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(54) ZIKA VIRUS IMMUNOGENIC **COMPOSITIONS**

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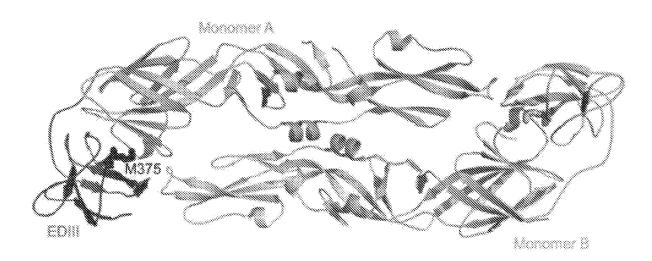
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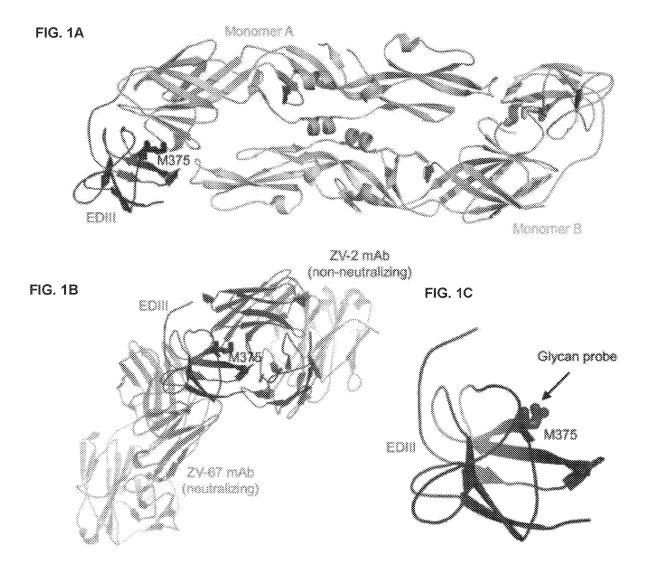
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(57)ABSTRACT

Provided herein are methods of use of subunit immunogenic compositions, in particular for the prevention and treatment of Zika vims infections.

Specification includes a Sequence Listing.





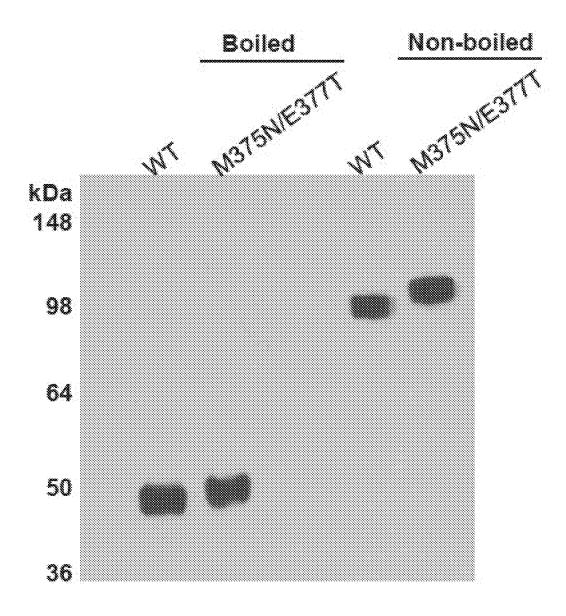
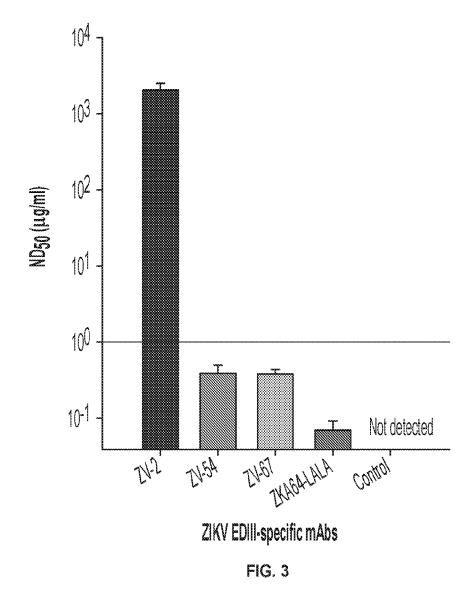
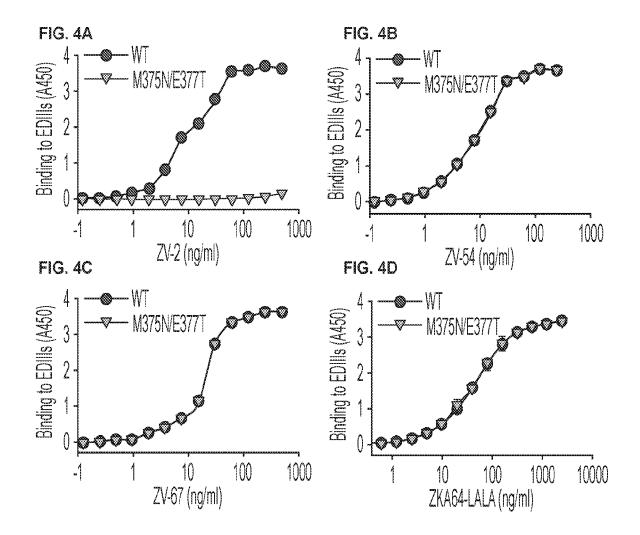


FIG. 2



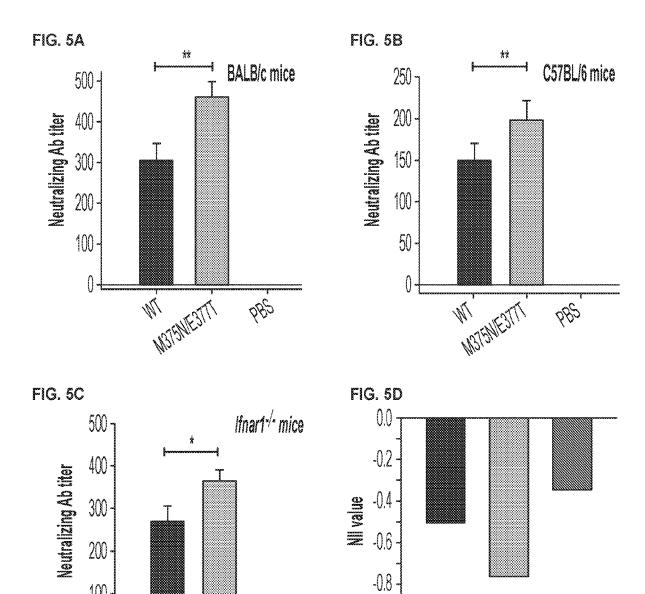


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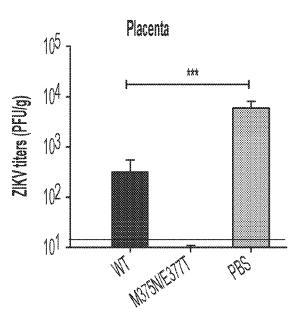
WT NSTEWESTT

685



BALBIC MICE CSTBLIG MICE HABITY' MICE





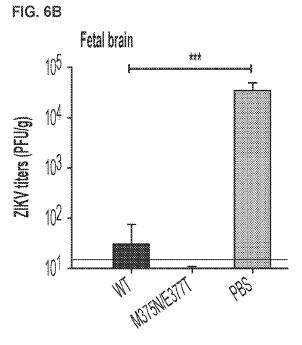
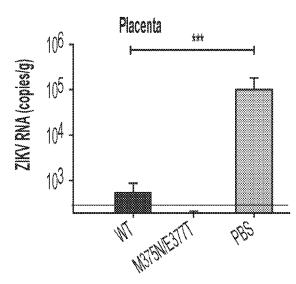
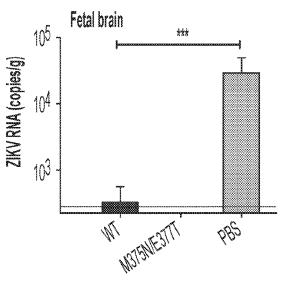


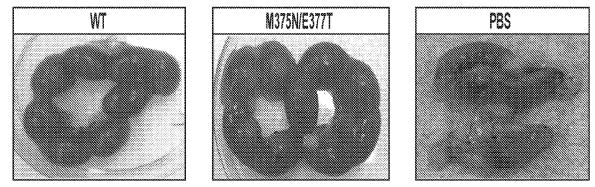


FIG. 6D









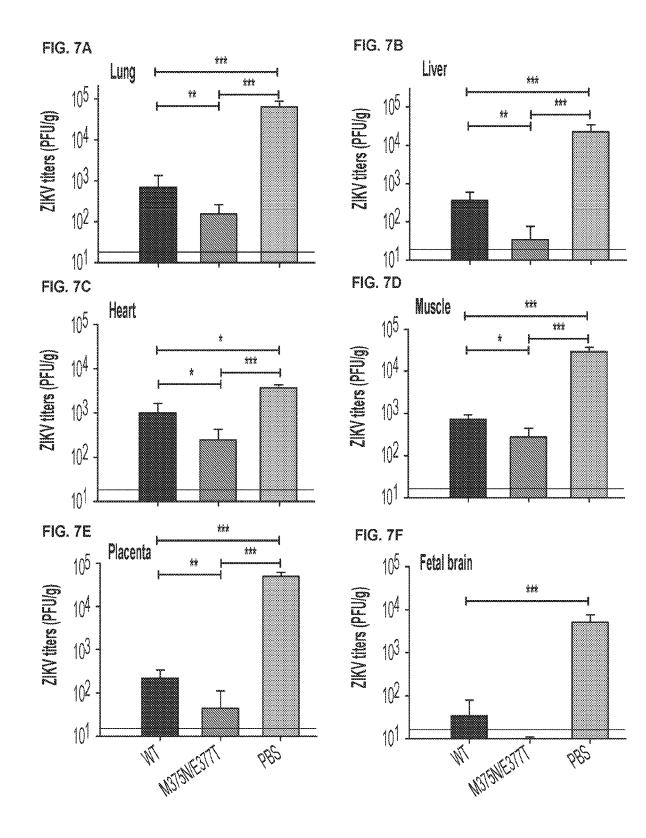
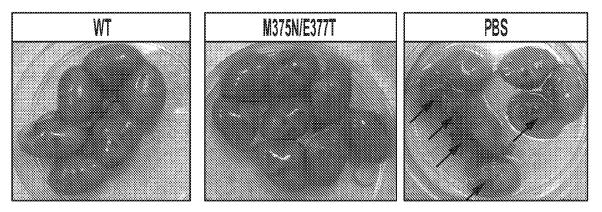
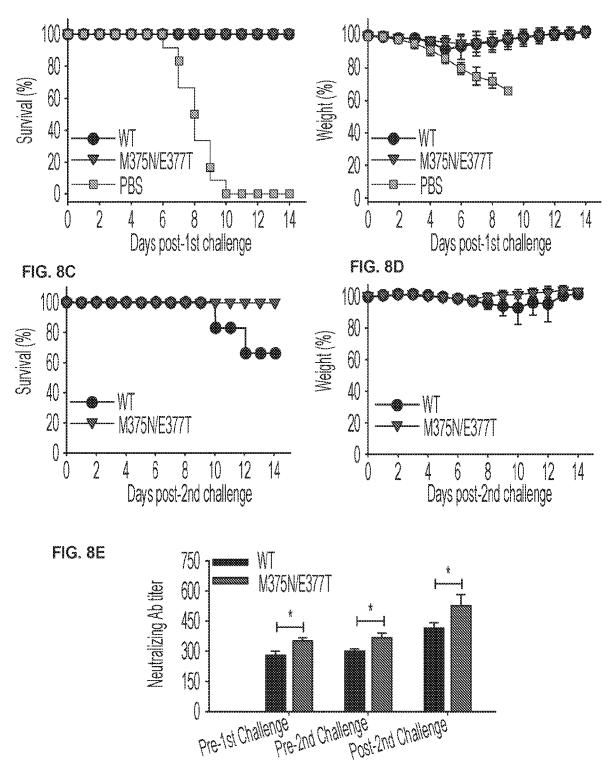


FIG. 7G









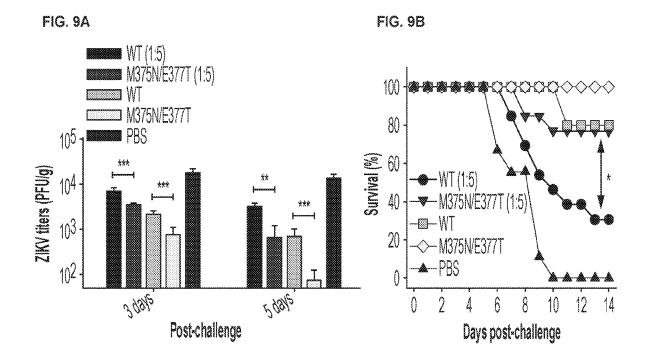
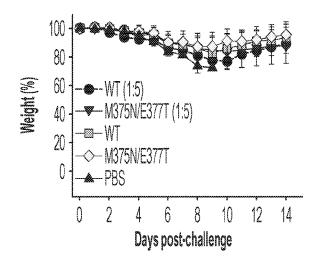
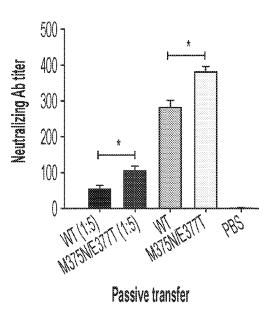
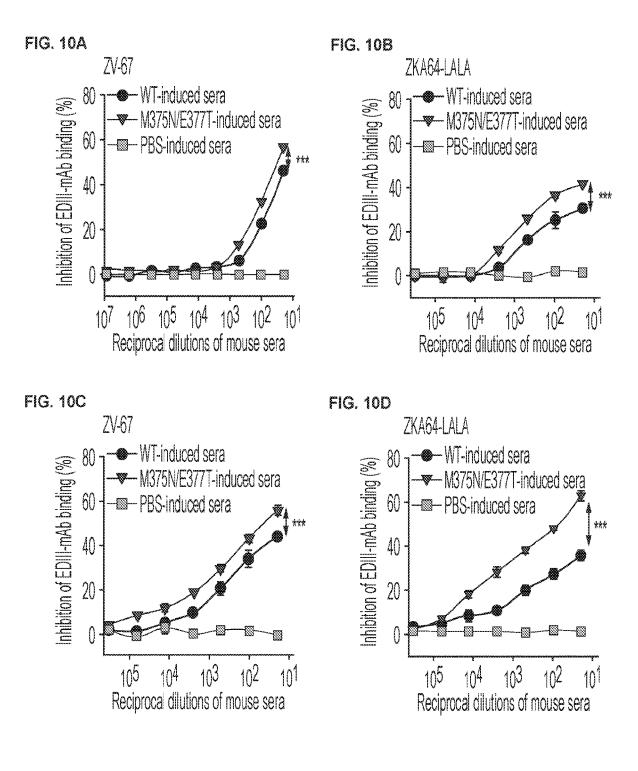


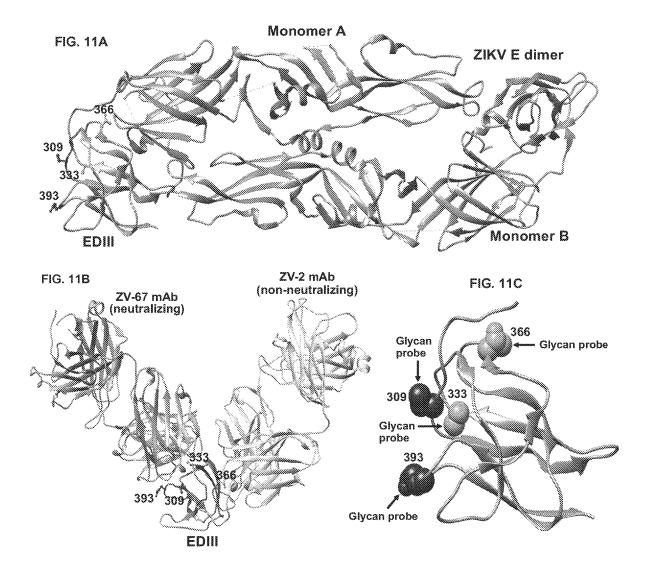
FIG. 9C











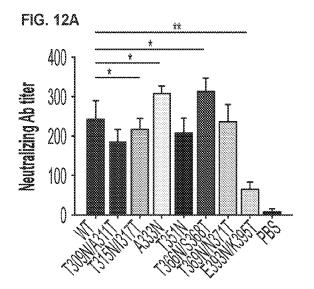


FIG. 12B

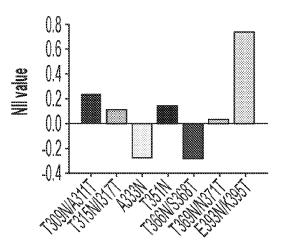
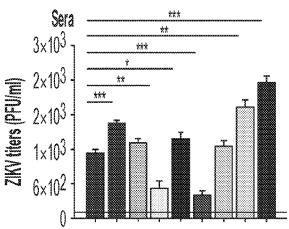
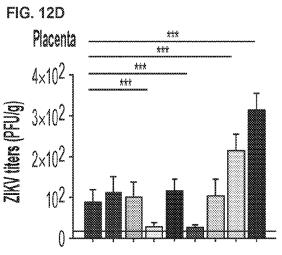
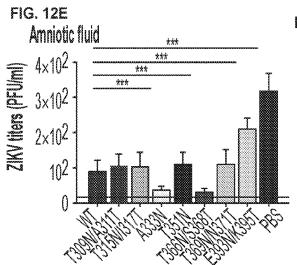
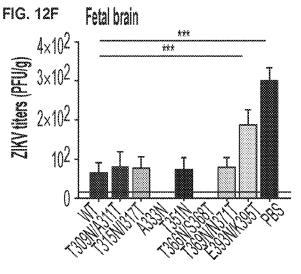


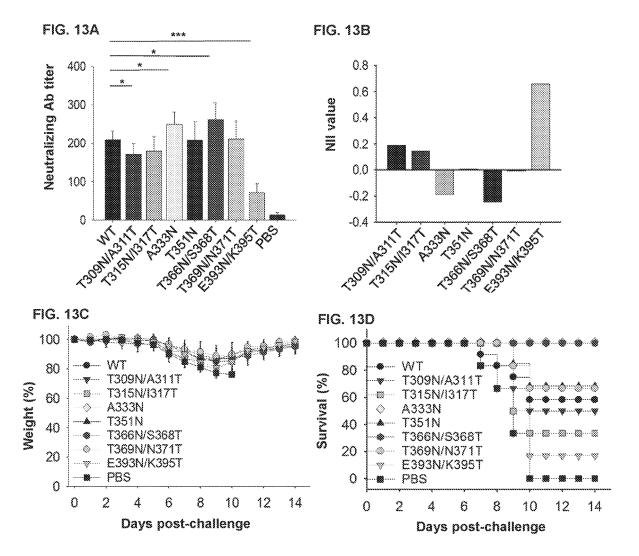
FIG. 12A











ZIKA VIRUS IMMUNOGENIC COMPOSITIONS

CROSS REFERENCE TO RELATED APPLICATIONS

[0001] The present application claims the benefit of U.S. Provisional Patent Application 62/751,470 filed Oct. 26, 2018, which is incorporated by reference herein in its entirety.

FEDERALLY SPONSORED RESEARCH OR DEVELOPMENT

[0002] This invention was made with government support under Grant No. R01A1139092 and R21A1137790 awarded by the National Institutes of Health and Grant No. NYB475, Grant No. NYB486, and Grant No. NYB552 awarded by the New York Blood Center. The government has certain rights in the invention.

TECHNICAL FIELD

[0003] The present disclosure relates to methods of use of subunit immunogenic compositions, in particular for use in the prevention and treatment of Zika virus infections.

BACKGROUND

[0004] Zika virus (ZIKV) is a flavivirus in the same genus of human pathogenic arboviruses, including dengue virus (DENV), West Nile virus (WNV), yellow fever virus (YFV), and Japanese encephalitis virus (JEV). ZIKV causes neurological diseases such as Guillain-Barre syndrome and congenital Zika syndrome (symptoms include microcephaly, brain abnormalities and other congenital malformations). Despite several ZIKV vaccines currently in clinical trials, no vaccines have been approved for preventing ZIKV infections in humans.

[0005] The genome of ZIKV is a single-stranded positivesense RNA encoding a number of structural and nonstructural proteins. The envelope (E) protein, a major structural protein, guides viral entry into host cells by first binding to a host receptor and then fusing viral and host membranes. It forms a viral-envelope-anchored dimer, and each monomer contains multiple structural domains, i.e., domain I (EDI), domain II (EDII), domain III (EDIII), stem region, and transmembrane domain (TM). The E protein is a major inducer of the host immune response and a main target for subunit vaccine design. The individual domains of the E protein may also function in subunit vaccines. Among them, EDI and EDII, but not EDIII, can trigger antibodyenhanced ZIKV entry. Due to its safety, EDIII, which is responsible for binding to host receptor(s), has been our focus for subunit vaccine development. We previously showed that a ZIKV EDIII-based recombinant subunit vaccine is effective in eliciting long-term and broad-spectrum neutralizing antibodies against divergent ZIKV strains. However, its efficacy is not optimal. Improving the efficacy of this subunit vaccine is key to its potential contribution to prevent ZIKV infections.

[0006] Despite their general safety, recombinant subunit vaccines often suffer insufficient efficacy. Using Middle East respiratory syndrome (MERS) coronavirus (MERS-CoV) as a model system, we have determined that when a recombinant subunit vaccine is made, the previously buried surface areas become artificially exposed and they contain immu-

nodominant non-neutralizing epitopes that distract the host immune system from reacting to neutralizing epitopes. We further developed a neutralizing immunogenicity index (NII) to quantitatively characterize the contribution of each of these epitopes to the overall immunogenicity of the subunit vaccine. In spite of its initial success in MERS-CoV subunit vaccine designs, the concepts of the intrinsic limitation of viral subunit vaccines and the NII have not been extended to subunit vaccine designs targeting other viruses.

SUMMARY

[0007] Provided herein are methods of use of subunit immunogenic compositions, in particular for use in the prevention and treatment of Zika virus infections.

[0008] Disclosed herein are methods for preventing and/or treating a Zika virurs infection in a subject in need thereof, the method comprising administering a therapeutically effective amount of an isolated polypeptide comprising an altered structural envelope (E) protein domain III (EDIII), wherein the alteration reduces or prevents the immunogenicity of one or more interfering epitopes of the EDW. In some embodiments, the method further comprises co-administration of an adjuvant.

[0009] In some embodiments, the interfering epitope surrounds residue 333, 366, or 375 of ZIKV EDIII polypeptide. In some embodiments, the alteration comprises creation of a site for N-linked glycosylation. In some embodiments, the N-linked glycosylation site is an NXT or NXS sequence, wherein X is any amino acid. In some embodiments, the alteration comprises at least an N at residue 375 and either a T or S at residue 377. In some embodiments, the alteration comprises at least an N at residue 333 and either a T or S at residue 335. In some embodiments, the alteration comprises at least an N at residue 366 and either a T or S at residue 368. In some embodiments, the alteration comprises M375N/ E377T. In some embodiments, the alteration comprises A333N. In some embodiments, the alteration comprises T366N/S368T. In some embodiments, the administration decreases viral titer levels.

[0010] In some embodiments of the methods disclosed herein, the administration decreases viral RNA copy number. In some embodiments, the administration increases production of neutralizing antibodies.

[0011] In some embodiments of the methods disclosed herein, the adjuvant comprises liposomes, virosomes, saponins, emulsions, or combinations thereof. In some embodiments, the adjuvant comprises aluminum, cholesterol, adjuvant system 04 (ASO4), adjuvant system 01E (AS01E), monophosphoryl lipid A (MPL), saponin, oil-inwater, or combinations thereof. In some embodiments, the adjuvant comprises aluminum, MPL, or combinations thereof. In some embodiments, the adjuvant comprises aluminum.

[0012] Also disclosed is an immunogenic composition comprising an isolated polypeptide comprising any of SEQ ID NOs 4-6 or 8-22, or a combination thereof. Also disclosed herein is an immunogenic composition comprising an isolated polypeptide consisting of any of SEQ ID NOs 4-6 and 8-22, or a combination thereof. In some embodiments, the immunogenic composition further comprises an adjuvant.

[0013] Also disclosed herein are methods for preventing and/or treating a Zika virus infection in a subject in need

thereof, the method comprising administering a therapeutically effective amount of an immunogenic composition disclosed herein.

[0014] Also disclosed herein are methods for preventing birth defects associated with a Zika virus infection, comprising immunizing a woman who is pregnant, who may become pregnant, or who plans to become pregnant, with an immunogenic composition disclosed herein.

[0015] In some embodiments of the methods disclosed herein, the administration increases production of neutralizing antibodies. In some embodiments, the administration decreases viral titer levels. In some embodiments, the administration decreases viral RNA copy number. In some embodiments, as a result of the administration, any pregnancy in the woman does not result in Zika virus-associated birth defects.

[0016] Also disclosed herein is the use of an immunogenic composition disclosed herein in the prevention and/or treatment of a Zika virus infection.

[0017] Also disclosed herein is the use of an immunogenic composition disclosed herein in the prevention of birth defects associated with Zika virus infection.

[0018] The details of one or more embodiments of the invention are set forth in the accompanying drawings and the description below. Other features, objects, and advantages of the invention will be apparent from the description and drawings, and from the claims.

DESCRIPTION OF DRAWINGS

[0019] FIG. 1A-C shows an exemplary structure-based design of ZIKV EDIII vaccine with enhanced efficacy. FIG. 1A depicts the crystal structure of ZIKV E protein dimer (PDB ID: 5LBV). The two monomeric subunits are shown on top and bottom, respectively. EDIII of monomeric subunit A is shown on left bottom, and residue 375 of EDIII is shown in sticks. FIG. 1B depicts the crystal structures of ZIKV EDIII (shown in the middle) complexed with ZIKV EDIII-specific monoclonal antibodies (mAbs). Neutralizing mAb ZV-67 is shown at bottom (PDB ID: 5KVG), and non-neutralizing mAb ZV-2 is shown on top (PDB ID: 5KVD). FIG. 1C depicts the introduction of an N-linked glycosylation site to residue 375 of ZIKV EDIII (after introducing double mutations M375N/E377T to EDIII).

[0020] FIG. 2 depcits exemplary Western blot detection of the ZIKV EDIII mutant (M375N/E377T) protein vaccine with residue 375 shielded by a glycan probe. ZIKV wildtype (WT) EDIII was included as a control. Boiled and non-boiled protein samples are shown. The protein molecular weight marker (kDa) is indicated on the left. ZIKV E-specific mouse polyclonal antibody (1:1,000) was used for test.

[0021] FIG. 3 depicts exemplary neutralizing activity of ZIKV EDIII-specific mAbs against ZIKV infection. Mouse mAbs ZV-2 and ZV-54, as well as human mAbs ZV-67 and ZKA64-LALA, were tested for neutralizing activity against ZIKV strain R103451 (2015/Honduras) by plaque reduction neutralization test (PRNT). PBS was included as control. Neutralizing activity is expressed as mean 50% neutralization dose (ND₅₀) ±standard error (s.e.m) (n=2).

[0022] FIG. **4**A-D depicts exemplary antigenicity of the ZIKV EDIII mutant (M375N/E377T) protein vaccine. ELISA was performed to test the binding of mutant M375N/E377T protein to ZIKV EDIII-specific mAbs ZV-2 (FIG. **4**A), ZV-54 (FIG. **4**B), ZV-67 (FIG. **4**C), and ZKA64-LALA

(FIG. 4D) mAbs. ZIKV EDIII-WT protein was used as a comparison. The data are presented as means \pm s.e.m (n=4). [0023] FIG. 5A-D depicts exemplary neutralizing antibodies induced by ZIKV EDIII mutant (M375N/E377T) protein vaccine against ZIKV infection. Immunocompetent BALB/c (FIG. 5A) and C57BL/6 (FIG. 5B) mice, as well as Type-I interferon (i.e., interferon-a/p) receptor (IFNAR)-deficient (Ifnar1^{-/-}) mice (C57BL/6 background) (FIG. 5C), were immunized with ZIKV WT and mutant (M375N/E377T) EDIII proteins, and sera were collected at 10 days post 2^{nd} immunization to detect neutralizing antibodies against ZIKV (strain R103451) by PRNT assay. PBS was used as background control. Neutralizing activity is expressed as 50% plaque reduction neutralizing antibody titer (PRNT₅₀), which was calculated through serum dilutions and neutralizations at 50% plaque reduction using the CalcuSyn computer program. The data are presented as means±s.e.m (n=6). Significant differences (*: P<0.05; **: P<0.01) are shown in FIG. 5A-C. FIG. 5D depicts the calculated neutralizing immunogenicity index (NII) for mutant M375N/ E377T protein using serum neutralizing antibody titers (PRNT₅₀) from BALB/c, C57BL/6, and Ifnar1^{-/-} mice described above. The NII value was calculated using (PRNT_{50-WT}-PRNT_{50-muntant})/PRNT_{50-WT}, where PRNT₅₀₋ wT and PRNT_{50-mutant} represent PRNT₅₀ neutralizing anti-body titers for VVT and mutant (M375N/E377T) EDIII proteins, respectively.

[0024] FIG. 6A-E depicts exemplary enhanced efficacy of the ZIKV EDIII mutant (M375N/E377T) protein vaccine in ZIKV-challenged immunocompetent pregnant BALB/c mice and their fetuses. Female BALB/c mice were immunized with WT and mutant (M375N/E377T) EDIII proteins, or PBS control, and mated with male BALB/c mice for pregnancy 10 days post-2nd immunization. The pregnant mice (E10-E13) were challenged with ZIKV (strain R103451, 2×10⁵ plaque-forming units: PFU), and detected for viral titers and viral RNAs in placenta (FIGS. 6A and 6C) and fetal brain (FIGS. 6B and 6D) by plaque (FIG. 6A-B) and qRT-PCR (FIG. 6C-D) assays 6 days after challenge. The data are presented as means±s.e.m (n=6). Significant differences (***: P<0.001) are shown in each figure. The detection limit for plaque assay was 20 PFU/g, and for qRT-PCR was 2.5×10^2 RNA copies/g, of tissue. FIG. 6E depicts the morphology of uteri and embryos in immunized pregnant BALB/c mice 6 days after ZIKV challenge.

[0025] FIG. 7A-G depicts exemplary inhibition of ZIKV replication of ZIKV EDIII mutant (M375N/E377T) protein vaccine against ZIKV challenge in pregnant Ifnar1^{-/-} mice and their fetuses. Ifnar1^{-/-} mice were immunized with VVT and mutant (M375N/E377T) EDIII proteins, or PBS control, and mated with male Ifnar $1^{-/-}$ mice for pregnancy 10 days post- 2^{nd} immunization. The pregnant mice (E10-E13) were challenged with ZIKV (strain R103451, 10³ PFU), and viral titers were measured by plaque assay in tissues, including lung (FIG. 7A), liver (FIG. 7B), heart (FIG. 7C), muscle (FIG. 7D), placenta (FIG. 7E), and fetal brain (FIG. 7F) 6 days post-challenge. The data are presented as means±s.e.m (n=6). Significant differences (*: P<0.05; **: P<0.01; ***: P<0.001) are shown in each figure. The detection limit for lung and heart was 25 PFU/g, and for liver, muscle, placenta, and fetal brain was 20 PFU/g, of tissue. FIG. 7G depicts the morphology of the uteri and embryos of immunized pregnant mice 6 days after ZIKV challenge as Ifnar1⁻ described above. Arrows indicate fetal death.

[0026] FIG. 8A-E depicts exemplary enhanced protective efficacy of ZIKV EDIII mutant (M375N/E377T) protein vaccine in ZIKV-challenged lethal Ifnar1^{-/-} mouse model. Adult male Ifnar1-/- mice were immunized with ZIKV WT and mutant (M375N/E377T) EDIII proteins, or PBS control, and challenged with ZIKV (R103451, 10³ PFU) 10 days post- 2^{nd} immunization to evaluate survival (FIG. 8A) and weight (FIG. 8B) for 14 days (n=6). The surviving Ifnar1^{-/-} mice in the WT and mutant M375N/E377T groups were further challenged with ZIKV (R103451, 5×10⁴ PFU), and observed for survival (FIG. 8C) and weight (FIG. 8D) as above (n=6). The % weight in FIG. 8B and FIG. 8D represents the mean % weight of all surviving mice at the indicated days after challenge, and error bars indicate s.e.m. FIG. 8E depicts serum neutralizing antibody titers of mice before and after challenge with ZIKV (strain R103451) by PRNT assay. The data are presented as mean PRNT₅₀±s.e.m (n=4-6). Significant differences (*: P<0.05) are shown in the figure.

[0027] FIG. 9A-D depicts exemplary enhanced protective efficacy of passively transferred sera from ZIKV EDIII mutant (M375N/E377T) protein vaccine-immunized mice against ZIKV challenge. Ifnar1^{-/-} mice received passively transferred M375N/E377T-immunized mouse sera, and 6 h later, they were challenged with ZIKV (strain R103451), followed by evaluation of serum viral titers at 3 and 5 days post-challenge, as well as survival and weight changes for 14 days. Mouse sera from ZIKV EDIII-WT- or PBS-immunized mice were included as controls. FIG. 9A depicts ZIKV titers in sera of challenged mice collected at 3 and 5 days after ZIKV challenge (n=5). Significant differences (**: P<0.01; ***: P<0.001) are shown. Survival (FIG. 9B) and weight (FIG. 9C) of serum transferred mice after ZIKV challenge (n=5-13). Significant difference (*: P<0.05) is shown between the VVT and mutant EDIII (M375N/E377T) groups (1:5 dilution). FIG. 9D depicts neutralizing antibodies induced by the above sera transferred into mice against ZIKV (strain R103451) by PRNT assay. Neutralizing activity is expressed as PRNT₅₀. The data are presented as means±s.e.m. Significant differences (*: P<0.05) are shown.

[0028] FIG. 10A-D depicts exemplary masking of a nonneutralizing epitope on residue 375 of ZIKV EDIII refocused neutralizing immunogenicity on neutralizing epitopes. An ELISA competition assay was performed to test the inhibition of the binding between human neutralizing mAbs ZV-67 or ZKA64-LALA and WT EDIII protein by mutant (M375N/E377T) EDIII-induced mouse sera. Inhibition of the binding between ZV-67 (FIG. 10A, 40C) or ZKA64-LALA (FIG. 10B, 10D) mAb and VVT EDIII protein by mutant M375N/E377T-induced sera from BALB/c (FIG. 10A, 10B) and Ifnar1^{-/-} (FIG. 10C, 10D) mice. The EDIII-WT-induced mouse sera were used as a comparison. Mouse sera induced by PBS were used as negative control. The inhibition in mAb-EDIII binding was calculated in the presence or absence of immunized sera. The data are presented as means±s.e.m. (n=4), and significant differences (***: P<0.001) are shown in each figure.

[0029] FIG. **11**A-C depicts an exemplary structure-based design of ZIKV EDIII mutant (A333N and T366N/S368T) protein vaccines with enhanced efficacy. FIG. **11**A depicts the crystal structure of ZIKV E protein dimer (PDB ID: SLBV). The two monomeric subunits (monomer A and B) are on top and bottom, respectively. EDIII of monomeric subunit A is shown on left bottom, and the four residues with

increased (A333 and T366) or decreased (T309 and E393) immunogenicity are shown in sticks. FIG. **11**B depicts the crystal structures of ZIKV EDIII (shown at bottom) complexed with ZIKV EDIII-specific mAbs. Neutralizing mAb ZV-67 (PDB ID: SKVG) and non-neutralizing mAb ZV-2 (PDB ID: SKVD) are shown on left and right, respectively. FIG. **11**C depicts the introduction of an N-linked glycosylation site at residues T309, A333, T366, and E393 of ZIKV EDIII (after introducing single or double mutations T309N/A311T, A333N, T366N/S368T, and E393N/K395T to EDIII).

[0030] FIG. 12A-F depicts exemplary neutralizing antibodies and protection induced by ZIKV EDIII mutant proteins (i.e., T309N/A311T, T315N/1317T, A333N, T351N, T366N/S368T, T369N/N371T, and E393N/K395T) in immunocompetent BALB/c mice and their fetuses. FIG. 12A depicts improved neutralizing activity of A333N and T366N/S368T ZIKV EDIII mutant proteins in BALB/c mice. Female mice were immunized with ZIKV WT and mutant EDIII proteins, and sera collected at 10 days post-last dose were measured for neutralizing antibodies against ZIKV strain PAN2016 (2016/Panama) by PRNT assay. PBS was used as background control. Neutralizing activity is expressed as PRNT₅₀. The data are presented as means \pm s. e.m (n=5). Significant differences (*: P<0.05; **: P<0.01) are shown. FIG. 12B depicts calculated neutralizing immunogenicity index (NII) for ZIKV EDIII mutant proteins based on serum neutralizing antibody titer (PRNT $_{50}$) using formula (PRNT_{50-WT}-PRNT_{50-muntant})/PRNT_{50-WT}, where PRNT_{50-WT} and PRNT_{50-mutant} represent PRNT₅₀ neutralizing antibody titers for WT and mutant EDIII proteins, respectively. (FIG. 12C-F) Demonstrate enhanced efficacy of A333N and T366N/S368T ZIKV EDIII mutant proteins in protecting pregnant BALB/c mice and their fetuses against high-dose ZIKV (strain PAN2016, 5×10^4 PFU) challenge. The immunized BALB/c mice were mated with male BALB/c mice and the pregnant mice (E10-E13) were challenged with ZIKV after injection with anti-Ifnar1 antibody, followed by detection of viral titers by plaque assay in sera (FIG. 12C) at 3 days, as well as in placenta (FIG. 12D), amniotic fluid (FIG. 12E), and fetal brain (FIG. 12F) at 5 days, post-challenge. Significant differences (*: P<0.05; **: P<0.01; ***: P<0.001) are shown in FIG. 12C-F, and the data are presented as means±s.e.m (n=5). The detection limit was 25 PFU/ml of sera and amniotic fluid, or 25 PFU/g of tissue (placenta and fetal brain).

[0031] FIG. 13A-D depicts exemplary neutralizing antibodies and protection induced by ZIKV EDIII mutant proteins (i.e., T309N/A311T, T315N/1317T, A333N, T351N, T366N/S368T, T369N/N371T, and E393N/K395T) in immunocompromised Ifnar1-/- mice. FIG. 13A depictsimproved neutralizing activity of A333N and T366N/S368T ZIKV EDIII mutant proteins in Ifnar1-/- mice. Male and female mice were immunized with ZIKV WT and mutant EDIII proteins, and sera collected at 10 days post last-dose were measured for neutralizing antibodies against ZIKV (strain R103451) by PRNT assay. PBS was used as background control. Neutralizing activity is expressed as PRNT₅₀. The data are presented as means \pm s.e.m (n=6). Significant differences (*: P<0.05; ***: P<0.001) are shown. FIG. 13B depicts the neutralizing immunogenicity index (NII) for ZIKV EDIII mutant proteins based on serum neutralizing antibody titer (PRNT₅₀), as calculated above. FIG. 13C-D depict enhanced efficacy of A333N and T366N/

S368T ZIKV EDIII mutant proteins in protecting ZIKVchallenged lethal Ifnar1^{-/-} mouse model. The immunized mice were challenged with ZIKV (strain R103451, 10³ PFU) 13 days post-last dose to evaluate weight (FIG. 13C) and survival (FIG. 13D) for 14 days (n=6). The % weight in FIG. 13C represents the mean % weight of all survived mice at the indicated days after challenge, and error bars indicate s.e.m.

DETAILED DESCRIPTION

[0032] A number of embodiments of the disclosed subunit immunogenic compositions have been described. Nevertheless, it will be understood that various modifications may be made without departing from the spirit and scope of the present disclosure.

[0033] As used herein an "immunogenic composition" refers to an expressed protein, with or without an adjuvant, and which elicits an immune response in the host. The immunogenic compositions disclosed herein are immunoprotective or therapeutic. When the immunogenic compositions may prevent, ameliorate, palliate, or eliminate disease from the host then the immunogenic composition may also optionally be referred to as a vaccine. In some embodiments, the immunogenic composition includes one or more pharmaceutically acceptable excipients and may optionally include an adjuvant.

[0034] Zika virus (ZIKV) infection in pregnant women can lead to fetal death and malformations. A previously reported ZIKV envelope protein domain III (EDIII) has been shown as a possible subunit vaccine candidate with crossneutralization but limited efficacy.

[0035] Provided herein are methods of use of subunit immunogenic compositions, such as vaccines, in particular for the prevention and treatment of ZIKV infection. In some embodiments, to improve its efficacy and overcome subunit vaccines' intrinsic limitations, a non-neutralizing epitope on ZIKV EDIII was identified surrounding residue 375 that is buried in the full-length envelope protein but becomes exposed in recombinant EDIII, and this residue was shielded with an engineered glycan probe. Further epitopes with similar properties were identified surrounding residues 333 or 366 of ZIKV EDIII protein. These three epitopes are thus referred to as interfering epitopes.

[0036] In various embodiments, EDIII is altered to reduce or prevent the immunogenicity of these three interfering epitopes, so that a higher titer antibody response to neutralizing epitopes is obtained upon immunization with the altered EDIII domains, as compared to that obtained upon immunization with the wild type EDIII. In particular embodiments, this is accomplished by mutating the sequence of the EDIII polypeptide sequence to create a glycosylation site in the vicinity of one or more of these interfering epitopes to obscure, and prevent binding of immunoglobulins to, the peptide epitope. In various embodiments, any one, two, or all three of the interfering epitopes are altered.

[0037] Sites for N-linked glycosylation are the amino acid sequences NXT or NXS (and rarely NXC), where X is any naturally encoded amino acid besides proline. In some embodiments, the EDIII polypeptide comprises an N at residue 375 and a T at residue 377, for example, M375N/E377T. In some embodiments, the EDIII polypeptide comprises an N at residue 333 and a T at residue 335, for example A333N/T335T (that is, residue 335 is T in the wild

type sequence used in the examples below). In some embodiments, the EDIII polypeptide comprises an N at residue 366 and a T at residue 368, for example, T366N/S368T. In other embodiments, one or more of these T residues are instead S.

[0038] In some embodiments, compared to the wild-type EDIII, the mutant EDIII proteins induce significantly stronger neutralizing antibodies in different mouse strains (i.e., BALB/c, C57BL/6, and Ifnar1^{-/-}), and also demonstrate significantly improved efficacy in fully protecting mice, particularly pregnant mice and their fetuses, against high-dose lethal ZIKV challenge. In some embodiments, the mutant EDIII immune sera also significantly enhanced the passive protective efficacy in fully protecting mice against lethal ZIKV challenge and the passive protection was positively associated with neutralizing antibody titers. In some embodiments, the enhanced efficacy of the mutant EDIII proteins was due to the shielding of the immunodominant non-neutralizing epitope, which led to immune refocusing on the neutralizing epitopes.

[0039] The altered EDIII proteins and fusion proteins are most often produced by recombinant expression in a glyco-sylation competent expression system, preferably a mammalian expression system, or one in which the glycosylation would not itself be immunogenic in the subjects to be immunized. In alternative embodiments, the EDIII protein is produced by chemical synthesis.

[0040] In some embodiments, the altered EDIII polypeptide is produced as a complete protein. In some embodiments, the altered EDIII polypeptide is produced to further comprise a peptide tag, such as a poly-histidine tag. In some embodiments, the altered EDIII polypeptide is produced as a fusion protein. In some embodiments, the EDIII polypeptide is fused to one or more of an Fc domain, hemagglutinin (HA), protein A, foldon, GCN4 trimerization motif, or glutathione S-transferase (GST). Such fusion proteins may also comprise a peptide tag, such as a poly-his tag.

[0041] Some embodiments comprise one or more isolated immunogenic polypeptides comprising an altered EDIII polypeptide as described herein. Other embodiments are immunogenic compositions or vaccines comprising one or more such polypeptides. Some embodiments are methods to prevent or treat Zika viral infection by administering such immunogenic polypeptides or compositions, or vaccines, to a person infected with, exposed to, or at risk of exposure to Zika virus. Other embodiments are methods to prevent or reduce the severity of birth defects associated with Zika virus infection by administering, Some embodiments are methods to prevent or treat Zika viral infection by administering such immunogenic polypeptides or compositions, or vaccines, to a woman who is pregnant, who may become pregnant, or who plans to become pregnant. In particular embodiments the isolated immunogenic polypeptide comprising an altered EDIII polypeptide is any one of SEQ ID NOs 4-6 and 8-22.

[0042] Some embodiments may be expressed using functional language, for example, means for inducing Zika virus neutralizing epitope-focused immune responses, or means for inducing anti-Zika virus immune responses with reduced reactivity to non-neutralizing (or interfering) epitopes. Examples of such means include SEQ ID NOs 4-6 and 8-22. Similarly, in some embodiments methods of treatment include a step for inducing Zika virus neutralizing epitopefocused immune responses, and the like, corresponding to administration of the herein disclosed immunogenic polypeptides.

	TABLE 1
	Sequence Identifiers
SEQ NO:	ZIKV pre-M and full-length envelope (E) protein (containing amino acids for ZIKV pre-M and Membrane protein (M), and E protein with 505 amino acids) VTRRGSAYYMYLDRNDAGEAISFPTTLGMNKCYIQIMDLGHMCDATMSYECPM LDEGVEPDDVDCWCNTTSTWVVGTCHHKKGEARRSRAVTLPSHSTRKLQTR SQTWLESREYTKHLIRVENWIFRNPGFALAAAAIAWLLGSSTSQKVIYLVMIL LIAPAYSIRCIGVSNRDFVEGMSGGTWVDIVLEHGGCVTVMAQDKPTVDIELV TTTVSNMAEVRSYCYEASISDMASDSRCPTQGEAYLDKQSDTQYVCKRTLVDR GWGNGCGLFGKGSLVTCAKFACSKKMTGKSIQPENLEYRIMLSVHGSQHSGMI VNDTCHETDENRAKVEITPNSPRAEATLGGFGSLGLDCEPRTGLDFSDLYYLT MNNKHWLVHKEWFHDIPLPWHAGADTGTPHWNNKEALVEFKDAHAKGVSYSLCTAA FTFTKIPAETLHGTVTVEVQYAGTDGPCKVPAQMAVDMQTLTPVGRLITANPV ITESTENSKMMLELDPPFGDSYIVIGVGEKKITHHWHRSGSTIGKAFEATVRG AKRMAVLGDTAWDFGSVGGALNSLGKGIHQIFGAAFKSLFGGMSWFSQILIGT LLMWLGLNTKNGSISLMCLALGGVLIFLSTAVSADVGCSV
SEQ NO:	ZIKV full-length E protein (containing ZIKV full-length E protein with amino acids 1-505) IRCIGVSNRDFVEGMSGGTWDIVLEHGGCVTVMAQDKPTVDIELVTTTVSNM AEVRSYCYEASISDMASDSRCPTQGEAYLDKQSDTQYVCKRTLVDRGWGNGCG LFGKGSLVTCAKFACSKKMTGKSIQPENLEYRIMLSVHGSQHSGMIVNDTGHE TDENRAKVEITPNSPRAEATLGGFGSLGLDCEPRTGLDFSDLYYLTMNNKHWL VHKEWFHDIPLPWHAGADTGTPHNNNKEALVEFKDAHAKRQTVVVLGSQEGAV HTALAGALEAEMDGAKGRLSSGHLKCRLKMDKLRLKGVSYSLCTAAFTFYLIP AETLHGTVTVEVQYAGTDGPCKVPAQMAVDMQTLTPVGRLITANPVITESTEN SKMMLELDPPFGDSYIVIGYGEKKITHHWHRSGSTIGKAFEATVRGAKRMAVL GDTAWDFGSVGGALNSLGKGIHQIFGAAFKSLFGGMSWFSQILIGTLLMWLGL NTKNGSISLMCLALGGVLIFLSTAVSAD
SEQ NO:	ZIKV E protein domain III (EDIII) wild-type (WT) (containing ZIKV E protein amino acids 298-409) LRLKGVSYSLCTAAFTFTKIPAETLHGTVTVEVQYAGTDGPCKVPAQMAVDMQ TLTPVGRLITANPVITESTENSKMMLELDPPFGDSYIVIGVGEKKITHHWHRS GSTIGK
SEQ NO:	ZIKV EDIII M375N/E377T mutant protein (containing ZIKV E protein amino acids 298-409 with M375N and E377T mutations) LRLKGVSYSLCTAAFTFTKIPAETLHGTVTVEVQYAGTDGPCKVPAQMAVDMQ TLTPVGRLITANPVITESTENSKM <u>NLT</u> LDPPFGDSYIVIGVGEKKITHHWHRS GSTIGK
SEQ NO:	ZIKV EDIII WT-Fc protein (containing ZIKV E wild-type protein amino acids 298- 409 with human Fc (hFc) sequences) LRLKGVSYSLCTAAFTFTKIPAETLHGTVTVEVQYAGTDGPCKVPAQMAVDMQ TLTPVGRLITANPVITESTENSKMMLELDPPFGDSYIVIGVGEKKITHHWHRS GSTIGKRSDKTHTCPPCPAPELLGGPSVFLFPPKPKDTLMISRTPEVTCVVD VSHEDPEVKFNWYVDGVEVHNAKTKPREEQYNSTYRVVSVLTVLHQDWLNGKE YKCKVSNKALPAPIEKTISKAKGQPREPQVYTLPPSREEMTKNQVSLTCLVKG FYPSDIAVEWESNGQPENNYKTTPPVLDSDGSFFLYSKLTVDKSRWQQGNVFS CSVMHEALHNHYTQKSLSLSPGK
SEQ NO:	ZIKV EDIII M375N/E377T-Fc mutant protein (containing ZIKV E wild-type protein amino acids 298-409 with M375N and E377T mutations, and hFc sequences) LRLKGVSYSLCTAAFTFTKIPAETLHGTVTVEVQYAGTDGPCKVPAQMAVDMQ TLTPVGRLITANPVITESTENSKMMLTLDPPFGDSYIVIGYGEKKITHHWHRS GSTIGKRSDKTHTCPPCPAPELLGGPSVFLFPPKPKDTLMISRTPEVTCVVVD VSHEDPEVKFNWYVDGVEVHNAKTKPREEQYNSTYRVVSVLTVLHQDWLNGKE YKCKVSNKALPAPIEKTISKAKGQPREPQVYTLPPSREEMTKNQVSLTCLVKG FYPSDIAVEWESNGQPENNYKTTPPVLDSDGSFFLYSKLTVDKSRWQQGNVFS CSVMHEALHNHYTQKSLSLSPGK
SEQ NO:	ZIKV EDIII WT-His protein (containing ZIKV E wild-type protein amino acids 298-409 and His ₆) LRLKGVSYSLCTAAFTFTKIPAETLHGTVTVEVQYAGTDGPCKVPAQMAVDMQ TLTPVGRLITANPVITESTENSKMMLELDPPFGDSYIVIGVGEKKITHHWHRS GSTIGKHHHHHH
SEQ NO:	ZIKV EDIII M375N/E377T-His mutant protein (containing ZIKV E wild-type protein amino acids 298-409 with M375N and E377T mutations, and His ₆) LRLKGVSYSLCTAAFTFTKIPAETLHGTVTVEVQYAGTDGPCKVPAQMAVDMQ

LRLKGVSYSLCTAAFTFTKIPAETLHGTVTVEVQYAGTDGPCKVPAQMAVDMQ TLTPVGRLITANPVITESTENSKM<u>NLT</u>LDPPFGDSYIVIGVGEKKITHHWHRS GSTIGKHHHHHH TABLE 1-continued

	Sequence Identifiers
SEQ ID NO: 9	ZIKV EDIII A333N mutant protein (containing ZIKV E protein amino acids 298- 409 with A333N mutation) LRLKGVSYSLCTAAFTFTKIPAETLHGTVTVEVQYNGTDGPCKVPAQMAVDMQ TLTPVGRLITANPVITESTENSKMMLELDPPFGDSYIVIGVGEKKITHHWHRS GSTIGK
SEQ ID NO: 10	ZIKV EDIII T366N/S368T mutant protein (containing ZIKV E protein amino acids 298-409 with T366N and S368T mutations) LRLKGVSYSLCTAAFTFTKIPAETLHGTVTVEVQYAGTDGPCKVPAQMAVDMQ TLTPVGRLITANPVI <u>NET</u> TENSKMMLELDPPFGDSYIVIGVGEKKITHHWHRS GSTIGK
SEQ ID NO: 11	ZIKV EDIII A333N-Fc mutant protein (containing ZIKV E protein amino acids 298-409 with A333N mutation, and hFc sequences) LRLKGVSYSLCTAAFTFTKIPAETLHGTVTVEVQY <u>N</u> GTDGPCKVPAQMAVDMQ TLTPVGRLITANPVITESTENSKMMLELDPPFGDSYIVIGVGEKKITHHWHRS GSTIGKRSDKTHTCPPCPAPELLGGPSVFLFPPKPKDTLMISRTPEVTCVVVD VSHEDPEVKFNWYVDGVEVHNAKTKPREEQYNSTYRVVSVLTVLHQDWLNGKE YKCKVSNKALPAPIEKTISKARGQPREPQVTLPPSREEMTKNQVSLTCLVKG FYPSDIAVEWESNGQPENNYKTTPPVLDSDGSFFLYSKLTVDKSRWQQGNVFS CSVMHEALHNHYTQKSLSLSPGK
SEQ ID NO: 12	ZIKV EDIII T366N/S368T-Fc mutant protein (containing ZIKV E protein amino acids 298-409 with T366N and S368T mutations, and hFc sequences) LRLKGVSYSLCTAAFTFTKIPAETLHGTVTVEVQYAGTDGPCKVPAQMAVDMQ TLTPVGRLITANPVINETENSKMMLELDPPFGDSYIVIGVGEKKITHHWHRS GSTIGKRSDKTHTCPPCPAFELLGGPSVFLFPPKPKDTLMISRTPEVTCVVVD VSHEDPEVKFNWYVDGVEVHNAKTKPREEQVNSTYRVVSVLTVLHQDWLNGKE YKCKVSNKALPAPIEKTISKAKGQPREPQVYTLPPSREEMTKNQVSLTCLVKG FYPSDIAVEWESNGQPENNYKTTPPVLDSDGSFFLYSKLTVDKSRWQQGNVFS CSVMHEALHNHYTQKSLSLSPGK
SEQ ID NO: 13	ZIKV EDIII T309N/A311T mutant protein (containing ZIKV E protein amino acids 298-409 with T309N and A311T mutations) LRLKGVSYSLC <u>NAT</u> FTFTKIPAETLHGTVTVEVQYAGTDGPCKVPAQMAVDMQ TLTPVGRLITANPVITESTENSKMMLELDPPFGDSYIVIGVGEKKITHHWHRS GSTIGK
SEQ ID NO: 14	ZIKV EDIII T315N/I317T mutant protein (containing ZIKV E protein amino acids 298-409 with T315N and I317T mutations) LRLKGVSYSLCTAAFTF <u>NKT</u> PAETLHGTVTVEVQYAGTDGPCKVPAQMAVDMQ TLTPVGRLITANPVITESTENSKMMLELDPPFGDSYIVIGVGEKKITHHWHRS GSTIGK
SEQ ID NO: 15	ZIKV EDIII T351N mutant protein (containing ZIKV E protein amino acids 298- 409 with T351N mutation) LRLKGVSYSLCTAAFTFTKIPAETLHGTVTVEVQYAGTDGPCKVPAQMAVDMQ <u>M</u> LTPVGRLITANPVITESTENSKMMLELDPPFGDSYIVIGVGEKKITHHWHRS GSTIGK
SEQ ID NO: 16	ZIKV EDIII T369N/N371T mutant protein (containing ZIKV E protein amino acids 298-409 with T369N and N371T mutations) LRLKGVSYSLCTAAFTFTKIPAETLHGTVTVEVQYAGTDGPCKVPAQMAVDMQ TLTPVGRLITANPVITES <u>NET</u> SKMMLELDPPFGDSYIVIGVGEKKITHHWHRS GSTIGK
SEQ ID NO: 17	ZIKV EDIII E393N/K395T mutant protein (containing ZIKV E protein amino acids 298-409 with E393N and K395T mutations) LRLKGVSYSLCTAAFTFTKIPAETLHGTVTVEVQYAGTDGPCKVPAQMAVDMQ TLTPVGRLITANPVITESTENSKMMLELDPPFGDSYIVIGVG <u>NKT</u> ITHHWHRS GSTIGK
SEQ ID NO: 18	ZIKV EDIII T309N/A311T-Fc mutant protein (containing ZIKV E protein amino acids 298-409 with T309N and A311T mutations, and hFc sequences) LRLKGVSYSLC <u>NATF</u> TFTKIPAETLHGTVTVEVQYAGTDGPCKVPAQMAVDMQ TLTPVGRLITANPVITESTENSKMMLELDPPFGDSYIVIGVGEKKITHHWHRS GSTIGKRSDKTHTCPPCPAFELLGGPSVFLPPPKPKDTLMISRTPEVTCVVD VSHEDPEVKFNWYVDGVEVHNAKTKPREEQYNSTYRVVSVLTVLHQDWLNGKE YKCKVSNKALPAPIEKTISKARGQPREPQVYTLPPSREEMTKNQVSLTCLVKG FYPSDIAVEWESNGQPENNYKTTPPVLDSDGSFFLYSKLTVDKSRWQQGNVFS CSVMHEALHNHYTQKSLSLSPGK
SEQ ID NO: 19	ZIKV EDIII T315N/I317T-Fc mutant protein (containing ZIKV E protein amino acids 298-409 with T315N and I317T mutations, and hFc sequences)

6

TABLE 1-continued

	TABLE 1-continued
	Sequence Identifiers
	LRLKGVSYSLCTAAFTF <u>M</u> K <u>T</u> PAETLHGTVTVEVQYAGTDGPCKVPAQMAVDMQ TLTPVGRLITANPVITESTENSKMMLELDPPFGDSYIVIGVGEKKITHHWHRS GSTIGKRSDKTHTCPPCPAPELLGGPSVFLFPPKPKDTLMISRTPEVTCVVVD VSHEDPEVKFNWYVDGVEVHNAKTKPREEQYNSTYRVVSVLTVLHQDWLNGKE YKCKVSNKALPAPIEKTISKAKGQPREPQVYTLPPSREEMTKNQVSLTCLVKG FYPSDIAVEWESNGQPENNYKTTPPVLDSDGSFFLYSKLTVDKSRWQQGNVFS CSVMHEALHNHYTQKSLSLSPGK
SEQ ID NO: 20	ZIKV EDIII T351N-Fc mutant protein (containing ZIKV E protein amino acids 298-409 with T351N mutation, and hFc sequences) LRLKGVSYSLCTAAFTFTKIPAETLHGTVTVEVQYAGTDGPCKVPAQMAVDMQ <u>NLTPVGRLITANPVITESTENSKMMLELDPPFGDSYIVIGVGEKKITHHWHRS</u> GSTIGKRSDKTHTCPPCPAPELLGGPSVFLFPPKPKDTLMISRTPEVTCVVD VSHEDPEVKFNWYVDGVEVHNAKTKPREEQYNSTYRVVSVLTVLHQDWLNGKE YKCKVSNKALPAPIEKTISKAKGQPREPQVYTLPPSREEMTKNQVSLTCLVKG FYPSDIAVEWESNGQPENNYKTTPPVLDSDGSFFLYSKLTVDKSRWQQGNVFS CSVMHEALHNHYTQKSLSLSPGK
SEQ ID NO: 21	ZIKV EDIII T369N/N371T-Fc mutant protein (containing ZIKV E protein amino acids 298-409 with T369N and N371T mutations, and hFc sequences) LRLKGVSYSLCTAAFTFTKIPAETLHGTVTVEVQYAGTDGPCKVPAQMAVDMQ TLTPVGRLITANPVITES <u>NET</u> SKMMLELDPPFGDSYIVIGVGEKKITHHWHRS GSTIGKRSDKTHTCPPCPAPELLGGPSVFLPPPKPKDTLMISRTPEVTCVVD VSHEDPEVKFNWYVDGVEVHNAKTKPREEQYNSTYRVVSVLTVLHQDWLNGKE YKCKVSNKALPAPIEKTISKAKGQPREPQVTLPPSREEMTKNQVSLTCLVKG FYPSDIAVEWESNGQPENNYKTTPPVLDSDGSFFLYSKLTVDKSRWQQGNVFS CSVMHEALHNHYTQKSLSLSPGK
SEQ ID NO: 22	ZIKV EDIII E393N/K395T-Fc mutant protein (containing ZIKV E protein amino acids 298-409 with E393N and K395T mutations, and hFc sequences) LRLKGVSYSLCTAAFTFTKIPAETLHGTVTVEVQYAGTDGPCKVPAQMAVDMQ TLTPVGRLITANPVITESTENSKMMLELDPPFGDSYIVIGVG <u>MKT</u> ITHHWHRS GSTIGKRSDKTHTCPPCPAPELLGGPSVFLFPPKPKDTLMISRTPEVTCVVD VSHEDPEVKFNWYVDGVEVHNAKTKPREEQYNSTYRVVSVLTVLHQDWLNGKE YKCKVSNKALPAPIEKTISKAKGQPREPQVYTLPPSREEMTKNQVSLTCLVKG FYPSDIAVEWESNGQPENNYKTTPPVLDSDGSFFLYSKLTVDKSRWQQGNVFS CSVMHEALHNHYTQKSLSLSPGK
SEQ ID NO: 23	Human Fc (hFc) sequence RSDKTHTCPPCPAPELLGGPSVFLFPPKPKDTLMISRTPEVTCVVVDVSHEDP EVKFNWYVDGVEVHNAKTKPREEQYNSTYRVVSVLTVLHQDWLNGKEYKCKVS NKALPAPIEKTISKAKGQPREPQVYTLPPSREEMTKNQVSLTCLVKGFYPSDI AVEWESNGQPENNYKTTPPVLDSDGSFFLYSKLTVDKSRWQQGNVFSCSVMHE ALHNHYTQKSLSLSPGK
SEQ ID NO: 24	Foldon GYIPEAPRDGQAYVRKDGEWVLLSTFL
SEQ ID NO: 25	GCN4 MKQIEDKIEEILSKIYHIENEIARIKKLIGEV

EXAMPLES

Example 1—Construction, Expression and Immunogenicity of Recombinant ZIKV EDIII Mutant (M375N/E377T) Protein

[0043] Materials and Methods

[0044] Expression and purification of recombinant proteins. ZIKV EDIII protein containing residues 298-409 (i.e., wild-type, WT) was constructed based on ZIKV E (ZikaSPH2015 strain, GenBank accession number KU321639.1) fused with a C-terminal human IgG1 Fc (human Fc) (SEQ. ID NO:23) tag. Mutant ZIKV EDIII protein containing a glycan probe surrounding residue 375 (i.e., M375N/E377T) was constructed by multi-site mutagenesis kits using the EDIII-WT plasmid as template. The recombinant proteins were expressed in 293T cell culture supernatants, and purified by Protein A affinity chromatography. **[0045]** Mouse immunization. The purified ZIKV EDIII VVT (SEQ ID NO:5) and M375N/E377T (SEQ ID NO:6) mutant proteins were used to immunize mice. The 4-6-week-old female BALB/c and C57BL/6, as well as 3-week-old female Ifnar1^{-/-} mice, were intramuscularly (i.m.) immunized with EDIII WT and mutant (M375N/E377T) proteins (10 µg/mouse), or PBS control, in the presence of aluminum (500 µg/mouse) and monophosphoryl lipid A (MPL) (10 µg/mouse) adjuvant combinations. The immunized mice were boosted once with the same immunogens at 4-week intervals, and sera were collected 10 days after last immunization to test neutralizing antibodies.

[0046] Western blot. The purified proteins were analyzed by Western blot. The proteins were separated by 10% Tris-Glycine SDS-PAGE gels, and transferred to nitrocellulose membranes. The transferred blot was blocked with 5% fat-free milk in PBST overnight at 4° C., and sequentially incubated with ZIKV E-protein-immunized mouse sera

(1:1000) and horseradish peroxidase (HRP)-conjugated goat anti-mouse IgG (1:5,000) for 1 h at room temperature. The signal was visualized with ECL Western blot substrate buffer and Amersham Hyperfilm.

[0047] ELISA. ELISA was used to test the binding between ZIKV EDIII proteins and EDIII-specific mouse mAbs ZV-2 and ZV-54, and human mAbs ZV-67 and ZKA64-LALA. ELISA plates were coated with EDIII WT or M375N/E377T mutant proteins overnight at 4° C. and blocked with 2% fat-free milk in PBST for 2 h at 37° C. After three washes, the plates were sequentially incubated with serially diluted mAbs and HRP-conjugated anti-mouse IgG-Fab antibody (for ZV-2 and ZV-54, 1:5,000) or anti-human IgG-Fab antibody (for ZV-67 and ZKA64-LALA, 1:5,000) for 1 h at 37° C. The reaction was detected by substrate 3,3',5,5'-tetramethylbenzidine and stopped with stop solution (1 N H₂SO₄). Absorbance at OD₄₅₀ nm was measured.

[0048] ZIKV plaque-forming assay and plaque reduction neutralization test (PRNT) assay. ZIKV human strain R103451 (2015/Honduras) was amplified in Vero E6 cells to determine viral titers by a standard plaque-forming assay. Viral titers in the tissues and/or sera from the challenged mice were detected using similar approaches as described above. A PRNT was carried out to measure neutralizing antibody titers in immunized mouse sera or EDIII-specific mAbs. ZIKV (100 PFU) was incubated with serially diluted sera or mAbs for 1.5 h at 37° C., and the antibody-virus mixtures were then added to Vero E6 cells and incubated for 1 h at 37° C. The cells were overlaid with medium (1% carboxymethyl cellulose in DMEM containing 2% FBS), and cultured at 37° C. for 4-5 days, followed by staining with 0.5% crystal violet. PRNT₅₀ or ND₅₀ was calculated as the highest dilution of sera or mAbs leading to complete inhibition of viral infectivity in at least 50% of the wells.

[0049] Results

[0050] Identification and Masking of a Non-Neutralizing Epitope on ZIKV EDIII WT Protein

[0051] To identify an immunodominant non-neutralizing epitope on ZIKV EDIII, we analyzed the crystal structure of ZIKV E protein dimer. We found a patch of surface area on EDIII that is buried in the full-length E protein dimer, but becomes exposed in recombinant EDIII. Met375 is located in the center of this patch and protrudes from a bent β -strand (FIG. 1A-B). Hence, we engineered a glycan probe onto the epitope surrounding residue 375 (i.e., epitope-375) (FIG. 1C). To this end, we introduced double mutations M375N/ E377T into EDIII, which changed residue 375 to an N-linked glycosylation site. The ZIKV EDIII mutant (M375N/E377T) protein, along with the EDIII wild-type (WT), was expressed and purified from transfected mammalian 293T cell culture supernatant. Both proteins contain a C-terminal Fc tag and were purified to homogeneity. The result from Western blot showed that both proteins ran as dimers without boiling (due to the Fc dimeric tag and incomplete denaturing), and as monomers after boiling (due to complete denaturing). The EDIII M375N/E377T mutant protein ran slower with a slightly larger molecular weight than the EDIII WT (FIG. 2), indicating that the EDIII mutant was glycosylated after addition of a glycan probe at residue 375.

[0052] To characterize the conformation of the ZIKV EDIII mutant protein, we investigated the binding interactions between ZIKV EDIII (WT or M375N/E377T mutant)

and ZIKV EDIII-specific mAbs using ELISA. Among the mAbs used in this study, ZV-54, ZV-67, and ZKA64-LALA can potently neutralize ZIKV infections in vitro, whereas ZV-2 has no neutralizing activity in vitro in a plaque-based neutralization assay (FIG. 3). The results showed that the ZIKV EDIII mutant (M375N/E377T) protein bound to ZV-54, ZV-67, and ZKA64-LALA neutralizing mAbs, but did not bind to ZV-2. In comparison, the ZIKV EDIII WT protein bound to all of the four mAbs (FIG. 4A-D). We then analyzed the binding sites of these mAbs on EDIII. The crystal structures of ZIKV E protein complexed with ZV-67 and of ZIKV EDIII complexed with ZV-2 revealed that ZV-67 binds to an area that is exposed on the E protein dimer and is likely the receptor-binding site, whereas ZV-2 binds to an area centering epitope-375 that is buried on the E protein dimer but becomes artificially exposed on the recombinant EDIII (FIG. 1B). The structural basis for the binding of ZV-54 and ZKA64-LALA are not known, but these two mAbs may also bind functionally important regions on EDIII and away from epitope-375. Overall, these data further confirm that the ZIKV EDIII mutant (M375N/ E377T) protein has successfully incorporated a glycan at residue 375. The data also reveal that despite the introduced glycan, the EDIII mutant (M375N/E377T) protein retains its native structural conformation and antigenicity by binding to at least three different neutralizing mAbs.

[0053] ZIKV EDIII Epitope-375 had Significantly Improved Neutralizing Activity with Negative Neutralizing Immunogenicity Index

[0054] We previously defined neutralizing immunogenicity index (NII) of an epitope as the contribution of the epitope to the overall neutralizing immunogenicity of the vaccine (Du L. et al., Nat Commun 7:13473, 2016). For epitope-375 on ZIKV EDIII, its NII can be calculated as difference between the neutralizing immunogenicity of the EDIII wild-type (WT) and that of the EDIII mutant (M375N/E377T), divided by the neutralizing immunogenicity of the EDIII WT. To measure the NII of epitope-375 on ZIKV EDIII, we immunized mice with the EDIII WT and mutant (M375N/E377T) proteins individually, and measured the induced serum neutralizing antibody titers. The mice included immunocompetent BALB/c and C57BL/6 mice and immunocompromised Ifnar $1^{-/-}$ mice. The results showed that compared to the EDIII WT, the EDIII mutant (M375N/E377T) protein induced significantly higher titers of anti-ZIKV neutralizing antibodies in all immunized mice (FIG. 5A-C), suggesting that epitope-375 makes negative contribution to the overall neutralizing immunogenicity of ZIKV EDIII vaccine and that masking epitope-375 increases the overall neutralizing immunogenicity of the vaccine. We further calculated the NII of epitope-375 on EDIII. The value of NII varies slightly in different mice: about -0.50 in BALB/c, -0.76 in C57BL/6, and -0.34 in Ifnar1^{-/-} mice (FIG. 5D). The negative sign of NII confirms that the contribution of epitope-375 to the overall neutralizing immunogenicity of the vaccine is negative, and the value of NII suggests that masking epitope-375 increases the overall neutralizing immunogenicity of the vaccine by about 50%, 76%, and 34% (calculated using formula: (PRNT_{50-WT}-PRNT_{50-mutant})/PRNT_{50-WT}*100), respectively, in BALB/c, C57BL/6, and Ifnar1^{-/-} mice. Therefore, the measured NII values reveal that epitope-375 is an immunodominant nonneutralizing epitope and that masking it can significantly improve the neutralizing immunogenicity of the ZIKV EDIII vaccine.

Example 2—Protective Efficacy of ZIKV EDIII Mutant (M375N/E377T) Protein Subunit Vaccine in Mouse Models

[0055] Materials and Methods

[0056] ZIKV challenge studies and protection evaluation. Ten days after last immunization, female BALB/c and Ifnar1-/- mice immunized with ZIKV EDIII WT (SEQ ID NO:5) and M375N/E377T mutant (SEQ ID NO:6) proteins were mated with respective naïve male mice within the same strains. Mice injected with PBS were included as controls. Pregnant female mice (E10-E13) were intraperitoneally (i.p.) challenged with ZIKV strain R103451 (10^3 PFU for Ifnar1^{-/-} mice, and 2×10^5 PFU for BALB/c mice; 200 µl/mouse). ZIKV viral titers and RNA copies in tissues, placentas and fetal brain collected 6 days post-infection (p.i.) were detected by plaque-forming assay and quantitative reverse transcriptase PCR (qRT-PCR) assay, respectively. In a separate experiment, male Ifnar1^{-/-} mice were (i.p.) challenged with ZIKV (R103451, 10³ PFU) 10 days post-last immunization, and evaluated for survival and weight for 14 days p.i. Ten days after completion of the 1st challenge experiment, the surviving Ifnar1^{-/-} mice in the EDIII WT and mutant (M375N/E377T) groups were further challenged with high-titer ZIKV (R103451, 5×10⁴ PFU), and observed for survival and weight as described above. Mice with ≥25% body weight loss were humanely euthanized.

[0057] qRT-PCR. ZIKV RNA copies in sera and tissues of challenged mice were detected by qRT-PCR. RNAs were extracted by QIAamp MinElute Virus Spin Kit (for sera) and RNeasy Mini Kit (for tissues), and quantified by one-step qRT-PCR using Power SYBR Green PCR Master Mix, MultiScribe Reverse Transcriptase, and Ambion[™] RNase Inhibitor in ViiA 7 Master Cycler PCR System. The forward and reverse primers 5'-TTGGTCATGATACTGCTGAT-TGC-3' (SEQ ID NO:26) and 5'-CCTTCCACAAAGTCCC-TATTGC-3' (SEQ ID NO:27) were used for the amplification.

[0058] Results

[0059] Enhanced Efficacy of the ZIKV EDIII Mutant (M375N/E377T) Protein in Protecting Immunized Mice and their Fetuses

[0060] The efficacy of the ZIKV EDIII mutant (M375N/ E377T) protein vaccine in protecting immunocompetent pregnant female mice and their fetuses from ZIKV infection was investigated, as immunocompetent adult mice, such as BALB/c, are non-lethal models for ZIKV infection. To this end, we immunized BALB/c mice with either the ZIKV EDIII wild-type (WT) or mutant (M375N/E377T) protein vaccine, and challenged the pregnant mice (E10-E13) with ZIKV (strain R103451, 2×10^5 PFU). We then collected placenta and fetal brain 6 days after challenge, and measured the viral titers and RNA copies of ZIKV of these samples using plaque and qRT-PCR assays, respectively. Uteri and embryos were collected simultaneously to observe morphological changes of the uteri and compare fetal status. The results from plaque assay revealed that different from the ZIKV EDIII-WT protein-treated group, viral titers in the placenta and fetal brain of ZIKV EDIII mutant (M375N/ E377T) protein-immunized pregnant mice were undetectable (FIG. 6A-B). Similarly, the results from qRT-PCR assay demonstrated that the viral RNA copies in the placenta and fetal brain of the ZIKV EDIII mutant (M375N/E377T) protein-immunized pregnant mice were also undetectable (FIG. 6C-D). As a negative control, the tissues of PBStreated pregnant mice contained significantly higher viral titers and RNA copies (FIG. 6A-D). In addition, there were no significant morphological changes of the uteri of ZIKV EDIII-WT and mutant (M375N/E377T) protein-immunized pregnant mice and all embryos from these mice were in good conditions, whereas the embryos from the PBS-treated mice were invisible, showing severe resorption in their uteri (FIG. 6E). Collectively, compared to the ZIKV EDIII WT protein, the EDIII mutant (M375N/E377T) protein vaccine protects immunocompetent pregnant mice more effectively from ZIKV infection as evidenced by undetectable viral titers and RNA copies, although both the EDIII WT and mutant vaccines protected the fetuses of immunocompetent pregnant mice from ZIKV infection.

[0061] Next, the efficacy of the ZIKV EDIII mutant (M375N/E377T) protein vaccine in protecting immunocompromised Ifnar1^{-/-} mice and their fetuses from ZIKV infection was evaluated, as these mice are lethal models for ZIKV infection. To this end, we immunized female Ifnar1^{-/-} mice and male Ifnar^{-/-} mice with either the ZIKV EDIII WT or mutant (M375N/E377T) vaccine, and then treated female and male mice differently. First, the immunized male and female mice were mated and pregnant female Ifnar1^{-/-} mice were challenged with ZIKV (strain R103451, 10³ PFU), the tissues were collected (including placenta and fetal brain) 6 days post-challenge, ZIKV titers were measured in these tissues using plaque assay. We also examined the uteri and embryos of these mice 6 days post-challenge. The results showed that compared to the ZIKV EDIII-WT proteintreated group, the viral titers in the ZIKV EDIII mutant (M375N/E377T) protein-immunized female Ifnar1^{-/-} pregnant mice were significantly lower in lung (FIG. 7A), liver (FIG. 7B), heart (FIG. 7C), muscle (FIG. D), and placenta (FIG. 7E), or undetectable in fetal brain (FIG. 7F). Moreover, there were no significant morphological changes of the uteri and all fetuses were in good conditions in ZIKV EDIII WT- and mutant (M375N/E377T) protein-treated mice (FIG. 7G). In the negative control group (i.e., PBS-treated female pregnant Ifnar $1^{-/-}$ mice), the viral titers were very high in all the tested tissues and fetal brain, resulting in significant morphological changes of the uteri with severe resorption or fetal death (FIG. 7A-G).

[0062] Second, we challenged adult male Ifnar1^{-/-} mice sequentially with ZIKV at low (R103451, 10³ PFU) and high (R103451, 5×10^4 PFU) doses, and observed them for survival and weight changes. The results showed that the male Ifnar1^{-/-} mice immunized with either the ZIKV EDIII WT or the mutant (M375N/E377T) protein vaccine and then challenged with 10³ PFU of ZIKV (strain R103451) all survived at 14 days post 1st challenge (FIG. 8A); their weight decreased slightly during days 4-7 post-infection but kept increasing steadily afterwards (FIG. 8B). In contrast, in the negative control group (i.e., PBS-treated mice), all mice showed ≥25% weight loss and were humanely euthanized at day 10 post-infection. All of the surviving mice immunized with either the ZIKV EDIII WT or mutant (M375N/E377T) vaccine were further challenged with a high dose of ZIKV (strain R103451, 5×10^4 PFU) to investigate whether they can survive lethal high-dose ZIKV infection. The results

showed that the EDIII mutant (M375N/E377T)-immunized mice all survived the second, high-dose ZIKV challenge (100% survival) (FIG. 8C), with no obvious weight loss (FIG. 8D). In contrast, only 67% of the EDIII-WT-immunized mice survived the second, high-dose ZIKV challenge, with considerable weight loss during days 7-12 after ZIKV challenge (FIG. 8C-D). Comparison of the serum neutralizing antibodies in the mice before and after ZIKV challenge demonstrated that anti-ZIKV neutralizing antibody titers were significantly higher in the EDIII mutant (M375N/ E377T)-immunized mice than in the EDIII-WT-immunized mice before the 1^{st} - and 2^{nd} -challenge, particularly after the 2^{nd} -challenge (FIG. 8E). Therefore, the enhanced efficacy of the mutant (M375N/E377T) protein against the high-dose ZIKV infection might be due to the higher serum neutralizing antibodies it induced. Overall, these data demonstrate that compared to the EDIII WT, the EDIII mutant (M375N/ E377T) vaccine protects immunocompromised female pregnant mice and their fetuses more effectively from ZIKV infection, as evidenced by significantly reduced or undetectable viral titers, and it also protects immunocompromised adult male mice from high-dose ZIKV infection more effectively, as evidenced by improved survival rate and steadily maintained weight.

Example 3—Passive Protective Efficacy of the ZIKV EDIII Mutant (M375N/E377T) Protein Subunit Vaccine in ZIKV-Susceptible Lethal Mouse Model

[0063] Materials and Methods

[0064] Passive protection of ZIKV EDIII mutant (M375N/ E377T)-immunized mouse sera against ZIKV challenge in lethal Ifnar1^{-/-} mouse model. The 5-6-week-old male and female Ifnar1^{-/-} mice (n=5-13) were injected (i.p.) with sera (200 μ l/mouse: normalized for equal ZIKV EDIII-specific IgG titers at 10⁵, or 1:5 dilution in PBS) of mice immunized with ZIKV EDIII WT (SEQ ID NO:5) or M375N/E377T mutant (SEQ ID NO:6) proteins. Six hours later, mice were i.p. challenged with ZIKV (strain R103451, 10³ PFU). Mice injected with PBS-induced Ifnar1^{-/-} mouse sera were included as controls. Neutralizing activity of passively transferred mouse sera was detected by PRNT assay. ZIKV titers were detected from sera collected at 3 days and 5 days post-infection (p.i.) by ZIKV plaque-forming assay. Mouse survival and weight were recorded for 14 days p.i.

[0065] Results

[0066] Enhanced Passive Protection of the ZIKV EDIII Mutant (M375N/E377T) Protein-Immunized Mouse Sera in Protecting Lethal Ifnar1^{-/-} Mouse Model Against ZIKV Challenge

[0067] To further elucidate the enhanced protective efficacy of the ZIKV EDIII mutant (M375N/E377T) protein vaccine and investigate the association between protection and neutralizing antibodies, passive protection was performed using the sera of mice immunized with ZIKV EDIII WT and mutant (M375N/E377T) proteins. As such, sera were pooled from each vaccination group, and normalized for equal ZIKV EDIII-specific IgG titers. Adult Ifnar1^{-/-} mice received passively transferred pooled sera (with or without 1:5 dilution), and challenged with ZIKV as described above, followed by comparison of their viremia, survival rate, and weight changes after ZIKV challenge. The results showed that ZIKV viremia in the mice receiving passively transferred EDIII mutant (M375N/E377T)-immunized sera was significantly lower than in the mice receiving passively transferred EDIII-WT-immunized sera at both 3 and 5 days post-challenge (FIG. 9A). In addition, while about 77% of the mice receiving the diluted EDIII mutant (M375N/E377T)-immunized sera survived the ZIKV challenge, all mice receiving the undiluted sera survived the ZIKV challenge, showing slight weight loss (FIG. 9B-C). In contrast, only about 31% and 80% of the mice survived the ZIKV challenge after receiving EDIII-WT-immunized sera with or without dilution at 1:5, respectively, while the former had significant weight loss during days 8-10 after ZIKV challenge (FIG. 9B-C). Moreover, the EDIII-mutant (M375N/E377T)-immunized sera (with or without 1:5 dilution) had significantly higher-titer anti-ZIKV neutralizing antibodies than the EDIII-WT-immunized sera (with or without 1:5 dilution) (FIG. 9D). As expected, mice transferred with the PBS-injected sera, which showed no anti-ZIKV neutralizing antibodies, had the highest viral titers and weight loss, and 0% survival rate at day 10 post-challenge (FIG. 9B-D). The above data indicate that compared to the ZIKV-EDIII-WT-immunized sera, the EDIII-mutant (M375N/E377T)-immunized sera contained significantly high-titer neutralizing antibodies, thus resulting in the enhanced passive protection against ZIKV challenge, suggesting their positive association.

> Example 4—Molecular Mechanism for the Enhanced Efficacy of the ZIKV EDIII Mutant (M375N/E377T) Protein Subunit Vaccine

[0068] Materials and Methods

[0069] The competition between ZIKV EDIII-specific human mAbs ZV-67 or ZKA64-LALA and EDIII mutant (M375N/E377T) protein-induced mouse sera for the binding of ZIKV EDIII wild-type (WT) protein was performed using ELISA as described above, except that the binding between EDIII VVT and ZV-67 or ZKA64-LALA was tested in the presence of serially diluted mouse sera induced by EDIII WT (SEQ ID NO:5) and M375N/E377T mutant (SEQ ID NO:6) proteins, or PBS, respectively. The binding between EDIII and mAbs was measured by addition of HRP-conjugated anti-human IgG-Fab antibody (1:5,000) and subsequent enzymatic reaction as described above.

[0070] Results

[0071] Molecular Mechanism for the Enhanced Efficacy of the ZIKV EDIII Mutant (M375N/E377T) Protein Subunit Vaccine

[0072] To explore the molecular mechanism for the enhanced efficacy of the ZIKV EDIII mutant (M375N/ E377T) protein vaccine, we investigated the binding interactions between the ZIKV EDIII wild-type (WT) and EDIIIspecific human neutralizing mAbs in the presence of the ZIKV EDIII mutant (M375N/E377T) protein-induced mouse sera using ELISA. The EDIII-WT-induced mouse sera were used as a comparison. The result showed that compared to the EDIII-WT-induced mouse sera, the EDIII mutant (M375N/E377T)-induced sera from both immunocompetent BALB/c mice (FIG. 10A-B) and immunocompromised Ifnar1^{-/-} mice (FIG. 10C-D) blocked the binding between the EDIII VVT protein and EDIII-specific neutralizing mAbs more significantly. Since the ZIKV EDIII VVT and mutant (M375N/E377T) proteins induced similar titers of total serum IgG antibodies, the above results indicate that the ZIKV EDIII mutant (M375N/E377T) protein induced high-titer serum neutralizing antibodies than the EDIII-WT

protein in mice. Therefore, masking the immunodominant non-neutralizing epitope-375 (i.e., NII is negative and has a relatively large absolute value) triggered the host immune system to refocus on less immunodominant but neutralizing epitopes (i.e., NIIs are positive and have relatively small absolute values), accounting for the production of more neutralizing antibodies.

> Example 5—Construction, Expression, Immunogenicity, and Efficacy of Other Recombinant ZIKV EDIII Mutant Proteins

[0073] Materials and Methods

[0074] Expression and purification of recombinant ZIKV EDIII mutant proteins. This was performed as described above. Briefly, mutant ZIKV EDIII proteins containing a glycan probe surrounding residue 309 (i.e., T309N/A311T) (SEQ ID NO:13), residue 315 (i.e., T315N/1317T) (SEQ ID NO:14), residue 333 (i.e., A333N) (SEQ ID NO:9), residue 351 (i.e., T351N) (SEQ ID NO:15), residue 366 (i.e., T366N/S368T) (SEQ ID NO:10), residue 369 (i.e., T369N/ N371T) (SEQ ID NO:16) and residue 393 (i.e., E393N/ K395T) (SEQ ID NO:17) were constructed by multi-site mutagenesis kits using the above ZIKV EDIII wild-type (WT) plasmid fused with a C-terminal human Fc tag as the template. The recombinant proteins were expressed in 293T cell culture supernatants, and purified by Protein A affinity chromatography, as described above.

[0075] Animal immunization. The above purified ZIKV EDIII mutant proteins (SEQ ID NO:11-12 and 18-22) were i.m. injected into female BALB/c (4-6-week-old) or male/ female Ifnar1^{-/-} (4-6-week old) mice (10 μ g/mouse) in the presence of aluminum (500 µg/mouse) and MPL (10 µg/mouse) adjuvant combination. ZIKV EDIII WT protein and PBS were included as controls. The immunized mice were boosted at four weeks, and sera were collected 10 days post-last dose to detect neutralizing antibodies. The immunized mice were subsequently used for the following experiments, including ZIKV challenge and protection evaluation. [0076] ZIKV plaque reduction neutralization test (PRNT). This assay was performed as described above to measure neutralizing antibody titers of immunized mouse sera. Briefly, recent ZIKV human strains PAN2016 or R103451 (100 PFU) were incubated with 2-fold serially diluted mouse sera for 1.5 h at 37° C., which were then added to Vero E6 cells and incubated for 1 h at 37° C. The cells were further overlaid with DMEM containing 1% carboxymethyl cellulose and 2% FBS, and cultured for 4-5 days at 37° C., followed by staining with 0.5% crystal violet, and plaques counted. PRNT₅₀ was calculated at 50% plaque reduction using the CalcuSyn computer program, as described above. [0077] ZIKV challenge and evaluation of protection against ZIKV infection. These experiments were carried out as described above. Briefly, ten days post-last dose, the immunized female BALB/c mice were mated with naïve male BALB/c mice. The pregnant BALB/c mice (E10-E13) were pretreated with anti-Ifnar1 blocking antibody (2 mg/mouse) (to become susceptible to ZIKV infection); 24 hours later, the mice were i.p. challenged with ZIKV (strain PAN2016, 5×10⁴ PFU). ZIKV titers in sera (collected at 3 days p.i.), placenta, fetal brain and amniotic fluid (collected at 5 days p.i.) were detected using plaque-forming assay. The ZIKV titers were detected in Vero E6 cells using around 40 µl of sera/amniotic fluid or 40 mg of tissue samples, so the detection limit was about 25 PFU/ml (for sera and amniotic fluid) or 25 PFU/g of tissue (for placenta and fetal brain).

[0078] In a separate experiment, immunized male/female Ifnar1^{-/-} mice were i.p. challenged with ZIKV (strain R103451, 10³ PFU) 13 days post-last dose, and evaluated for survival and weight for 14 days p.i. Mice losing \geq 25% body weight were humanely euthanized.

[0079] Results

[0080] Identification of Other Epitopes (i.e., Residues 333 and 366) on ZIKV EDIII with Significantly Improved Neutralizing Activity and Enhanced Protection of BALB/c mice and their Fetuses

[0081] Similar to epitope-375 (i.e., M375N/E377T mutant) described above, we identified several other epitopes (including 309, 315, 333, 351, 366, 369, and 393) on the ZIKV EDIII wild-type (WT) protein, and engineered a N-linked glycan probe onto each of these epitopes to generate single mutations (i.e., A333N and T351N), or double mutations (T309N/A311T, T315N/1317T, T366N/ S368T, T369N/N371T, and E393N/K395T), which formed seven N-linked glycosylation sites (SEQ ID NO:9-10 and 13-17). An exemplary structure-based design of ZIKV EDIII mutant (A333N and T366N/S368T) protein vaccines with enhanced efficacy is depicted in FIG. 11. Crystal structure of ZIKV E protein dimer (PDB ID: SLBV) is depicted in FIG. 11A. The two monomeric subunits (monomer A and B) are on top and bottom, respectively. EDIII of monomeric subunit A is shown on left bottom, and the four residues with increased (A333 and T366) or decreased (T309 and E393) immunogenicity are shown in sticks. Crystal structures of ZIKV EDIII (shown at bottom) complexed with ZIKV EDIII-specific mAbs are depicted in FIG. 11B. Neutralizing mAb ZV-67 (PDB ID: SKVG) and nonneutralizing mAb ZV-2 (PDB ID: SKVD) are shown on left and right, respectively. FIG. 11C depicts the introduction of an N-linked glycosylation site to residues T309, A333, T366, and E393 of ZIKV EDIII (after introducing single or double mutations T309N/A311T, A333N, T366N/S368T, and E393N/K395T to EDIII).

[0082] We expressed and purified these proteins from 293T cell culture supernatants, and used the mutant, as well as WT (as a control). EDIII proteins to immunize BALB/c mice. We then measured the serum neutralizing antibodies against ZIKV (strain PAN2016), based on which to calculate neutralizing immunogenicity index (NII). The results showed that compared to the EDIII WT, A333N (SEO ID NO:11) and T366N/S368T (SEQ ID NO:12) mutant EDIII proteins induced significantly high-titer neutralizing antibodies, whereas T309N/A311T (SEQ ID NO:18) and E393N/K395T (SEQ ID NO:22) mutant EDIII proteins elicited significantly reduced neutralizing antibodies (FIG. 12A). We further calculated the NII values for each of these epitopes on EDIII (using formula: PRNT_{50-WT}-PRNT₅₀₋ mutant)/PRNT_{50-WT}*100). The results revealed that epitope-333 and epitope-366 had negative values (FIG. 11B), suggesting that these epitopes made negative contributions to the overall neutralizing immunogenicity of ZIKV EDIII vaccine, thus masking them significantly improved vaccine's neutralizing activity. The results also showed that other epitopes, particularly epitope-309 and epitope-393, had positive values (FIG. 12B), suggesting that these epitopes made positive contributions to the overall neutralizing immunogenicity of ZIKV EDIII vaccine, therefore masking them significantly reduced vaccine's neutralizing activity.

[0083] We further investigated the efficacy of these mutant ZIKV EDIII protein vaccines in protecting pregnant BALB/c mice and their fetuses against high-dose ZIKV infection. To this end, the above ZIKV VVT and mutant EDIII-immunized female BALB/c mice were mated with naïve male BALB/mice after last-dose, and the pregnant mice were then challenged with ZIKV (strain PAN2016, 5×10^4 PFU), followed by detection of viral titers in adult mouse sera (3 days after challenge), placenta, amniotic fluid, and fetal brain (5 days after challenge) by plaque assay. The results indicated that compared to the EDIII-WT-proteinimmunized mice, the mutant-EDIII (A333N and T366N/ S368T)-immunized mice had significantly reduced viral titers in their sera, placenta, and amniotic fluid (FIG. 12C-E), or undetectable viral titers in the fetal brain (FIG. 12F). In contrast, while the T309N/A311T (SEQ ID NO:18) and T351N (SEQ ID NO:20)-immunized mice had significantly increased viral titers in their sera (FIG. 12C), the E393N/ K395T (SEQ ID NO:22)-immunized mice had significantly increased viral titers in all samples tested, including sera, placenta, amniotic fluid, and fetal brain (FIG. 12C-F). Moreover, the control PBS-treated mice also had significantly higher viral titers in all above samples (FIG. 12C-F). Therefore, compared to the ZIKV EDIII VVT protein, the A333N (SEQ ID NO:11) and T366N/S368T (SEQ ID NO:12) mutant EDIII proteins had more potent efficacy in protecting immunocompetent pregnant BALB/c mice and their fetuses against high-dose ZIKV challenge.

[0084] Enhanced Neutralizing Activity and Efficacy of ZIKV EDIII Mutant (A333N and T366N/S368T) Proteins in Protecting Immunocompromised Mice Against ZIKV Infection

[0085] We further evaluated the neutralizing immunogenicity and efficacy of the identified A333N and T366N/S368T, as well as other mutant, ZIKV EDIII protein vaccines in protecting ZIKV-susceptible immunocompromised Ifnar1^{-/-} mice. To this end, we immunized Ifnar1^{-/-} mice with each of the ZIKV mutant, or VVT (as a control), EDIII proteins, collected sera to detect neutralizing antibodies and calculate NII values as described above, and then challenged the mice with a ZIKV strain (R103451, 10³PFU) different from that tested in BABL/c mice 13 days post-last dose, followed by investigation of their survival and weight changes.

[0086] The results from neutralization assay showed that compared to the ZIKV EDIII WT, the mutant A333N (SEQ ID NO:11) and T366N/S368T (SEQ ID NO:12) EDIII proteins elicited significantly high-titer neutralizing antibodies against the above ZIKV strain (FIG. 13A), and that the NII values for epitope-333 and epitope-366 were negative in Ifnar1^{-/-} mice (FIG. **13**B). In contrast, the T315N/1317T (SEQ ID NO:19), T309N/A311T (SEQ ID NO:18), and E393N/K395T (SEQ ID NO:22) mutant EDIII proteins induced reduced, or significantly reduced, neutralizing antibody titers (FIG. 13A), and thus the NII values corresponding to the epitope-315, epitope-309, and epitope-393 were positive (FIG. 13B). The above results were generally consistent with those from the immunized BALB/c mice, further confirming that the contribution of epitope-333 and epitope-366 to the overall neutralizing immunogenicity of the ZIKV EDIII vaccine is negative, and thus masking them significantly improved the vaccine's neutralizing immunogenicity.

[0087] The results from challenge study revealed that similar to the EDIII WT-immunized mice, the A333N and T366N/S368T-immunized mice exhibited slightly reduced weight after ZIKV challenge and then increased consistently afterwards (FIG. 13C). Importantly, all mice immunized with either A333N (SEQ ID NO:11) or T366N/S368T (SEQ ID NO:12) mutant EDIII proteins survived, but only about 58% of the mice immunized with EDIII VVT protein survived, at 14 days post-challenge (FIG. 13D), confirming the enhanced efficacy of above two mutant ZIKV EDIII proteins in protecting immunocompromised Ifnar1^{-/-} mice from ZIKV infection. Compared to the EDIII WT-immunized mice, more mice died in the groups immunized with T309N/A311T (SEQ ID NO:18), T315N/1317T (SEQ ID NO:19), and E393N/K395T (SEQ ID NO:22), with survival rates about 50%, 33%, and 17%, respectively, at 14 days after ZIKV challenge (FIG. 13D), indicating the reduced protection of these mutant EDIII proteins with positive NII values. In contrast, mice in the PBS control group exhibited continuously reduced weight, all of which were humanely sacrificed at day 10 after challenge (FIG. 13C-D).

[0088] Overall, the protection results from mice immunized with each mutant or WT ZIKV EDIII protein are consistent with respective serum neutralizing antibody titers, suggesting that EDIII vaccine-induced neutralizing antibodies play an important role in preventing ZIKV infection. This line of experimentation further identified A333N and T366N/S368T mutant ZIKV EDIII proteins as novel subunit vaccines against infection of divergent ZIKV strains.

[0089] Unless otherwise indicated, all numbers expressing quantities of ingredients, properties such as molecular weight, reaction conditions, and so forth used in the specification and claims are to be understood as being modified in all instances by the term "about." As used herein the terms "about" and "approximately" means within 10 to 15%, preferably within 5 to 10%. Accordingly, unless indicated to the contrary, the numerical parameters set forth in the specification and attached claims are approximations that may vary depending upon the desired properties sought to be obtained by the present invention. At the very least, and not as an attempt to limit the application of the doctrine of equivalents to the scope of the claims, each numerical parameter should at least be construed in light of the number of reported significant digits and by applying ordinary rounding techniques. Notwithstanding that the numerical ranges and parameters setting forth the broad scope of the invention are approximations, the numerical values set forth in the specific examples are reported as precisely as possible. Any numerical value, however, inherently contains certain errors necessarily resulting from the standard deviation found in their respective testing measurements.

[0090] The terms "a," "an," "the" and similar referents used in the context of describing the invention (especially in the context of the following claims) are to be construed to cover both the singular and the plural, unless otherwise indicated herein or clearly contradicted by context. Recitation of ranges of values herein is merely intended to serve as a shorthand method of referring individually to each separate value falling within the range. Unless otherwise indicated herein, each individual value is incorporated into the specification as if it were individually recited herein. All methods described herein can be performed in any suitable order unless otherwise indicated herein or otherwise clearly contradicted by context. The use of any and all examples, or exemplary language (e.g., "such as") provided herein is intended merely to better illuminate the invention and does not pose a limitation on the scope of the invention otherwise claimed. No language in the specification should be construed as indicating any non-claimed element essential to the practice of the invention.

[0091] Groupings of alternative elements or embodiments of the invention disclosed herein are not to be construed as limitations. Each group member may be referred to and claimed individually or in any combination with other members of the group or other elements found herein. It is anticipated that one or more members of a group may be included in, or deleted from, a group for reasons of convenience and/or patentability. When any such inclusion or deletion occurs, the specification is deemed to contain the group as modified thus fulfilling the written description of all Markush groups used in the appended claims.

[0092] Certain embodiments of this invention are described herein, including the best mode known to the inventors for carrying out the invention. Of course, variations on these described embodiments will become apparent to those of ordinary skill in the art upon reading the foregoing description. The inventor expects skilled artisans to employ such variations as appropriate, and the inventors intend for the invention to be practiced otherwise than specifically described herein. Accordingly, this invention includes all modifications and equivalents of the subject

matter recited in the claims appended hereto as permitted by applicable law. Moreover, any combination of the abovedescribed elements in all possible variations thereof is encompassed by the invention unless otherwise indicated herein or otherwise clearly contradicted by context.

[0093] Specific embodiments disclosed herein may be further limited in the claims using consisting of or consisting essentially of language. When used in the claims, whether as filed or added per amendment, the transition term "consisting of" excludes any element, step, or ingredient not specified in the claims. The transition term "consisting essentially of" limits the scope of a claim to the specified materials or steps and those that do not materially affect the basic and novel characteristic(s). Embodiments of the invention so claimed are inherently or expressly described and enabled herein.

[0094] Furthermore, numerous references have been made to patents and printed publications throughout this specification. Each of the above-cited references and printed publications are individually incorporated herein by reference in their entirety.

[0095] In closing, it is to be understood that the embodiments of the invention disclosed herein are illustrative of the principles of the present invention. Other modifications that may be employed are within the scope of the invention. Thus, by way of example, but not of limitation, alternative configurations of the present invention may be utilized in accordance with the teachings herein. Accordingly, the present invention is not limited to that precisely as shown and described.

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Pro	Thr	Gln	Gly	Glu 245	Ala	Tyr	Leu	Asp	Lys 250	Gln	Ser	Asp	Thr	Gln 255	Tyr
Val	Сүз	Lys	Arg 260	Thr	Leu	Val	Asp	Arg 265	Gly	Trp	Gly	Asn	Gly 270	Суз	Gly
Leu	Phe	Gly 275		Gly	Ser	Leu	Val 280	Thr	Суз	Ala	Lys	Phe 285	Ala	Суз	Ser
ГÀа	Lys 290	Met	Thr	Gly	ГÀа	Ser 295	Ile	Gln	Pro	Glu	Asn 300	Leu	Glu	Tyr	Arg
Ile 305	Met	Leu	Ser	Val	His 310	Gly	Ser	Gln	His	Ser 315	Gly	Met	Ile	Val	Asn 320
Asp	Thr	Gly	His	Glu 325	Thr	Asp	Glu	Asn	Arg 330	Ala	ГÀа	Val	Glu	Ile 335	Thr
Pro	Asn	Ser	Pro 340	Arg	Ala	Glu	Ala	Thr 345	Leu	Gly	Gly	Phe	Gly 350	Ser	Leu
Gly	Leu	Asp 355		Glu	Pro	Arg	Thr 360	Gly	Leu	Asp	Phe	Ser 365	Asp	Leu	Tyr
Tyr	Leu 370	Thr	Met	Asn	Asn	Lys 375	His	Trp	Leu	Val	His 380	Lys	Glu	Trp	Phe
His 385	Asp	Ile	Pro	Leu	Pro 390	Trp	His	Ala	Gly	Ala 395	Asp	Thr	Gly	Thr	Pro 400
His	Trp	Asn	Asn	Lys 405		Ala	Leu	Val	Glu 410	Phe	Lys	Asp	Ala	His 415	Ala
Lys	Arg	Gln	Thr 420	Val	Val	Val	Leu	Gly 425	Ser	Gln	Glu	Gly	Ala 430	Val	His
Thr	Ala	Leu 435	Ala	Gly	Ala	Leu	Glu 440	Ala	Glu	Met	Asp	Gly 445	Ala	Гла	Gly
Arg	Leu 450	Ser	Ser	Gly	His	Leu 455	Lys	Суз	Arg	Leu	Lys 460	Met	Asp	Lys	Leu
Arg 465	Leu	Lys	Gly	Val	Ser 470	Tyr	Ser	Leu	Суа	Thr 475	Ala	Ala	Phe	Thr	Phe 480
	Lys	Ile	Pro	Ala 485		Thr	Leu	His	Gly 490		Val	Thr	Val	Glu 495	
Gln	Tyr	Ala	-		Asp	Gly	Pro	-		Val	Pro	Ala			Ala
Val	Asp		500 Gln	Thr	Leu	Thr		505 Val	Gly	Arg	Leu		510 Thr	Ala	Asn
Pro	Val	515 Ile	Thr	Glu	Ser	Thr	520 Glu	Asn	Ser	Lys	Met	525 Met	Leu	Glu	Leu
Asp	530 Pro	Pro	Phe	Glv	Asp	535 Ser	Tvr	Ile	Val	Ile	540 Glv	Val	Glv	Glu	Lvs
545	FIO	110	rne	Gry	550	Der	TÄT	116	Var	555	Gry	Vai	Gry	Gru	560

Lys Ile Thr His His Trp His Arg Ser Gly Ser Thr Ile Gly Lys Ala Phe Glu Ala Thr Val Arg Gly Ala Lys Arg Met Ala Val Leu Gly Asp Thr Ala Trp Asp Phe Gly Ser Val Gly Gly Ala Leu Asn Ser Leu Gly Lys Gly Ile His Gln Ile Phe Gly Ala Ala Phe Lys Ser Leu Phe Gly Gly Met Ser Trp Phe Ser Gln Ile Leu Ile Gly Thr Leu Leu Met Trp625630635640 Leu Gly Leu Asn Thr Lys Asn Gly Ser Ile Ser Leu Met Cys Leu Ala Leu Gly Gly Val Leu Ile Phe Leu Ser Thr Ala Val Ser Ala Asp Val Gly Cys Ser Val <210> SEO ID NO 2 <211> LENGTH: 505 <212> TYPE: PRT <213> ORGANISM: Zika Virus <400> SEOUENCE: 2 Ile Arg Cys Ile Gly Val Ser Asn Arg Asp Phe Val Glu Gly Met Ser Gly Gly Thr Trp Val Asp Ile Val Leu Glu His Gly Gly Cys Val Thr Val Met Ala Gln Asp Lys Pro Thr Val Asp Ile Glu Leu Val Thr Thr Thr Val Ser Asn Met Ala Glu Val Arg Ser Tyr Cys Tyr Glu Ala Ser Ile Ser Asp Met Ala Ser Asp Ser Arg Cys Pro Thr Gln Gly Glu Ala Tyr Leu Asp Lys Gln Ser Asp Thr Gln Tyr Val Cys Lys Arg Thr Leu Val Asp Arg Gly Trp Gly Asn Gly Cys Gly Leu Phe Gly Lys Gly Ser Leu Val Thr Cys Ala Lys Phe Ala Cys Ser Lys Lys Met Thr Gly Lys Ser Ile Gln Pro Glu Asn Leu Glu Tyr Arg Ile Met Leu Ser Val His Gly Ser Gln His Ser Gly Met Ile Val Asn Asp Thr Gly His Glu Thr Asp Glu Asn Arg Ala Lys Val Glu Ile Thr Pro Asn Ser Pro Arg Ala Glu Ala Thr Leu Gly Gly Phe Gly Ser Leu Gly Leu Asp Cys Glu Pro Arg Thr Gly Leu Asp Phe Ser Asp Leu Tyr Tyr Leu Thr Met Asn Asn Lys His Trp Leu Val His Lys Glu Trp Phe His Asp Ile Pro Leu Pro Trp His Ala Gly Ala Asp Thr Gly Thr Pro His Trp Asn Asn Lys Glu

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225					230					235					240
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Leu	Glu	Ala 275	Glu	Met	Asp	Gly	Ala 280	Lys	Gly	Arg	Leu	Ser 285	Ser	Gly	His
Leu	Lys 290	Cys	Arg	Leu	Lys	Met 295	Asp	Lys	Leu	Arg	Leu 300	Lys	Gly	Val	Ser
Tyr 305	Ser	Leu	Суз	Thr	Ala 310	Ala	Phe	Thr	Phe	Thr 315	Lys	Ile	Pro	Ala	Glu 320
Thr	Leu	His	Gly	Thr 325	Val	Thr	Val	Glu	Val 330	Gln	Tyr	Ala	Gly	Thr 335	Asp
Gly	Pro	Cys	Lys 340	Val	Pro	Ala	Gln	Met 345	Ala	Val	Asp	Met	Gln 350	Thr	Leu
Thr	Pro	Val 355	Gly	Arg	Leu	Ile	Thr 360	Ala	Asn	Pro	Val	Ile 365	Thr	Glu	Ser
Thr	Glu 370	Asn	Ser	ГЛа	Met	Met 375	Leu	Glu	Leu	Asp	Pro 380	Pro	Phe	Gly	Asp
Ser 385	Tyr	Ile	Val	Ile	Gly 390	Val	Gly	Glu	Lys	Lys 395	Ile	Thr	His	His	Trp 400
His	Arg	Ser	Gly	Ser 405	Thr	Ile	Gly	Lys	Ala 410	Phe	Glu	Ala	Thr	Val 415	Arg
Gly	Ala	Lys	Arg 420	Met	Ala	Val	Leu	Gly 425	Asp	Thr	Ala	Trp	Asp 430	Phe	Gly
Ser	Val	Gly 435	Gly	Ala	Leu	Asn	Ser 440	Leu	Gly	Lys	Gly	Ile 445	His	Gln	Ile
Phe	Gly 450	Ala	Ala	Phe	Lys	Ser 455	Leu	Phe	Gly	Gly	Met 460	Ser	Trp	Phe	Ser
Gln 465	Ile	Leu	Ile	Gly	Thr 470	Leu	Leu	Met	Trp	Leu 475	Gly	Leu	Asn	Thr	Lys 480
Asn	Gly	Ser	Ile	Ser 485	Leu	Met	Сүз	Leu	Ala 490	Leu	Gly	Gly	Val	Leu 495	Ile
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	0> SH	-			17 - 7	G -	m	G -	т -	<u></u>	m1-	7 7		D ¹ -	m¹-
Leu 1	Arg	ьеч	гла	GIY 5	vai	ser	TYr	ser	Leu 10	сув	Tnr	АІА	AIA	Phe 15	Tnr
Phe	Thr	Lys	Ile 20	Pro	Ala	Glu	Thr	Leu 25	His	Gly	Thr	Val	Thr 30	Val	Glu
Val	Gln	Tyr 35	Ala	Gly	Thr	Asp	Gly 40	Pro	Суз	ГЛа	Val	Pro 45	Ala	Gln	Met
Ala	Val 50	Asp	Met	Gln	Thr	Leu 55	Thr	Pro	Val	Gly	Arg 60	Leu	Ile	Thr	Ala
Asn 65	Pro	Val	Ile	Thr	Glu 70	Ser	Thr	Glu	Asn	Ser 75	ГЛЗ	Met	Met	Leu	Glu 80

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Leu Asp Pro Pro Phe Gly Asp Ser Tyr Ile Val Ile Gly Val Gly Glu Lys Lys Ile Thr His His Trp His Arg Ser Gly Ser Thr Ile Gly Lys <210> SEQ ID NO 4 <211> LENGTH: 112 <212> TYPE: PRT <213> ORGANISM: Artificial Sequence <220> FEATURE: <223> OTHER INFORMATION: ZIKV EDIII M375N/E377T protein <400> SEQUENCE: 4 Leu Arg Leu Lys Gly Val Ser Tyr Ser Leu Cys Thr Ala Ala Phe Thr Phe Thr Lys Ile Pro Ala Glu Thr Leu His Gly Thr Val Thr Val Glu Val Gln Tyr Ala Gly Thr Asp Gly Pro Cys Lys Val Pro Ala Gln Met Ala Val Asp Met Gln Thr Leu Thr Pro Val Gly Arg Leu Ile Thr Ala Asn Pro Val Ile Thr Glu Ser Thr Glu Asn Ser Lys Met Asn Leu Thr Leu Asp Pro Pro Phe Gly Asp Ser Tyr Ile Val Ile Gly Val Gly Glu Lys Lys Ile Thr His His Trp His Arg Ser Gly Ser Thr Ile Gly Lys <210> SEQ ID NO 5 <211> LENGTH: 341 <212> TYPE: PRT <213> ORGANISM: Artificial Sequence <220> FEATURE: <223> OTHER INFORMATION: ZIKV EDIII WT-Fc protein <400> SEQUENCE: 5 Leu Arg Leu Lys Gly Val Ser Tyr Ser Leu Cys Thr Ala Ala Phe Thr Phe Thr Lys Ile Pro Ala Glu Thr Leu His Gly Thr Val Thr Val Glu Val Gln Tyr Ala Gly Thr Asp Gly Pro Cys Lys Val Pro Ala Gln Met Ala Val Asp Met Gln Thr Leu Thr Pro Val Gly Arg Leu Ile Thr Ala Asn Pro Val Ile Thr Glu Ser Thr Glu Asn Ser Lys Met Met Leu Glu Leu Asp Pro Pro Phe Gly Asp Ser Tyr Ile Val Ile Gly Val Gly Glu Lys Lys Ile Thr His His Trp His Arg Ser Gly Ser Thr Ile Gly Lys Arg Ser Asp Lys Thr His Thr Cys Pro Pro Cys Pro Ala Pro Glu Leu Leu Gly Gly Pro Ser Val Phe Leu Phe Pro Pro Lys Pro Lys Asp Thr Leu Met Ile Ser Arg Thr Pro Glu Val Thr Cys Val Val Val Asp Val Ser His Glu Asp Pro Glu Val Lys Phe Asn Trp Tyr Val Asp Gly Val Glu Val His Asn Ala Lys Thr Lys Pro Arg Glu Glu Gln Tyr Asn Ser Thr Tyr Arg Val Val Ser Val Leu Thr Val Leu His Gln Asp Trp Leu Asn Gly Lys Glu Tyr Lys Cys Lys Val Ser Asn Lys Ala Leu Pro Ala Pro Ile Glu Lys Thr Ile Ser Lys Ala Lys Gly Gln Pro Arg Glu Pro225230235240 Gln Val Tyr Thr Leu Pro Pro Ser Arg Glu Glu Met Thr Lys Asn Gln Val Ser Leu Thr Cys Leu Val Lys Gly Phe Tyr Pro Ser Asp Ile Ala 260 265 Val Glu Trp Glu Ser Asn Gly Gln Pro Glu Asn Asn Tyr Lys Thr Thr Pro Pro Val Leu Asp Ser Asp Gly Ser Phe Phe Leu Tyr Ser Lys Leu Thr Val Asp Lys Ser Arg Trp Gln Gln Gly Asn Val Phe Ser Cys Ser Val Met His Glu Ala Leu His Asn His Tyr Thr Gln Lys Ser Leu Ser Leu Ser Pro Gly Lys <210> SEQ ID NO 6 <211> LENGTH: 341 <212> TYPE: PRT <213> ORGANISM: Artificial Sequence <220> FEATURE: <223> OTHER INFORMATION: ZIKV EDIII M375N/E377T-Fc protein <400> SEQUENCE: 6 Leu Arg Leu Lys Gly Val Ser Tyr Ser Leu Cys Thr Ala Ala Phe Thr Phe Thr Lys Ile Pro Ala Glu Thr Leu His Gly Thr Val Thr Val Glu Val Gln Tyr Ala Gly Thr Asp Gly Pro Cys Lys Val Pro Ala Gln Met Ala Val Asp Met Gln Thr Leu Thr Pro Val Gly Arg Leu Ile Thr Ala Asn Pro Val Ile Thr Glu Ser Thr Glu Asn Ser Lys Met Asn Leu Thr Leu Asp Pro Pro Phe Gly Asp Ser Tyr Ile Val Ile Gly Val Gly Glu Lys Lys Ile Thr His His Trp His Arg Ser Gly Ser Thr Ile Gly Lys Arg Ser Asp Lys Thr His Thr Cys Pro Pro Cys Pro Ala Pro Glu Leu Leu Gly Gly Pro Ser Val Phe Leu Phe Pro Pro Lys Pro Lys Asp Thr Leu Met Ile Ser Arg Thr Pro Glu Val Thr Cys Val Val Val Asp Val

<212> TYPE: PRT

<213> ORGANISM: Artificial Sequence

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Ser His Glu Asp Pro Glu Val Lys Phe Asn Trp Tyr Val Asp Gly Val Glu Val His Asn Ala Lys Thr Lys Pro Arg Glu Glu Gln Tyr Asn Ser Thr Tyr Arg Val Val Ser Val Leu Thr Val Leu His Gln Asp Trp Leu Asn Gly Lys Glu Tyr Lys Cys Lys Val Ser Asn Lys Ala Leu Pro Ala Pro Ile Glu Lys Thr Ile Ser Lys Ala Lys Gly Gln Pro Arg Glu Pro225230235240 Gln Val Tyr Thr Leu Pro Pro Ser Arg Glu Glu Met Thr Lys Asn Gln Val Ser Leu Thr Cys Leu Val Lys Gly Phe Tyr Pro Ser Asp Ile Ala 260 265 Val Glu Trp Glu Ser Asn Gly Gln Pro Glu Asn Asn Tyr Lys Thr Thr Pro Pro Val Leu Asp Ser Asp Gly Ser Phe Phe Leu Tyr Ser Lys Leu Thr Val Asp Lys Ser Arg Trp Gln Gln Gly Asn Val Phe Ser Cys Ser Val Met His Glu Ala Leu His Asn His Tyr Thr Gln Lys Ser Leu Ser Leu Ser Pro Gly Lys <210> SEQ ID NO 7 <211> LENGTH: 118 <212> TYPE: PRT <213> ORGANISM: Artificial Sequence <220> FEATURE: <223> OTHER INFORMATION: ZIKV EDIII WT-His protein <400> SEQUENCE: 7 Leu Arg Leu Lys Gly Val Ser Tyr Ser Leu Cys Thr Ala Ala Phe Thr Phe Thr Lys Ile Pro Ala Glu Thr Leu His Gly Thr Val Thr Val Glu Val Gln Tyr Ala Gly Thr Asp Gly Pro Cys Lys Val Pro Ala Gln Met Ala Val Asp Met Gln Thr Leu Thr Pro Val Gly Arg Leu Ile Thr Ala Asn Pro Val Ile Thr Glu Ser Thr Glu Asn Ser Lys Met Met Leu Glu Leu Asp Pro Pro Phe Gly Asp Ser Tyr Ile Val Ile Gly Val Gly Glu Lys Lys Ile Thr His His Trp His Arg Ser Gly Ser Thr Ile Gly Lys His His His His His His <210> SEQ ID NO 8 <211> LENGTH: 118

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Val Gln Tyr Ala Gly Thr Asp Gly Pro Cys Lys Val Pro Ala Gln Met Ala Val Asp Met Gln Thr Leu Thr Pro Val Gly Arg Leu Ile Thr Ala Asn Pro Val Ile Asn Glu Thr Thr Glu Asn Ser Lys Met Met Leu Glu Leu Asp Pro Pro Phe Gly Asp Ser Tyr Ile Val Ile Gly Val Gly Glu Lys Lys Ile Thr His His Trp His Arg Ser Gly Ser Thr Ile Gly Lys <210> SEQ ID NO 11 <211> LENGTH: 341 <212> TYPE: PRT <213> ORGANISM: Artificial Sequence <220> FEATURE: <223> OTHER INFORMATION: ZIKV EDIII A333N-Fc protein <400> SEOUENCE: 11 Leu Arg Leu Lys Gly Val Ser Tyr Ser Leu Cys Thr Ala Ala Phe Thr Phe Thr Lys Ile Pro Ala Glu Thr Leu His Gly Thr Val Thr Val Glu Val Gln Tyr Asn Gly Thr Asp Gly Pro Cys Lys Val Pro Ala Gln Met Ala Val Asp Met Gln Thr Leu Thr Pro Val Gly Arg Leu Ile Thr Ala Asn Pro Val Ile Thr Glu Ser Thr Glu Asn Ser Lys Met Met Leu Glu Leu Asp Pro Pro Phe Gly Asp Ser Tyr Ile Val Ile Gly Val Gly Glu Lys Lys Ile Thr His His Trp His Arg Ser Gly Ser Thr Ile Gly Lys Arg Ser Asp Lys Thr His Thr Cys Pro Pro Cys Pro Ala Pro Glu Leu Leu Gly Gly Pro Ser Val Phe Leu Phe Pro Pro Lys Pro Lys Asp Thr Leu Met Ile Ser Arg Thr Pro Glu Val Thr Cys Val Val Val Asp Val Ser His Glu Asp Pro Glu Val Lys Phe Asn Trp Tyr Val Asp Gly Val Glu Val His Asn Ala Lys Thr Lys Pro Arg Glu Glu Gln Tyr Asn Ser Thr Tyr Arg Val Val Ser Val Leu Thr Val Leu His Gln Asp Trp Leu Asn Gly Lys Glu Tyr Lys Cys Lys Val Ser Asn Lys Ala Leu Pro Ala Pro Ile Glu Lys Thr Ile Ser Lys Ala Lys Gly Gln Pro Arg Glu Pro Gln Val Tyr Thr Leu Pro Pro Ser Arg Glu Glu Met Thr Lys Asn Gln Val Ser Leu Thr Cys Leu Val Lys Gly Phe Tyr Pro Ser Asp Ile Ala

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Val Glu Trp Glu Ser Asn Gly Gln Pro Glu Asn Asn Tyr Lys Thr Thr Pro Pro Val Leu Asp Ser Asp Gly Ser Phe Phe Leu Tyr Ser Lys Leu Thr Val Asp Lys Ser Arg Trp Gln Gln Gly Asn Val Phe Ser Cys Ser Val Met His Glu Ala Leu His Asn His Tyr Thr Gln Lys Ser Leu Ser Leu Ser Pro Gly Lys <210> SEQ ID NO 12 <211> LENGTH: 341 <212> TYPE: PRT <213> ORGANISM: Artificial Sequence <220> FEATURE: <223> OTHER INFORMATION: ZIKV EDIII T366N/S368T-Fc protein <400> SEOUENCE: 12 Leu Arg Leu Lys Gly Val Ser Tyr Ser Leu Cys Thr Ala Ala Phe Thr Phe Thr Lys Ile Pro Ala Glu Thr Leu His Gly Thr Val Thr Val Glu Val Gln Tyr Ala Gly Thr Asp Gly Pro Cys Lys Val Pro Ala Gln Met Ala Val Asp Met Gln Thr Leu Thr Pro Val Gly Arg Leu Ile Thr Ala Asn Pro Val Ile Asn Glu Thr Thr Glu Asn Ser Lys Met Met Leu Glu Leu Asp Pro Pro Phe Gly Asp Ser Tyr Ile Val Ile Gly Val Gly Glu Lys Lys Ile Thr His His Trp His Arg Ser Gly Ser Thr Ile Gly Lys Arg Ser Asp Lys Thr His Thr Cys Pro Pro Cys Pro Ala Pro Glu Leu Leu Gly Gly Pro Ser Val Phe Leu Phe Pro Pro Lys Pro Lys Asp Thr Leu Met Ile Ser Arg Thr Pro Glu Val Thr Cys Val Val Val Asp Val Ser His Glu Asp Pro Glu Val Lys Phe Asn Trp Tyr Val Asp Gly Val Glu Val His Asn Ala Lys Thr Lys Pro Arg Glu Glu Gln Tyr Asn Ser Thr Tyr Arg Val Val Ser Val Leu Thr Val Leu His Gln Asp Trp Leu Asn Gly Lys Glu Tyr Lys Cys Lys Val Ser Asn Lys Ala Leu Pro Ala Pro Ile Glu Lys Thr Ile Ser Lys Ala Lys Gly Gln Pro Arg Glu Pro Gln Val Tyr Thr Leu Pro Pro Ser Arg Glu Glu Met Thr Lys Asn Gln Val Ser Leu Thr Cys Leu Val Lys Gly Phe Tyr Pro Ser Asp Ile Ala

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Val Glu Trp Glu Ser Asn Gly Gln Pro Glu Asn Asn Tyr Lys Thr Thr Pro Pro Val Leu Asp Ser Asp Gly Ser Phe Phe Leu Tyr Ser Lys Leu Thr Val Asp Lys Ser Arg Trp Gln Gln Gly Asn Val Phe Ser Cys Ser Val Met His Glu Ala Leu His Asn His Tyr Thr Gln Lys Ser Leu Ser Leu Ser Pro Gly Lys <210> SEQ ID NO 13 <211> LENGTH: 112 <212> TYPE: PRT <213> ORGANISM: Artificial Sequence <220> FEATURE: <223> OTHER INFORMATION: ZIKV EDIII T309N/A311T protein <400> SEOUENCE: 13 Leu Arg Leu Lys Gly Val Ser Tyr Ser Leu Cys Asn Ala Thr Phe Thr Phe Thr Lys Ile Pro Ala Glu Thr Leu His Gly Thr Val Thr Val Glu Val Gln Tyr Ala Gly Thr Asp Gly Pro Cys Lys Val Pro Ala Gln Met Ala Val Asp Met Gln Thr Leu Thr Pro Val Gly Arg Leu Ile Thr Ala Asn Pro Val Ile Thr Glu Ser Thr Glu Asn Ser Lys Met Met Leu Glu Leu Asp Pro Pro Phe Gly Asp Ser Tyr Ile Val Ile Gly Val Gly Glu Lys Lys Ile Thr His His Trp His Arg Ser Gly Ser Thr Ile Gly Lys <210> SEQ ID NO 14 <211> LENGTH: 112 <212> TYPE: PRT <213> ORGANISM: Artificial Sequence <220> FEATURE: <223> OTHER INFORMATION: ZIKV EDIII T315N/I317T protein <400> SEQUENCE: 14 Leu Arg Leu Lys Gly Val Ser Tyr Ser Leu Cys Thr Ala Ala Phe Thr Phe Asn Lys Thr Pro Ala Glu Thr Leu His Gly Thr Val Thr Val Glu Val Gln Tyr Ala Gly Thr Asp Gly Pro Cys Lys Val Pro Ala Gln Met Ala Val Asp Met Gln Thr Leu Thr Pro Val Gly Arg Leu Ile Thr Ala Asn Pro Val Ile Thr Glu Ser Thr Glu Asn Ser Lys Met Met Leu Glu Leu Asp Pro Pro Phe Gly Asp Ser Tyr Ile Val Ile Gly Val Gly Glu Lys Lys Ile Thr His His Trp His Arg Ser Gly Ser Thr Ile Gly Lys

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Val	Gln	Tyr 35	Ala	Gly	Thr	Asp	Gly 40	Pro	Суз	Lys	Val	Pro 45	Ala	Gln	Met
Ala	Val 50	Asp	Met	Gln	Thr	Leu 55	Thr	Pro	Val	Gly	Arg 60	Leu	Ile	Thr	Ala
Asn 65	Pro	Val	Ile	Thr	Glu 70	Ser	Thr	Glu	Asn	Ser 75	Lys	Met	Met	Leu	Glu 80
Leu	Asp	Pro	Pro	Phe 85	Gly	Asp	Ser	Tyr	Ile 90	Val	Ile	Gly	Val	Gly 95	Asn
Lys	Thr	Ile	Thr 100	His	His	Trp	His	Arg 105	Ser	Gly	Ser	Thr	Ile 110	Gly	Lys
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Phe	Thr	Гла	Ile 20	Pro	Ala	Glu	Thr	Leu 25	His	Gly	Thr	Val	Thr 30	Val	Glu
Val	Gln	Tyr 35	Ala	Gly	Thr	Asp	Gly 40	Pro	Суз	Lys	Val	Pro 45	Ala	Gln	Met
Ala	Val 50	Asp	Met	Gln	Thr	Leu 55	Thr	Pro	Val	Gly	Arg 60	Leu	Ile	Thr	Ala
Asn 65	Pro	Val	Ile	Thr	Glu 70	Ser	Thr	Glu	Asn	Ser 75	Lys	Met	Met	Leu	Glu 80
Leu	Asp	Pro	Pro	Phe 85	Gly	Asp	Ser	Tyr	Ile 90	Val	Ile	Gly	Val	Gly 95	Glu
Lys	Lys	Ile	Thr 100	His	His	Trp	His	Arg 105	Ser	Gly	Ser	Thr	Ile 110	Gly	Lys
Arg	Ser	Asp 115	Lys	Thr	His	Thr	Cys 120	Pro	Pro	CÀa	Pro	Ala 125	Pro	Glu	Leu
Leu	Gly 130	Gly	Pro	Ser	Val	Phe 135	Leu	Phe	Pro	Pro	Lys 140	Pro	Lys	Asp	Thr
Leu 145	Met	Ile	Ser	Arg	Thr 150	Pro	Glu	Val	Thr	Cys 155	Val	Val	Val	Asp	Val 160
Ser	His	Glu	Asp	Pro 165	Glu	Val	Lys	Phe	Asn 170	Trp	Tyr	Val	Asp	Gly 175	Val
Glu	Val	His	Asn 180	Ala	Lys	Thr	Lys	Pro 185	Arg	Glu	Glu	Gln	Tyr 190	Asn	Ser
Thr	Tyr	Arg 195	Val	Val	Ser	Val	Leu 200	Thr	Val	Leu	His	Gln 205	Asp	Trp	Leu
Asn	Gly 210		Glu	Tyr	ГЛа	Cys 215	Lys	Val	Ser	Asn	Lys 220	Ala	Leu	Pro	Ala
Pro 225	Ile	Glu	Lys	Thr	Ile 230	Ser	Lys	Ala	Lys	Gly 235		Pro	Arg	Glu	Pro 240
Gln	Val	Tyr	Thr	Leu 245	Pro	Pro	Ser	Arg	Glu 250	Glu	Met	Thr	Lys	Asn 255	Gln

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Val	Ser	Leu	Thr 260	Сув	Leu	Val	Lys	Gly 265	Phe	Tyr	Pro	Ser	Asp 270	Ile	Ala
Val	Glu	Trp 275	Glu	Ser	Asn	Gly	Gln 280	Pro	Glu	Asn	Asn	Tyr 285	Lys	Thr	Thr
Pro	Pro 290	Val	Leu	Asp	Ser	Asp 295		Ser	Phe	Phe	Leu 300	Tyr	Ser	Lys	Leu
Thr 305	Val	Aap	Lys	Ser	Arg 310	Trp	Gln	Gln	Gly	Asn 315	Val	Phe	Ser	Сүз	Ser 320
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Leu	Ser	Pro	Gly 340	Lys											
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1. A method for preventing and/or treating a Zika viral infection in a subject in need thereof, the method comprising administering a therapeutically effective amount of an isolated polypeptide comprising an altered structural envelope (E) protein domain III (EDIII), wherein the alteration reduces or prevents the immunogenicity of one or more interfering epitopes of the EDW.

2. The method of claim 1 further comprising co-administration of an adjuvant.

3. The method of claim **1**, wherein the interfering epitope surrounds residue 333, 366, or 375 of ZIKV EDIII polypetide.

4. The method of claim **1**, wherein the alteration comprises creation of a site for N-linked glycosylation.

5. (canceled)

6. The method of claim **4**, wherein the alteration comprises at least an N at residue 375 and either a T or S at residue 377.

7. The method of claim 4, wherein the alteration comprises at least an N at residue 333 and either a T or S at residue 335.

8. The method of claim **4**, wherein the alteration comprises at least an N at residue 366 and either a T or S at residue 368.

9. The method of claim **6**, wherein the alteration comprises M375N/E377T.

10. The method of claim 7, wherein the alteration comprises A333N.

11. The method of claim 8, wherein the alteration comprises T366N/S368T.

12. The method of claim **1**, wherein the administration decreases viral titer levels, or viral RNA copy number.

13. (canceled)

14. The method of claim **1**, wherein the administration increases production of neutralizing antibodies.

15. (canceled)

16. (canceled)

17. (canceled)

19. An immunogenic composition comprising an isolated polypeptide comprising any of SEQ ID NOs 4-6 or 8-22, or a combination thereof.

20. (canceled)

21. The immunogenic composition of claim **19**, further comprising an adjuvant.

22. A method for preventing and/or treating a Zika virus infection in a subject in need thereof, the method comprising administering a therapeutically effective amount of an immunogenic composition of claim 19 to a subject in need thereof.

23. The method of claim **22**, wherein the subject is a woman who is pregnant, who may become pregnant, or who plans to become pregnant.

24. The method of claim **22**, wherein the administration increases production of neutralizing antibodies.

25. The method of claim **22**, wherein the administration decreases viral titer levels, or viral RNA copy number.

26. (canceled)

27. The method of claim 23, wherein as a result of the administration, any pregnancy in the woman does not result in Zika virus-associated birth defects.

28. (canceled)

29. (canceled)

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