thus fulfilling the written description of all Markush groups used in the appended claims.

Certain embodiments of this invention are described herein, including the best mode known to the inventors for carrying out the invention. Of course, variations on these described embodiments will become apparent to those of ordinary skill in the art upon reading the foregoing description. The inventor expects skilled artisans to employ such variations as appropriate, and the inventors intend for the invention to be practiced otherwise than specifically described herein. Accordingly, this invention includes all modifications and equivalents of the subject matter recited in the claims appended hereto as permitted by applicable law. Moreover, any combination of the above-described elements in all possible variations thereof is encompassed by the invention unless otherwise indicated herein or otherwise clearly contradicted by context.

Specific embodiments disclosed herein may be further limited in the claims using consisting of or consisting essentially of language. When used in the claims, whether as filed or added per amendment, the transition term “consisting of” excludes any element, step, or ingredient not specified in the claims. The transition term “consisting essentially of” limits the scope of a claim to the specified materials or steps and those that do not materially affect the basic and novel characteristic(s). Embodiments of the invention so claimed are inherently or expressly described and enabled herein.

Furthermore, numerous references have been made to patents and printed publications throughout this specification. Each of the above-cited references and printed publications are individually incorporated herein by reference in their entirety.

In closing, it is to be understood that the embodiments of the invention disclosed herein are illustrative of the principles of the present invention. Other modifications that may be employed are within the scope of the invention. Thus, by way of example, but not of limitation, alternative configurations of the present invention may be utilized in accordance with the teachings herein. Accordingly, the present invention is not limited to that precisely as shown and described.

## SEQUENCE LISTING

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Gln	Ser	Asn	Leu	Leu	Arg	Ala	Ile	Glu	Ala	Gln	Gln	His	Leu	Leu	Gln
			20					25					30		
Leu	Thr	Val	Trp	Gly	Ile	Lys	Gln	Leu	Gln	Ala	Arg	Val	Leu		
		35					40					45			

<400> SEQUENCE: 10

Thr	Leu	Thr	Val	Gln	Val	Lys	Lys	Leu	Leu	Phe	Gly	Ile	Val	Gln	Gln
1				5					10					15	
Gln	Ser	Asn	Leu	Leu	Arg	Ala	Ile	Glu	Ala	Gln	Gln	His	Leu	Leu	Gln
			20					25					30		
Leu	Thr	Val	Trp	Gly	Ile	Lys	Gln	Leu	Gln	Ala	Arg	Val	Leu		
		35					40					45			

<400> SEQUENCE: 11

Ala Leu Thr Val Arg Thr His Thr Leu Ile Lys Gly Ile Val Gln Gln  
1 5 10 15

Gln Asp Asn Leu Leu Arg Ala Ile Gln Ala Gln Gln Gln Leu Leu Arg  
20 25 30

Leu Ser Val Trp Gly Ile Arg Gln Leu Arg Ala Arg Leu Leu  
35 40 45

<210> SEQ ID NO 12



[illegible]

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<210> SEQ ID NO 17  
<211> LENGTH: 63  
<212> TYPE: PRT  
<213> ORGANISM: Human immunodeficiency virus type 1  
  
<400> SEQUENCE: 17  
  
Ser Thr Met Gly Ala Ala Ser Met Thr Leu Thr Val Gln Ala Arg Gln  
1 5 10 15  
  
Leu Leu Ser Gly Ile Val Gln Gln Gln Asn Asn Leu Leu Arg Ala Ile  
20 25 30  
  
Glu Ala Gln Gln His Leu Leu Gln Leu Thr Val Trp Gly Ile Lys Gln  
35 40 45  
  
Leu Gln Ala Arg Ile Leu Ala Val Glu Arg Tyr Leu Lys Asp Gln  
50 55 60  
  
<210> SEQ ID NO 18  
<211> LENGTH: 27  
<212> TYPE: PRT  
<213> ORGANISM: Bacteriophage T4  
  
<400> SEQUENCE: 18  
  
Gly Tyr Ile Pro Glu Ala Pro Arg Asp Gly Gln Ala Tyr Val Arg Lys  
1 5 10 15  
  
Asp Gly Glu Trp Val Leu Leu Ser Thr Phe Leu  
20 25  
  
<210> SEQ ID NO 19  
<211> LENGTH: 28  
<212> TYPE: PRT  
<213> ORGANISM: Human immunodeficiency virus  
  
<400> SEQUENCE: 19  
  
Arg Met Lys Gln Ile Glu Asp Lys Ile Glu Glu Ile Glu Ser Lys Gln  
1 5 10 15  
  
Lys Lys Ile Glu Asn Glu Ile Ala Arg Ile Lys Lys  
20 25  
  
<210> SEQ ID NO 20  
<211> LENGTH: 26  
<212> TYPE: PRT  
<213> ORGANISM: Human immunodeficiency virus  
  
<400> SEQUENCE: 20  
  
Ile Lys Lys Glu Ile Glu Ala Ile Lys Lys Glu Gln Glu Ala Ile Lys  
1 5 10 15  
  
Lys Lys Ile Glu Ala Ile Glu Lys Glu Ile  
20 25  
  
<210> SEQ ID NO 21  
<211> LENGTH: 229  
<212> TYPE: PRT  
<213> ORGANISM: Homo sapiens  
  
<400> SEQUENCE: 21  
  
Arg Ser Asp Lys Thr His Thr Cys Pro Pro Cys Pro Ala Pro Glu Leu  
1 5 10 15  
  
Leu Gly Gly Pro Ser Val Phe Leu Phe Pro Pro Lys Pro Lys Asp Thr  
20 25 30  
  
Leu Met Ile Ser Arg Thr Pro Glu Val Thr Cys Val Val Val Asp Val  
35 40 45



[illegible]

<400> SEQUENCE: 22

Arg 1	Ser	Pro	Arg	Gly 5	Pro	Thr	Ile	Lys	Pro 10	Cys	Pro	Pro	Cys	Lys 15	Cys
Pro	Ala	Pro	Asn 20	Leu	Leu	Gly	Gly	Pro 25	Ser	Val	Phe	Ile	Phe 30	Pro	Pro
Lys	Ile	Lys 35	Asp	Val	Leu	Met	Ile 40	Ser	Leu	Ser	Pro	Ile 45	Val	Thr	Cys
Val	Val 50	Val	Asp	Val	Ser	Glu 55	Asp	Asp	Pro	Asp	Val 60	Gln	Ile	Ser	Trp
Phe 65	Val	Asn	Asn	Val	Glu 70	Val	His	Thr	Ala	Gln 75	Thr	Gln	Thr	His	Arg 80
Glu	Asp	Tyr	Asn 85	Ser	Thr	Leu	Arg	Val 90	Val	Ser	Ala	Leu	Pro	Ile 95	Gln
His	Gln	Asp	Trp 100	Met	Ser	Gly	Lys	Glu 105	Phe	Lys	Cys	Lys	Val 110	Asn	Asn
Lys	Asp	Leu 115	Pro	Ala	Pro	Ile	Glu 120	Arg	Thr	Ile	Ser	Lys 125	Pro	Lys	Gly
Ser 130	Val	Arg	Ala	Pro	Gln	Val	Tyr 135	Val	Leu	Pro	Pro 140	Pro	Glu	Glu	Glu
Met 145	Thr	Lys	Lys	Gln 150	Val	Thr	Leu	Thr	Cys	Met 155	Val	Thr	Asp	Phe	Met 160
Pro	Glu	Asp	Ile 165	Tyr	Val	Glu	Trp	Thr	Asn 170	Asn	Gly	Lys	Thr	Glu 175	Leu
Asn	Tyr	Lys	Asn 180	Thr	Glu	Pro	Val	Leu 185	Asp	Ser	Asp	Gly	Ser 190	Tyr	Phe

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Met	Tyr	Ser	Lys	Leu	Arg	Val	Glu	Lys	Lys	Asn	Trp	Val	Glu	Arg	Asn	
		195					200					205				
Ser	Tyr	Ser	Cys	Ser	Val	Val	His	Glu	Gly	Leu	His	Asn	His	His	Thr	
	210					215					220					
Thr	Lys	Ser	Phe	Ser	Arg	Thr	Pro	Gly	Lys							
225					230											
<210> SEQ ID NO 23																
<211> LENGTH: 225																
<212> TYPE: PRT																
<213> ORGANISM: Oryctolagus cuniculus																
<400> SEQUENCE: 23																
Arg	Ser	Ser	Lys	Pro	Thr	Cys	Pro	Pro	Pro	Glu	Leu	Leu	Gly	Gly	Pro	
1				5					10					15		
Ser	Val	Phe	Ile	Phe	Pro	Pro	Lys	Pro	Lys	Asp	Thr	Leu	Met	Ile	Ser	
			20					25					30			
Arg	Thr	Pro	Glu	Val	Thr	Cys	Val	Val	Val	Asp	Val	Ser	Gln	Asp	Asp	
		35					40					45				
Pro	Glu	Val	Gln	Phe	Thr	Trp	Tyr	Ile	Asn	Asn	Glu	Gln	Val	Arg	Thr	
	50					55					60					
Ala	Arg	Pro	Pro	Leu	Arg	Glu	Gln	Gln	Phe	Asn	Ser	Thr	Ile	Arg	Val	
65					70					75					80	
Val	Ser	Thr	Leu	Pro	Ile	Ala	His	Gln	Asp	Trp	Leu	Arg	Gly	Lys	Glu	
				85					90					95		
Phe	Lys	Cys	Lys	Val	His	Asn	Lys	Ala	Leu	Pro	Ala	Pro	Ile	Glu	Lys	
			100					105					110			
Thr	Ile	Ser	Lys	Ala	Arg	Gly	Gln	Pro	Leu	Glu	Pro	Lys	Val	Tyr	Thr	
		115					120					125				
Met	Gly	Pro	Pro	Arg	Glu	Glu	Leu	Ser	Ser	Arg	Ser	Val	Ser	Leu	Thr	
	130						135				140					
Cys	Met	Ile	Asn	Gly	Phe	Tyr	Pro	Ser	Asp	Ile	Ser	Val	Glu	Trp	Glu	
145					150					155					160	
Lys	Asn	Gly	Lys	Ala	Glu	Asp	Asn	Tyr	Lys	Thr	Thr	Pro	Ala	Val	Leu	
				165					170					175		
Asp	Ser	Asp	Gly	Ser	Tyr	Phe	Leu	Tyr	Ser	Lys	Leu	Ser	Val	Pro	Thr	
		180						185					190			
Ser	Glu	Trp	Gln	Arg	Gly	Asp	Val	Phe	Thr	Cys	Ser	Val	Met	His	Glu	
		195					200					205				
Ala	Leu	His	Asn	His	Tyr	Thr	Gln	Lys	Ser	Ile	Ser	Arg	Ser	Pro	Gly	
	210						215					220				
Lys																
225																
<210> SEQ ID NO 24																
<211> LENGTH: 73																
<212> TYPE: PRT																
<213> ORGANISM: Artificial																
<220> FEATURE:																
<223> OTHER INFORMATION: N46Fd fusion protein comprising N46 fragment of gp41 and foldon sequence																
<400> SEQUENCE: 24																
Thr	Leu	Thr	Val	Gln	Ala	Arg	Gln	Leu	Leu	Ser	Gly	Ile	Val	Gln	Gln	
1				5					10					15		
Gln	Asn	Asn	Leu	Leu	Arg	Ala	Ile	Glu	Ala	Gln	Gln	His	Leu	Leu	Gln	
			20					25					30			



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<210> SEQ ID NO 26
<211> LENGTH: 302
<212> TYPE: PRT
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<213> ORGANISM: Artificial  
<220> FEATURE:  
<223> OTHER INFORMATION: N46FdhFc fusion protein comprising the N46  
fragment of gp41, foldon sequence and human Fc  
  
<400> SEQUENCE: 26  
  
Thr Leu Thr Val Gln Ala Arg Gln Leu Leu Ser Gly Ile Val Gln Gln  
1 5 10 15  
  
Gln Asn Asn Leu Leu Arg Ala Ile Glu Ala Gln Gln His Leu Leu Gln  
20 25 30  
  
Leu Thr Val Trp Gly Ile Lys Gln Leu Gln Ala Arg Ile Leu Gly Tyr  
35 40 45  
  
Ile Pro Glu Ala Pro Arg Asp Gly Gln Ala Tyr Val Arg Lys Asp Gly  
50 55 60  
  
Glu Trp Val Leu Leu Ser Thr Phe Leu Arg Ser Asp Lys Thr His Thr  
65 70 75 80  
  
Cys Pro Pro Cys Pro Ala Pro Glu Leu Leu Gly Gly Pro Ser Val Phe  
85 90 95  
  
Leu Phe Pro Pro Lys Pro Lys Asp Thr Leu Met Ile Ser Arg Thr Pro  
100 105 110  
  
Glu Val Thr Cys Val Val Val Asp Val Ser His Glu Asp Pro Glu Val  
115 120 125  
  
Lys Phe Asn Trp Tyr Val Asp Gly Val Glu Val His Asn Ala Lys Thr  
130 135 140  
  
Lys Pro Arg Glu Glu Gln Tyr Asn Ser Thr Tyr Arg Val Val Ser Val  
145 150 155 160  
  
Leu Thr Val Leu His Gln Asp Trp Leu Asn Gly Lys Glu Tyr Lys Cys  
165 170 175  
  
Lys Val Ser Asn Lys Ala Leu Pro Ala Pro Ile Glu Lys Thr Ile Ser  
180 185 190  
  
Lys Ala Lys Gly Gln Pro Arg Glu Pro Gln Val Tyr Thr Leu Pro Pro  
195 200 205  
  
Ser Arg Glu Glu Met Thr Lys Asn Gln Val Ser Leu Thr Cys Leu Val  
210 215 220  
  
Lys Gly Phe Tyr Pro Ser Asp Ile Ala Val Glu Trp Glu Ser Asn Gly  
225 230 235 240  
  
Gln Pro Glu Asn Asn Tyr Lys Thr Thr Pro Pro Val Leu Asp Ser Asp  
245 250 255  
  
Gly Ser Phe Phe Leu Tyr Ser Lys Leu Thr Val Asp Lys Ser Arg Trp  
260 265 270  
  
Gln Gln Gly Asn Val Phe Ser Cys Ser Val Met His Glu Gly Leu His  
275 280 285  
  
Asn His Tyr Thr Gln Lys Ser Leu Ser Leu Ser Pro Gly Lys  
290 295 300  
  
<210> SEQ ID NO 27  
<211> LENGTH: 302  
<212> TYPE: PRT  
<213> ORGANISM: Artificial  
<220> FEATURE:  
<223> OTHER INFORMATION: N46FdmFc comprising the N46 fragment of gp41,  
foldon sequence and murine Fc  
  
<400> SEQUENCE: 27  
  
Thr Leu Thr Val Gln Ala Arg Gln Leu Leu Ser Gly Ile Val Gln Gln  
1 5 10 15  
  
Gln Asn Asn Leu Leu Arg Ala Ile Glu Ala Gln Gln His Leu Leu Gln



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20							25					30			
Leu	Thr	Val	Trp	Gly	Ile	Lys	Gln	Leu	Gln	Ala	Arg	Ile	Leu	Gly	Tyr
		35					40					45			
Ile	Pro	Glu	Ala	Pro	Arg	Asp	Gly	Gln	Ala	Tyr	Val	Arg	Lys	Asp	Gly
		50			55						60				
Glu	Trp	Val	Leu	Leu	Ser	Thr	Phe	Leu	Arg	Ser	Asp	Lys	Thr	His	Thr
65					70				75				80		
Cys	Pro	Pro	Cys	Pro	Ala	Pro	Glu	Leu	Leu	Gly	Gly	Pro	Ser	Val	Phe
				85				90				95			
Leu	Phe	Pro	Pro	Lys	Pro	Lys	Asp	Thr	Leu	Met	Ile	Ser	Arg	Thr	Pro
		100						105				110			
Glu	Val	Thr	Cys	Val	Val	Val	Asp	Val	Ser	His	Glu	Asp	Pro	Glu	Val
		115				120						125			
Lys	Phe	Asn	Trp	Tyr	Val	Asp	Gly	Val	Glu	Val	His	Asn	Ala	Lys	Thr
		130				135				140					
Lys	Pro	Arg	Glu	Glu	Gln	Tyr	Asn	Ser	Thr	Tyr	Arg	Val	Val	Ser	Val
145					150				155				160		
Leu	Thr	Val	Leu	His	Gln	Asp	Trp	Leu	Asn	Gly	Lys	Glu	Tyr	Lys	Cys
				165				170				175			
Lys	Val	Ser	Asn	Lys	Ala	Leu	Pro	Ala	Pro	Ile	Glu	Lys	Thr	Ile	Ser
		180						185				190			
Lys	Ala	Lys	Gly	Gln	Pro	Arg	Glu	Pro	Gln	Val	Tyr	Thr	Leu	Pro	Pro
		195				200						205			
Ser	Arg	Glu	Glu	Met	Thr	Lys	Asn	Gln	Val	Ser	Leu	Thr	Cys	Leu	Val
		210				215				220					
Lys	Gly	Phe	Tyr	Pro	Ser	Asp	Ile	Ala	Val	Glu	Trp	Glu	Ser	Asn	Gly
225					230				235				240		
Gln	Pro	Glu	Asn	Asn	Tyr	Lys	Thr	Thr	Pro	Pro	Val	Leu	Asp	Ser	Asp
				245				250				255			
Gly	Ser	Phe	Phe	Leu	Tyr	Ser	Lys	Leu	Thr	Val	Asp	Lys	Ser	Arg	Trp
		260						265				270			
Gln	Gln	Gly	Asn	Val	Phe	Ser	Cys	Ser	Val	Met	His	Glu	Gly	Leu	His
		275				280						285			
Asn	His	Tyr	Thr	Gln	Lys	Ser	Leu	Ser	Leu	Ser	Pro	Gly	Lys		
		290				295				300					

<210> SEQ ID NO 28  
<211> LENGTH: 302  
<212> TYPE: PRT  
<213> ORGANISM: Artificial  
<220> FEATURE:  
<223> OTHER INFORMATION: N46FdrFc comprising N46 sequence of gp41, foldon sequence and rabbit Fc  
  
<400> SEQUENCE: 28

Thr	Leu	Thr	Val	Gln	Ala	Arg	Gln	Leu	Leu	Ser	Gly	Ile	Val	Gln	Gln	
1			5			10			15							
Gln	Asn	Asn	Leu	Leu	Arg	Ala	Ile	Glu	Ala	Gln	Gln	His	Leu	Leu	Gln	
20					25					30						
Leu	Thr	Val	Trp	Gly	Ile	Lys	Gln	Leu	Gln	Ala	Arg	Ile	Leu	Gly	Tyr	
35					40					45						
Ile	Pro	Glu	Ala	Pro	Arg	Asp	Gly	Gln	Ala	Tyr	Val	Arg	Lys	Asp	Gly	
50					55					60						
Glu	Trp	Val	Leu	Leu	Ser	Thr	Phe	Leu	Arg	Ser	Asp	Lys	Thr	His	Thr	
65					70					75					80	

Cys	Pro	Pro	Cys	Pro	Ala	Pro	Glu	Leu	Leu	Gly	Gly	Pro	Ser	Val	Phe	
				85					90					95		
Leu	Phe	Pro	Pro	Lys	Pro	Lys	Asp	Thr	Leu	Met	Ile	Ser	Arg	Thr	Pro	
				100					105					110		
Glu	Val	Thr	Cys	Val	Val	Val	Asp	Val	Ser	His	Glu	Asp	Pro	Glu	Val	
				115					120					125		
Lys	Phe	Asn	Trp	Tyr	Val	Asp	Gly	Val	Glu	Val	His	Asn	Ala	Lys	Thr	
				130					135					140		
Lys	Pro	Arg	Glu	Glu	Gln	Tyr	Asn	Ser	Thr	Tyr	Arg	Val	Val	Ser	Val	
				145					150					155		
Leu	Thr	Val	Leu	His	Gln	Asp	Trp	Leu	Asn	Gly	Lys	Glu	Tyr	Lys	Cys	
				165					170					175		
Lys	Val	Ser	Asn	Lys	Ala	Leu	Pro	Ala	Pro	Ile	Glu	Lys	Thr	Ile	Ser	
				180					185					190		
Lys	Ala	Lys	Gly	Gln	Pro	Arg	Glu	Pro	Gln	Val	Tyr	Thr	Leu	Pro	Pro	
				195					200					205		
Ser	Arg	Glu	Glu	Met	Thr	Lys	Asn	Gln	Val	Ser	Leu	Thr	Cys	Leu	Val	
				210					215					220		
Lys	Gly	Phe	Tyr	Pro	Ser	Asp	Ile	Ala	Val	Glu	Trp	Glu	Ser	Asn	Gly	
				225					230					235		
Gln	Pro	Glu	Asn	Asn	Tyr	Lys	Thr	Thr	Pro	Pro	Val	Leu	Asp	Ser	Asp	
				245					250					255		
Gly	Ser	Phe	Phe	Leu	Tyr	Ser	Lys	Leu	Thr	Val	Asp	Lys	Ser	Arg	Trp	
				260					265					270		
Gln	Gln	Gly	Asn	Val	Phe	Ser	Cys	Ser	Val	Met	His	Glu	Gly	Leu	His	
				275					280					285		
Asn	His	Tyr	Thr	Gln	Lys	Ser	Leu	Ser	Leu	Ser	Pro	Gly	Lys			
				290					295					300		

<400> SEQUENCE: 29

[illegible]

<400> SEQUENCE: 30



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Arg	Met	Lys	Gln	Ile	Glu	Asp	Lys	Ile	Glu	Glu	Ile	Glu	Ser	Lys	Gln
1				5					10					15	
Lys	Lys	Ile	Glu	Asn	Glu	Ile	Ala	Arg	Ile	Lys	Lys	Thr	Leu	Thr	Val
			20					25					30		
Gln	Ala	Arg	Gln	Leu	Leu	Ser	Gly	Ile	Val	Gln	Gln	Gln	Asn	Asn	Leu
		35					40					45			
Leu	Arg	Ala	Ile	Glu	Ala	Gln	Gln	His	Leu	Leu	Gln	Leu	Thr	Val	Trp
	50					55					60				
Gly	Ile	Lys	Gln	Leu	Gln	Ala	Arg	Ile	Leu	Arg	Ser	Asp	Lys	Thr	His
65				70						75					80
Thr	Cys	Pro	Pro	Cys	Pro	Ala	Pro	Glu	Leu	Leu	Gly	Gly	Pro	Ser	Val
				85					90					95	
Phe	Leu	Phe	Pro	Pro	Lys	Pro	Lys	Asp	Thr	Leu	Met	Ile	Ser	Arg	Thr
			100					105						110	
Pro	Glu	Val	Thr	Cys	Val	Val	Val	Asp	Val	Ser	His	Glu	Asp	Pro	Glu
		115					120					125			
Val	Lys	Phe	Asn	Trp	Tyr	Val	Asp	Gly	Val	Glu	Val	His	Asn	Ala	Lys
		130					135					140			
Thr	Lys	Pro	Arg	Glu	Glu	Gln	Tyr	Asn	Ser	Thr	Tyr	Arg	Val	Val	Ser
145					150						155				160
Val	Leu	Thr	Val	Leu	His	Gln	Asp	Trp	Leu	Asn	Gly	Lys	Glu	Tyr	Lys
				165					170					175	
Cys	Lys	Val	Ser	Asn	Lys	Ala	Leu	Pro	Ala	Pro	Ile	Glu	Lys	Thr	Ile
			180					185					190		
Ser	Lys	Ala	Lys	Gly	Gln	Pro	Arg	Glu	Pro	Gln	Val	Tyr	Thr	Leu	Pro
		195					200					205			
Pro	Ser	Arg	Glu	Glu	Met	Thr	Lys	Asn	Gln	Val	Ser	Leu	Thr	Cys	Leu
		210				215					220				
Val	Lys	Gly	Phe	Tyr	Pro	Ser	Asp	Ile	Ala	Val	Glu	Trp	Glu	Ser	Asn
225					230					235					240
Gly	Gln	Pro	Glu	Asn	Asn	Tyr	Lys	Thr	Thr	Pro	Pro	Val	Leu	Asp	Ser
				245					250					255	
Asp	Gly	Ser	Phe	Phe	Leu	Tyr	Ser	Lys	Leu	Thr	Val	Asp	Lys	Ser	Arg
			260					265					270		
Trp	Gln	Gln	Gly	Asn	Val	Phe	Ser	Cys	Ser	Val	Met	His	Glu	Gly	Leu
		275				280						285			
His	Asn	His	Tyr	Thr	Gln	Lys	Ser	Leu	Ser	Leu	Ser	Pro	Gly	Lys	
	290					295					300				
<210> SEQ ID NO 31															
<211> LENGTH: 72															
<212> TYPE: PRT															
<213> ORGANISM: Artificial															
<220> FEATURE:															
<223> OTHER INFORMATION: IZN46 fusion protein comprising the N46															
sequence of gp41 and IZ sequence															
<400> SEQUENCE: 31															
Ile	Lys	Lys	Glu	Ile	Glu	Ala	Ile	Lys	Lys	Glu	Gln	Glu	Ala	Ile	Lys
1				5					10					15	
Lys	Lys	Ile	Glu	Ala	Ile	Glu	Lys	Glu	Ile	Thr	Leu	Thr	Val	Gln	Ala
			20					25					30		
Arg	Gln	Leu	Leu	Ser	Gly	Ile	Val	Gln	Gln	Gln	Asn	Asn	Leu	Leu	Arg
		35				40					45				
Ala	Ile	Glu	Ala	Gln	Gln	His	Leu	Leu	Gln	Leu	Thr	Val	Trp	Gly	Ile
	50					55				60					

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<210> SEQ ID NO 33
<211> LENGTH: 26
<212> TYPE: DNA
<213> ORGANISM: Artificial
<220> FEATURE:
<223> OTHER INFORMATION: N46Fd forward primer
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<400> SEQUENCE: 33	
cgggaattcg acgctgacgg tacagg	26
<210> SEQ ID NO 34	
<211> LENGTH: 42	
<212> TYPE: DNA	
<213> ORGANISM: Artificial	
<220> FEATURE:	
<223> OTHER INFORMATION: N46Fd reverse primer	
<400> SEQUENCE: 34	
ggaagatctt cagtgggtgt ggtgggtggt cagaaaggta ga	42
<210> SEQ ID NO 35	
<211> LENGTH: 24	
<212> TYPE: DNA	
<213> ORGANISM: Artificial	
<220> FEATURE:	
<223> OTHER INFORMATION: N46FdhFc reverse primer	
<400> SEQUENCE: 35	
ggaagatctc agaaaggtag acag	24
<210> SEQ ID NO 36	
<211> LENGTH: 81	
<212> TYPE: DNA	
<213> ORGANISM: Bacteriophage T4	
<400> SEQUENCE: 36	
ggctatattc cggaagcgcc gcgatgatggc caggcgatatg tgcgtaaaga tggcgaatgg	60
gtgctgctgt ctacctttct g	81

- What is claimed is:
1. An immunogenic composition for induction of an immune response against a lentivirus, said composition comprising a fusion protein, said fusion protein consisting of the following, in the order from the N-terminus to the C-terminus:
    - a lentivirus gp41 N-terminal heptad repeat (NHR or HR1) region;
    - a foldon trimerization motif; and
    - an immunoenhancer sequence.
  2. The immunogenic composition of claim 1 wherein the lentivirus is selected from the group consisting of HIV-1, HIV-2, and SIV.
  3. The immunogenic composition of claim 1 wherein the immunoenhancer is selected from the group consisting of the Fc domain of immunoglobulin G, complement component C3d, and Onchocerca volvulus activation associated protein-1 (Ov-ASP-1).
  4. The immunogenic composition of claim 3 wherein the Fc domain of immunoglobulin G or complement component C3d is from a mammal selected from the group consisting of mouse, rabbit, pig, non-human primate, and human.
  5. The immunogenic composition of claim 1 wherein the composition further comprises an adjuvant.
  6. An immunogenic composition for induction of an immune response against a lentivirus, said composition comprising a fusion protein, said fusion protein consisting of:
    - (a) the following, in the order from the N-terminus to the C-terminus:
      - a lentivirus gp41 N-terminal heptad repeat (NHR or HR1) region,
      - a foldon trimerization motif, and
      - an immunoenhancer sequence; and
    - (b) a His tag or a GST sequence at either the N-terminus or the C-terminus.
  7. A method of inducing an immune response to HIV comprising the steps of  
administering the immunogenic composition of claim 1 to a mammal in need thereof; and  
inducing an immune response in said mammal to said HIV.
  8. The method of claim 7 wherein said immunogenic composition is administered by a route selected from the group consisting of subcutaneous, intramuscular, intraperitoneal, and mucous immunization.
  9. The method of claim 7 wherein said immune response results in the production of neutralizing antibodies against HIV in said mammal.
  10. An immunogenic composition for induction of an immune response against a lentivirus, said composition comprising a fusion protein, said fusion protein consisting of the following, in the order from the N-terminus to the C-terminus:
    - an N46 sequence of human immunodeficiency virus gp41;
    - a foldon trimerization motif; and
    - a human immunoglobulin G Fc sequence.
  11. The immunogenic composition of claim 6 wherein the lentivirus is selected from the group consisting of HIV-1, HIV-2, and SIV.
  12. The immunogenic composition of claim 6 wherein the immunoenhancer is selected from the group consisting of the Fc domain of immunoglobulin G, complement component C3d, and Onchocerca volvulus activation associated protein-1 (Ov-ASP-1).

13. The immunogenic composition of claim 12 wherein the Fc domain of immunoglobulin G or complement component C3d is from a mammal selected from the group consisting of mouse, rabbit, pig, non-human primate, and human.

14. The immunogenic composition of claim 6 wherein the composition further comprises an adjuvant.

15. The method of claim 7 wherein the immunogenic composition is administered in combination with an adjuvant.

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